Overview
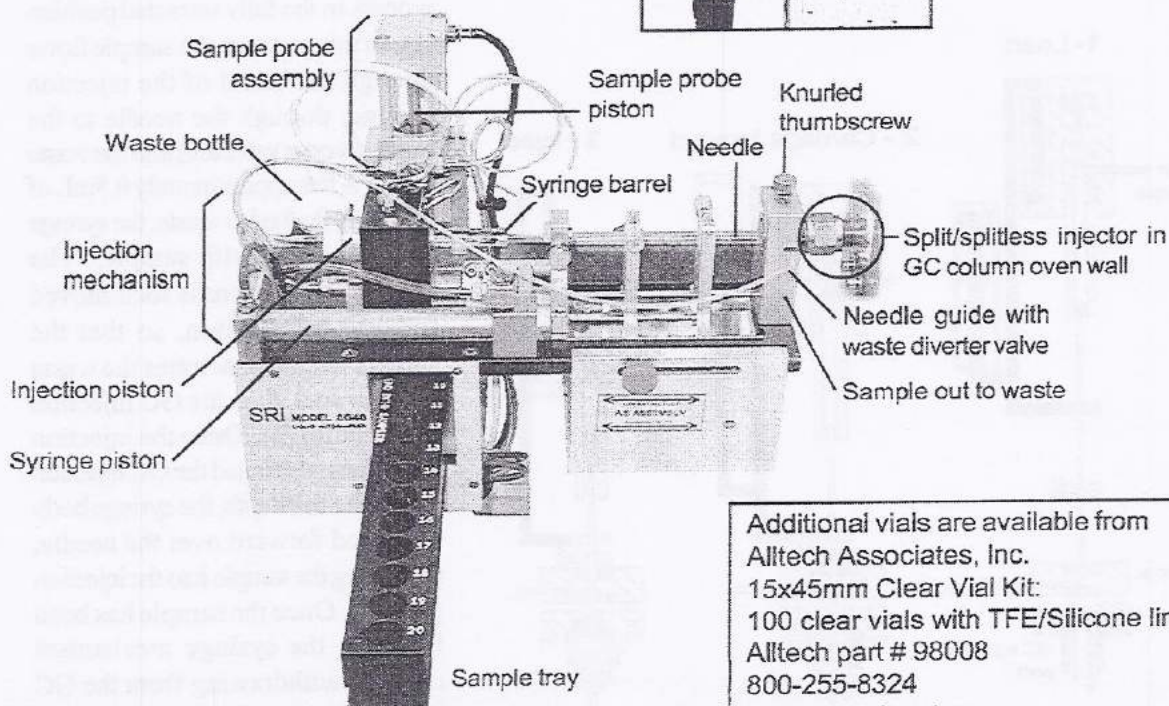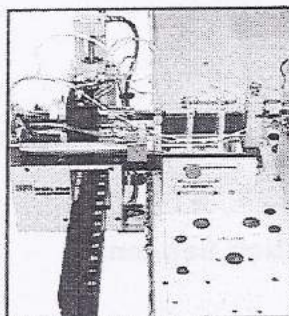
The SRI Model 8640 20-vial liquid autosampler is installed on the left-hand side of the SRI 8610C GC. The 8640 connects to an additional injector on the left side of the column oven. This additional injector may be on-column, heated, or split/splitless. It uses a sample tray to hold up to twenty 2mL vials, a sample probe to transfer the sample from the vials into the syringe barrel, and an injection mechanism to deliver the sample from the syringe barrel, through the needle, into the injector in the GC column oven wall. The 8640 uses 60psi of air or nitrogen to actuate its moving parts. The 8640 functions are assigned relays so that the autosampler may be operated automatically using a PeakSimple event table.

The 8640 is shipped with 100 screw-top vials and septa, replacements for which are available from a variety of suppliers. Extra sample trays and cooled sample trays are available. The cooled sample trays require an external refrigerated lab circulator.
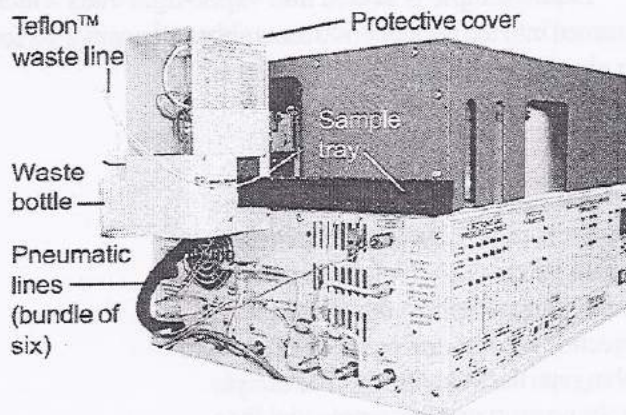
8640 with the protective cover removed

Additional vials are available from
Alltech Associates, Inc.
15x45mm Clear Vial Kit:
100 clear vials with TFE/Silicone liners
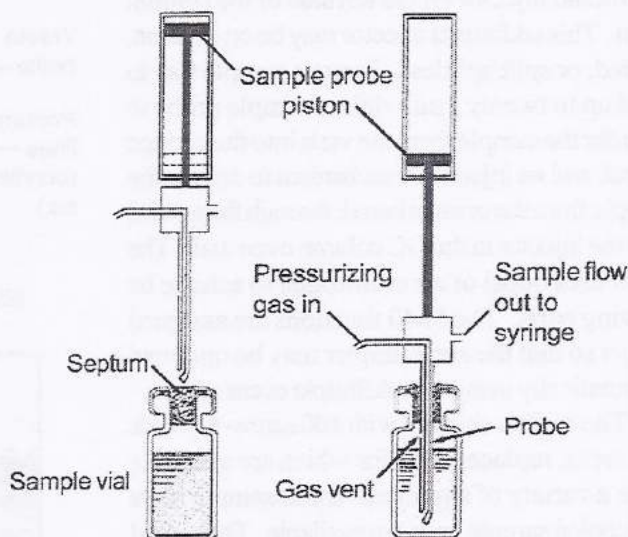Alltech part # 98008
800-255-8324
www.alltechweb.com
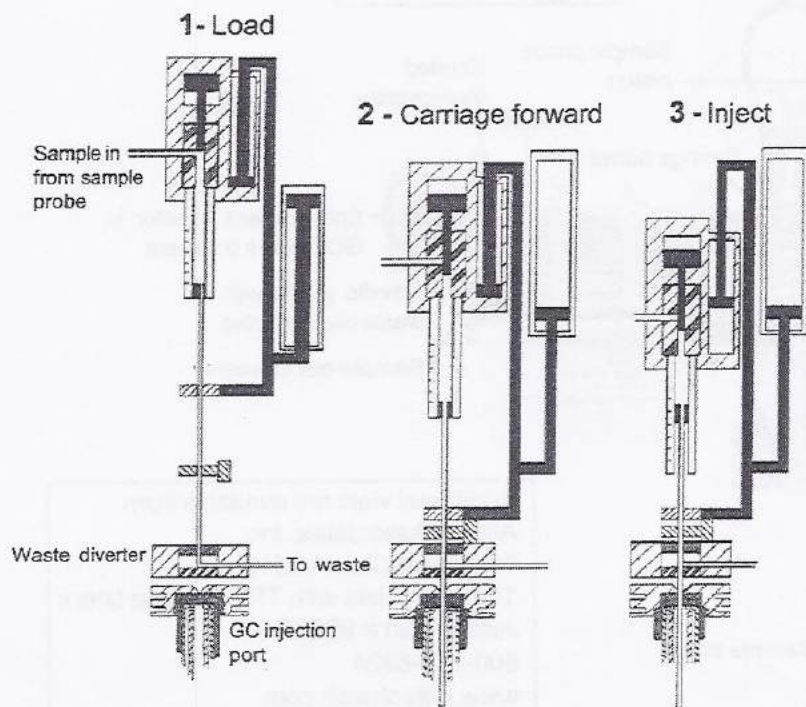
Theory of Operation

Liquid sample is sealed into vapor-tight vials which are inserted into the sample tray. The tray is then inserted into the Model 8640 assembly and positioned for the sampling sequence. The 8640 uses 60psi of air or nitrogen to actuate its moving parts.

The sampling probe, a concentric needle, is moved down by a piston to pressurize the vial with helium or other gas, causing sample to flow through the injection syringe and out the Teflon™ tubing into the waste bottle. The sample probe pressurizes the sample vial for a period of time long enough to rinse the previous sample to waste and fill the syringe with sample.

Sampling Probe Sequence

Sample probe piston
Pressurizing gas in
Sample flow out to syringe
Septum
Sample vial
Probe
Gas vent

Injection Sequence

1 - Load

Sample in from sample probe

2 - Carriage forward

3 - Inject

Waste diverter — To waste
GC injection port

The syringe mechanism begins the process in the fully retracted position (1). In this position, the sample flows through the barrel of the injection syringe, through the needle to the waste diverter valve and into the waste bottle. After approximately 0.5mL of sample is flushed to waste, the syringe barrel is filled with sample. The syringe mechanism is then moved forward by a piston, so that the syringe needle penetrates the waste diverter seal, then the GC injection port septum (2). Once the injection needle has penetrated the GC injection port to the full depth, the syringe body is pushed forward over the needle, displacing the sample into the injection port (3). Once the sample has been injected, the syringe mechanism retracts, withdrawing from the GC injection port.

General Operating Procedures

1. Fill each 2mL vial at least 75% full with liquid sample. Close the vials so that they are vapor-tight, with the Teflon side of the vial septa facing downward into the vial.

2. The sample tray is inserted and removed from the 8640 in one direction only. To remove the sample vial tray, push it away from you, toward the back of the GC, until it is free of the autosampler assembly. Place the filled sample vials in the tray. Reinsert the sample tray into the 8640 assembly from the front. Push it gently toward the back of the GC until the white lines at the tip of the white arrow on the sample tray are aligned with the front edge of the 8640. The sample tray is then in the ready position, with vial number one in place under the sample probe. The sample tray shown below, right is almost in the ready position (it was left partially out for visibility of the lines and arrow).

3. Activate and heat the GC detector(s).

4. Load or create a column oven temperature program.

5. Load or create an event table. Version 2.74 (and higher) of the PeakSimple software includes an event table file called "8640as.evt" as a general event table for use with the 8640 autosampler. When you load this event file, the default relay descriptions will not match the actual 8640 autosampler relay descriptions. These autosampler-specific descriptions must be entered by you, the user. The relays assigned to the autosampler are as follows:
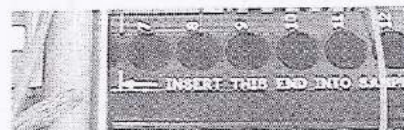
> Relay A - moves the sample probe DOWN
> Relay B - moves the sample probe UP.
> Relay C - moves the syringe carriage FORWARD
> Relay D - INJECTS the contents of the syringe
> Relay E - ADVANCES the tray one position
> Relay F - PRESSURIZES the sample vial

WARNING!
To avoid injury, keep your hands clear of the 8640 during operation.

See the event table shown at right for appropriate descriptions. The 8640 relay descriptions are also labeled on the right-hand side of the GC.

6. Set the autosampler air or nitrogen tank to 60psi. Set the carrier gas to 10mL/minute (the equivalent psi setting for your machine is labeled on the right panel of the GC). The amount of sample used to flush the needle can be adjusted by varying the pressure of the gas used to force the sample from the vial. This gas pressure is adjusted with the EPC trimpot on the top edge of the GC's front control panel, located directly above the vertical label "VIAL PRESSURE" on the front control panel. Using the event table at right, you should count 25 drops during the time that the gas is pressurizing the sample (0.600 minutes).

8640.evt		
EVENT TIME	EVENT	EVENT FUNCTION
0.000	ZERO	Zero data system signal
0.050	A ON	Sample probe DOWN
0.100	F ON	Vial pressure ON (pressurize the sample vial)
0.650	A OFF	Release pressure holding sample probe DOWN
0.700	F OFF	Vial pressure OFF
0.750	B ON	Sample probe UP
0.800	C ON	Syringe carriage FORWARD
0.850	D ON	Sample syringe INJECT
1.000	C OFF	Syringe carriage RETRACT
1.050	D OFF	Sample syringe RETURN
1.100	E ON	Tray advance ON
1.200	E OFF	Tray advance OFF
1.300	B OFF	Release pressure holding sample probe UP

7. The injection volume is factory set at 1µL, but is adjustable to 0-3µL. Loosen the 2 hex-head lock nuts, then turn the knurled nut while observing the needle in the syringe barrel to achieve the desired injection volume (please see the picture on the *Changing the Needle* page to locate the lock nuts and knurled nut).

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Changing the Needle

In the course of normal operation, the 8640 sample injection needle may become bent or otherwise damaged and require replacing. Make sure the syringe mechanism is fully retracted before starting; this is the default position to which it should return after a sample injection sequence.

Replacement needles are available from Central Instruments under part number 502743. Syringe barrel and needle sets are available under part number 503188. Call Central Instruments at:
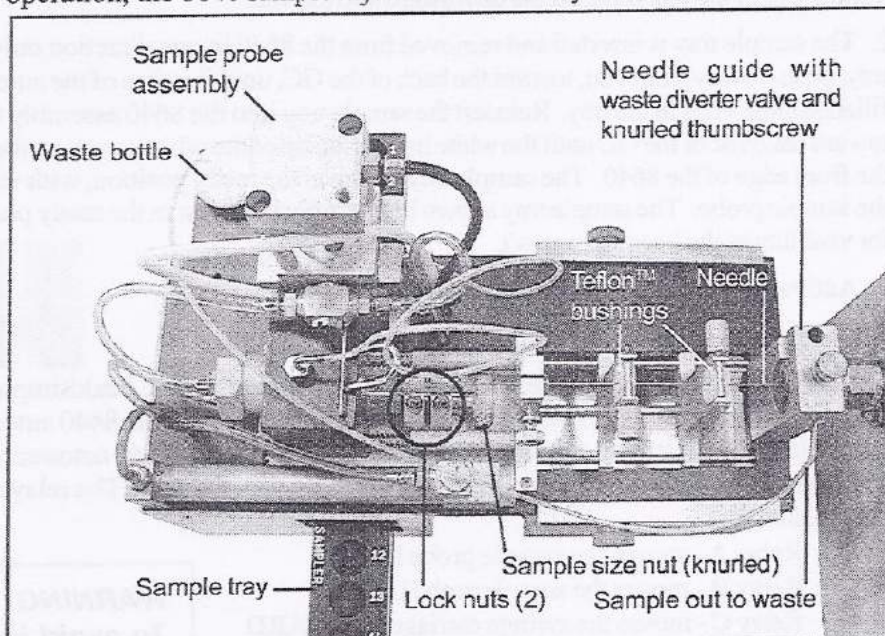
225-261-1917

Or write to:

P.O. Box 337
Greenwell Springs, LA 70739
USA.

Sample probe assembly

Waste bottle

Needle guide with waste diverter valve and knurled thumbscrew

Teflon™ bushings

Needle

Sample size nut (knurled)

Sample tray

Lock nuts (2)

Sample out to waste

1. To remove the needle, loosen the thumbscrew on the top of the waste diverter and the thumbscrew on the needle guide closest to the syringe barrel. Loosen and remove the two bushing retainers. Carefully lift out the needle, the two Teflon™ bushings and the waste diverter valve together. You will have to push the waste diverter valve out of the needle guide, and angle the needle tip out through the slot in the side of the waste diverter needle guide as you pull the needle from the syringe barrel.
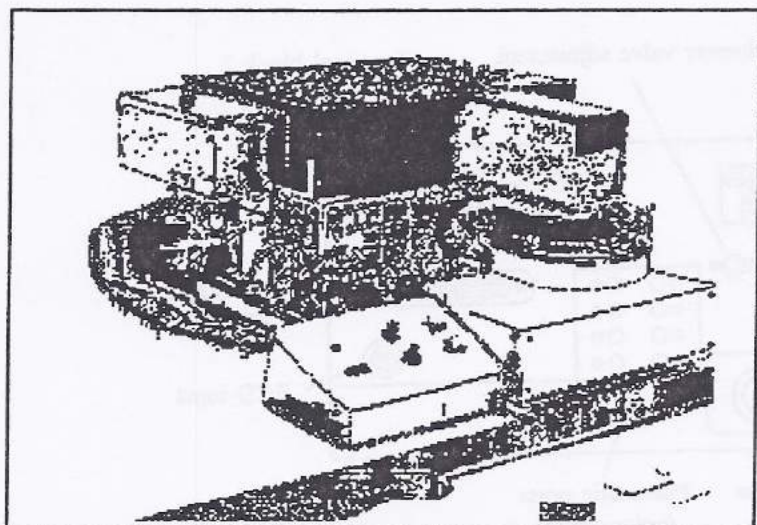
2. Slide the waste diverter valve and the two Teflon™ bushings off the old needle and onto the replacement needle.

3. Place the needle into the thumbscrew needle guide and the syringe barrel, and carefully angle the needle with the bushings and waste diverter valve into place, using the slot in the waste diverter needle guide to get the tip of the needle into alignment with the syringe barrel.

4. Position the two bushings in their cradles, then replace and tighten the bushing retainers. Tighten the thumbscrews on the needle guide and waste diverter.

5. Adjust the sample injection volume by loosening both hex-head lock nuts, then turning the knurled thumbscrew to achieve the desired volume. Tighten the lock nuts.

Loosen these 2 thumbscrews

Loosen and remove these 2 bushing retainers, then remove the 2 bushings

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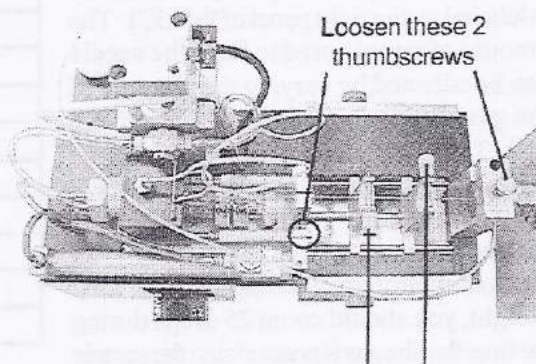
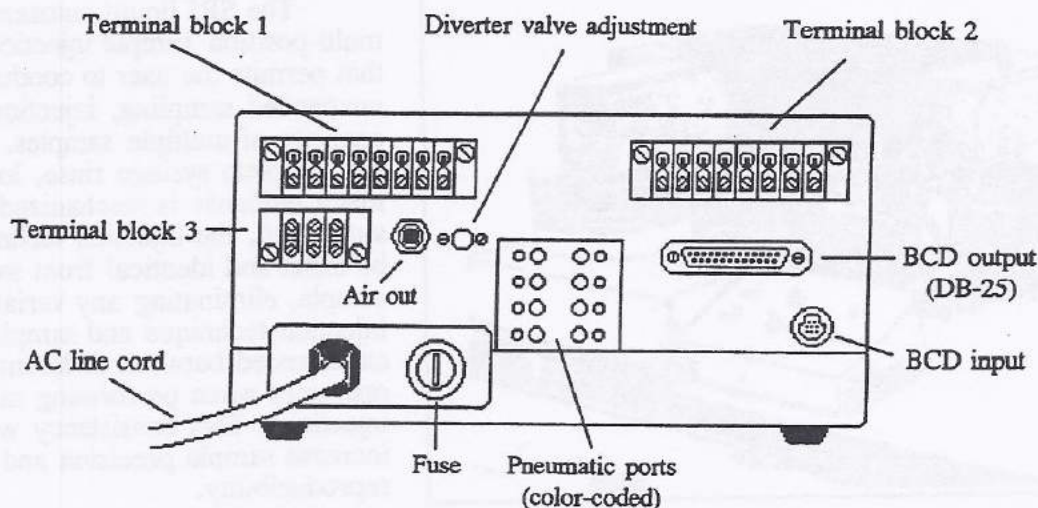SRI liquid autosampler connected to SRI 8610 gas chromatograph

The SRI liquid autosampler is a multi-position sample injection system that permits the user to conduct unattended sampling, injection and analyses of multiple samples. Because the complete syringe rinse, load and inject sequence is mechanized and automated, the injection technique will be exact and identical from sample to sample, eliminating any variation in injection technique and sample delivery experienced between different operators when performing manual injections. This consistency will increase sample precision and reproducibility.
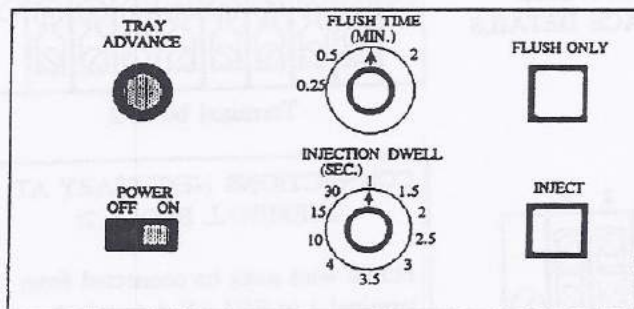
Front control panel of SRI autosampler controller

The autosampler is controlled automatically by PeakSimple software and/or manually at the autosampler control panel shown at left. Through software control, the autosampler is stepped through the sample vial positions until all samples inserted in the carousel have been injected and analyzed (without the need for operator intervention). A simple command in the event table (momentary activation of relay A) causes the autosampler to insert, flush, draw and inject the needle contents into the injection port. As soon as the needle has been withdrawn from the injection port, the autosampler is stepped to the next vial position to remain at the ready for the next actuation of relay A.
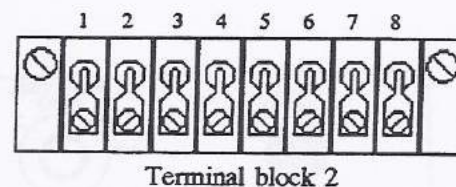
The sample vial tray may be manually advanced by hand or by pressing the TRAY ADVANCE actuator button. The amount of time (in minutes) that the syringe needle is flushed to clear the preceding sample is selectable using the FLUSH TIME control. The amount of solvent used to flush the syringe may also be varied by adjusting the sample pressurizing gas pressure. The actual volume of flush required will be dictated by the characteristics of the sample being injected. If not overly viscous, a sample flush volume of approximately 100 microliters should be adequate. If sample availability is limited to small volumes, then the flush may need to be reduced to economize on sample consumption. The amount of time that the needle remains in the injection port after the sample has been discharged from the syringe is also selctable using the INJECTION DWELL control. Selectable in seconds, the control permits the user to choose having the needle withdraw immediately upon having deposited its sample, or to maintain the needle in the injection port for an extended period, permitting any sample containing higher boiling or thermally labile components adequate time to exit the syringe and enter the injection port. High boiling components require longer needle-injection port dwell times than volatile components. A setting of 1 second signifies that the needle penetrates the septum, injects the sample and is withdrawn immediately, all within the duration of one second. A setting of 4 seconds permits the needle to dwell in the injection port for an additional 3.5 seconds.

Terminal block 1   Diverter valve adjustment   Terminal block 2

Terminal block 3

BCD output (DB-25)

Air out

AC line cord

BCD input

Fuse   Pneumatic ports (color-coded)

View of rear panel of SRI autosampler controller unit
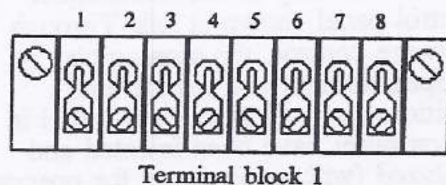
REAR PANEL INTERFACE DETAILS

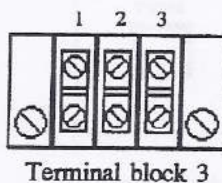1 2 3 4 5 6 7 8

Terminal block 1

1 2 3 4 5 6 7 8

Terminal block 2

CONNECTIONS NECESSARY AT TERMINAL BLOCK 1:

A jumper wire must be connected between terminals 7 and 8 of this terminal block (index enable position). Black wire provided.

1 2 3

Terminal block 3

CONNECTIONS NECESSARY AT TERMINAL BLOCK 2:

Purple wire must be connected from terminal 1 to RELAY A terminal on interface board. Black wire must be connected from terminal 2 to D.GND terminal on interface board.

CONNECTIONS NECESSARY AT TERMINAL BLOCK 3:

Three-wire control cable from autosampler connects here. Black wire connects to terminal 1, red wire to terminal 2 and green wire to terminal 3

PNEUMATIC HOSE TERMINAL

White hose from autosampler

Green hose from autosampler

Red hose from autosampler

White hose from autosampler vial pressurization regulator

Clear hose from autosampler

Black hose from autosampler

Yellow hose from autosampler

Blue hose from tray rotation pneumatic pressure regulator

Terminal block is color-coded

The procedure for installation of the autosampling carousel is simple and straight-forward. Pneumatic and electrical connections must be made at the controller unit and at the data system interface board (if in use by the system). Once these connections have been made, the autosampler is inspected for proper docking height with the chromatograph injection port. If the autosampler is intended for use with an early version of the model 8610 chromatograph (low-profile chassis with an injection port 6.5" above the countertop), the carousel will be equipped with three rubber feet that elevate the injection needle axis to exactly 6.5". Current production models are mounted on a platform that elevates the injection needle axis to exactly 10". This corresponds to the injection port height of the current production model 8610 chromatograph. When proper unit height has been verified at the injection port, the autosampler is mated to the chromatograph and operation may begin.

PNEUMATIC CONNECTIONS

In addition to connecting the six color-coded hoses from the autosampler control harness to their respective pneumatic terminals on the rear of the autosampler controller unit, two gas connections must be made at the lower two pneumatic hose terminals on the bulkhead. The white hose provided must be connected to the autosampler vial pressurization gas supply, using the provided regulator. This regulator should be set to 15 psi. Gas (typically air or nitrogen) is injected into the vial by the outer sleeve of the concentric sampling needle (needle within a needle), forcing sample to flow out of the vial through the center needle and into the injection syringe. The blue hose should be connected to the tray rotation gas supply, set to 60psi with the other regulator provided with the unit. This gas enables the tray mechanism to rotate, advance the samples and operate the injection mechanism.
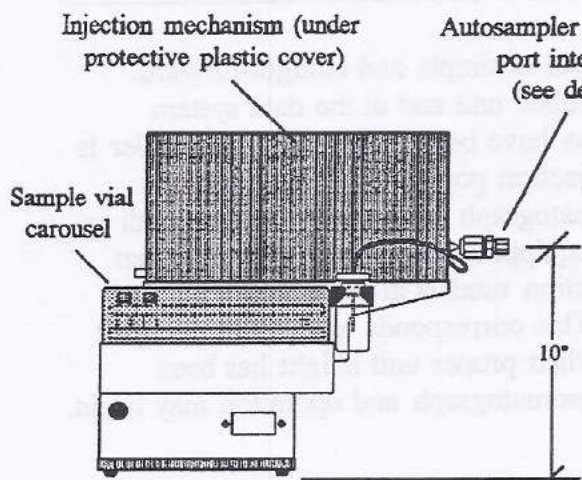
Pneumatic hose terminal

Each hose connector is accompanied by a color dot that specifies which hose to connect

Color-coded dots

White hose must be connected via a gas pressure regulator (provided), set to 15psi, to the autosampler vial pressurization gas supply (typically air).

Blue hose must be connected via a gas pressure regulator (also provided), set to 60psi, to the tray rotation gas supply (also typically air).

If air is used by both regulators, the main air supply pressure should be regulated to (at least) 75psi in order to maintain a stable gas supply.

ELECTRICAL CONNECTIONS

There are only three connections to be made by the user. A black jumper wire (provided) must be connected between terminals 7 and 8 of terminal block 1. The purple and black wires (also provided) must be connected to the interface board terminals labeled RELAY A and D.GND, respectively. These two wires provide the remote activation of the autosampler advance and sample circuitry (same as INJECT on the controller unit) by the data system or other remote device.
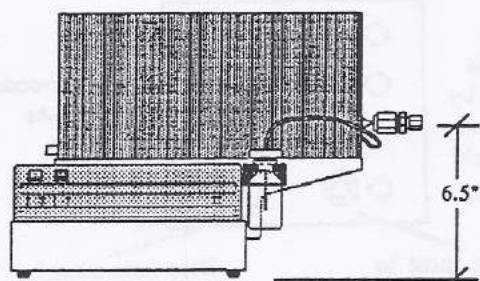
Terminal block 1

Black jumper wire connected between #7 and #8

Terminal block 2

Purple wire to interface board RELAY A terminal

Black wire to interface board D.GND terminal

C:\EP2\DOCS\LIQAUT

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Injection mechanism (under protective plastic cover)

Autosampler / injection port interface (see detail)

Standard SRI 1/8" injection port

Sample vial carousel

Autosampler injector extends from injection mechanism (under cover) at a height of 10" from the countertop, equal to the SRI chromatograph injection port height
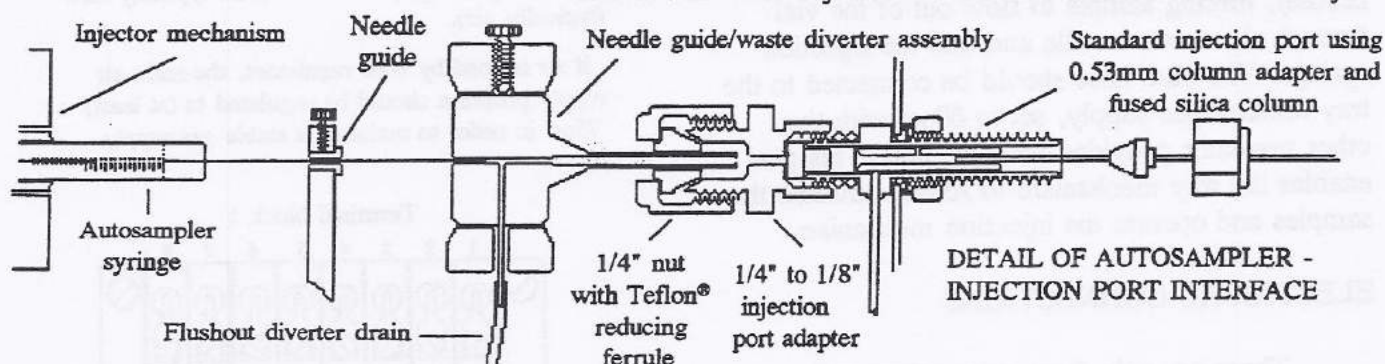
10"

10"

SRI

Right side view of SRI liquid autosampling carousel
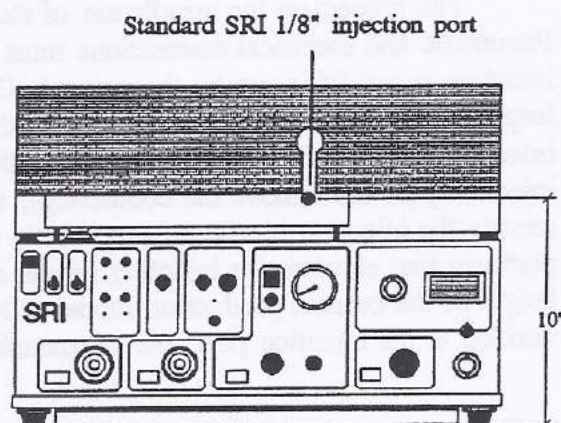
Front view of SRI model 8610 gas chromatograph

The SRI liquid autosampler injector mechanism is situated at a height equal to the injection port of the chromatograph. The injection port height of current production units is 10" (25.4cm). Previous models (pre-1992) employed an injector height of 6.5" (16.5cm). A special fitting is supplied with the autosampler that replaces the septum nut normally used to seal the injection port of the chromatograph. This fitting, also containing a septum, consists of a 1/8" to 1/4" adapter that is connected to the injection port. A special cylindrical brass fitting, employed as a needle guide and waste solvent diverter, is inserted into the needle end of the injector mechanism frame and secured by a

6.5"

Autosampler configured for early production chromatographs with 6.5" high injection port

Injector mechanism

Needle guide

Needle guide/waste diverter assembly

Standard injection port using 0.53mm column adapter and fused silica column

Autosampler syringe

Flushout diverter drain

1/4" nut with Teflon® reducing ferrule

1/4" to 1/8" injection port adapter

DETAIL OF AUTOSAMPLER - INJECTION PORT INTERFACE

thumbscrew. From this cylindrical fitting, a 1/8" metal tube protrudes. This tube, an extension of the needle guide, is secured to the chromatograph's injection port adapter by means of a 1/4" nut with a 1/4" to 1/8" Teflon® reducing ferrule. Once both fittings are in place, the units are docked together and the 1/4" nut is secured. Then the autosampler controller unit is located in a convenient location and normal operation can begin.
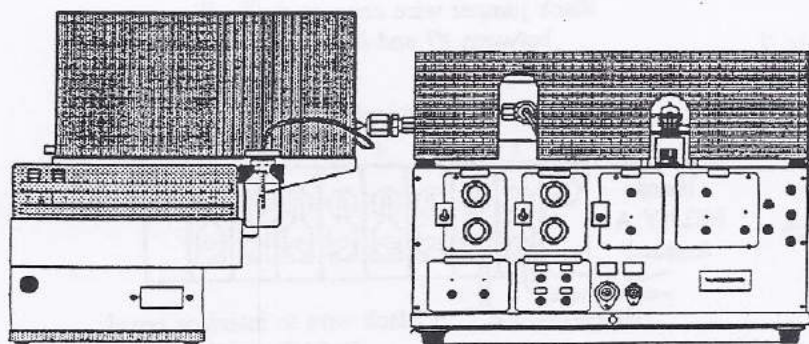
SRI Instruments
PeakSimple 2000
Chromatography Integration Software
Basic Tutorial
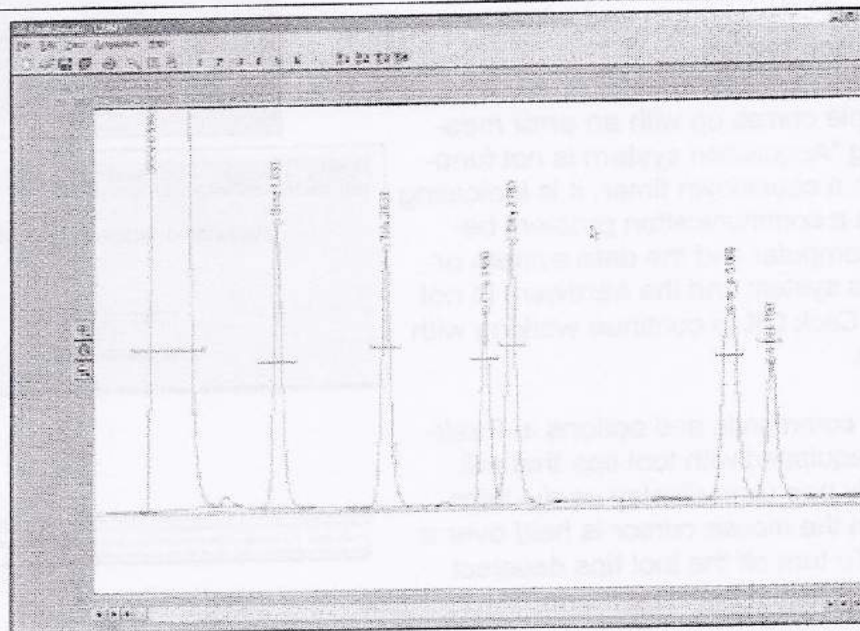
Installing PeakSimple 2000 from floppy disk or CD-Rom

A. Start the Windows operating system in use on your computer. (Windows 95, 98, ME, 2000)

B. Insert the PeakSimple 2000 disk or CD into your floppy disk drive.

C. Go to the **Start** menu in the bottom left hand corner of the windows screen and select **Run** from the set of icons.

D. From the run menu, type **X:\setup** (where **X** is the letter of your computers disk drive).

E. Now click on the **Continue** button with your mouse cursor or press the enter key on your keyboard to begin installation.

F. To complete installation follow the onscreen instructions provided by the installation wizard.

Installing PeakSimple 2000 from software download

A. Start the Windows operating system and use an online browser to access www.srigc.com.

B. From the menu on the left hand side of the screen select **Download our Software** and then download PeakSimple 2000 from the following page.

C. Save the file to a temporary folder and then double click on it from My Computer to allow the program to self-extract.

D. Once all the files have been extracted successfully double-click the install file and press the **Continue** button when prompted.

E. Follow the onscreen instructions to complete the installation of PeakSimple.

SRI Instruments 20720 Earl Street Torrance, CA 90503 U.S.A
Telephone: (310) 214-5092 Fax: (310) 214-5097 sales@srigc.com **www.srigc.com**

Launching PeakSimple 2000

1. Click on the windows **Start** button in the bottom left-hand corner of the screen. Select **Programs** and then **PeakSimple** from the list of program groups on the screen and then click on **PeakSimple**.

2. This will launch PeakSimple and initialize the data acquisition system.

3. If PeakSimple comes up with an error message stating "Acquisition system is not functioning" with a countdown timer, it is indicating that there is a communication problem between the computer and the data system or that the data system and the hardware is not connected. Click **OK** to continue working with PeakSimple.

4. Most of the commands and options in Peak-Simple are equipped with tool tips that will automatically pop up to display useful information when the mouse cursor is held over a command. To turn off the tool tips deselect the tool tips option in the Help menu.
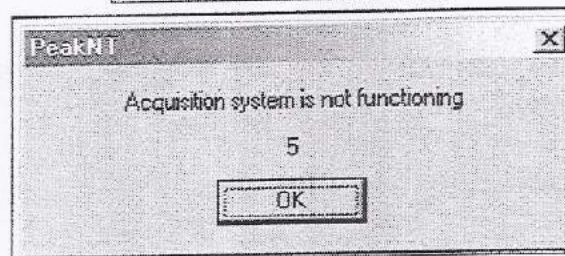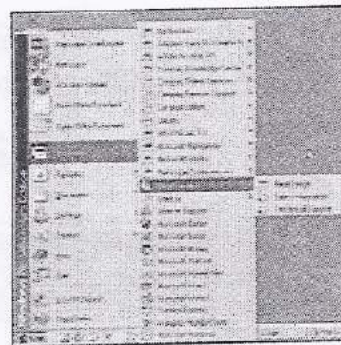
Opening a PeakSimple Data File

1. To open a PeakSimple data file or chromatogram, begin by selecting **File** in the Peak-Simple menu bar and then choose **Open...** from the set of options.

2. The Load Chromatogram File window is now open. The PeakSimple software includes a number of sample chromatogram data files that can be opened, displayed, and manipulated. One file, 602.CHR, will be used throughout the rest of the tutorial. Select file **602.CHR** from the PeakSimple directory, choose **Channel 1** as a destination channel, and then select **Open** to load the file.

PeakNT

Acquisition system is not functioning

5

OK

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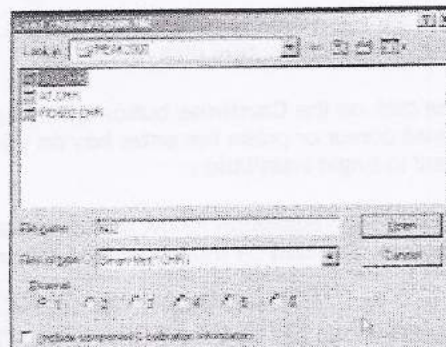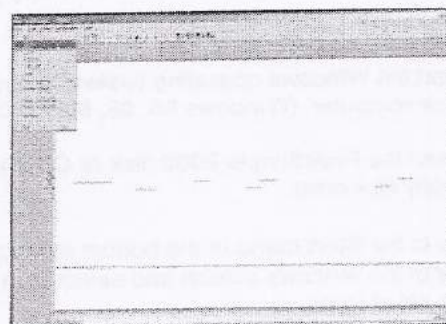
Adjusting Display Limits

1. To adjust the display limits of a chromatogram click on either the + magnifying glass icon or the - magnifying glass icon to the left of the chromatogram. This will increase or decrease the limits by a factor of two each time you click on the icons.

2. After opening chromatogram 602.CHR, practice making the display limits smaller but the peaks larger by clicking the + magnifying glass icon.

3. Practice making the display limits larger but the peaks smaller by clicking on the - magnifying glass icon.
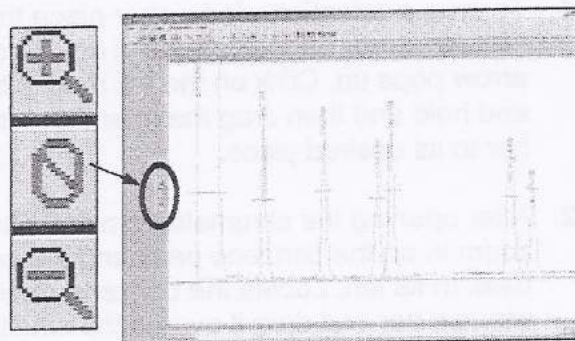
Zooming

1. To zoom in on a specific part of a PeakSimple chromatogram, click and hold the left mouse button and drag it over the desired area.

2. After opening chromatogram 602.CHR hold the left mouse button and drag it over the base of the toluene peak. Let go of the mouse button and there will be a larger view of the area that was selected.

3. To return to the original display limits of the chromatogram and unzoom the area selected press **F6** or select the unzoom icon located in the PeakSimple toolbar at the top of the screen.

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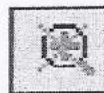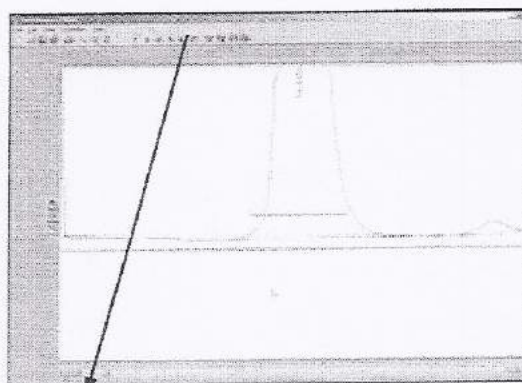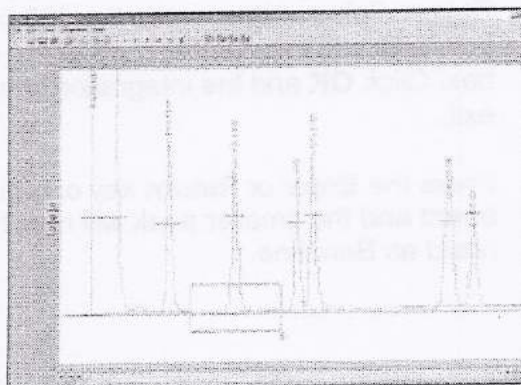
Dragging Retention Windows

1. To drag a retention window bar place the mouse cursor on the bar until a double sided arrow pops up. Click on the left mouse button and hold and then drag the retention window bar to its desired place.

2. After opening the chromatogram 602.CHR zoom in on the benzene peak and the smaller peak to its left. Locate the benzene retention window bar and drag it over to the smaller un-named peak to the left of the benzene. Because this is a small peak it is not immediately recognized.

3. Right click on the chromatogram over the un-named peak and select **Integration** from the resulting menu.

4. From the integration window locate the **Area Reject** dialogue box, erase the 100.0 in the box, and add the number **10.0** to the dialogue box. Click **OK** and the integration window will exit.

5. Press the **Enter** or **Return** key on your keyboard and the smaller peak will now be recognized as Benzene.

Manual Integration

1. To manually adjust the integration baseline and peak separation in a chromatogram use the manual integration toolbar provided by PeakSimple. To open up the manual integration toolbar select **Edit** in the PeakSimple menu bar and then click on the **Manual Integration** option. The manual integration toolbar will now appear to the left of the chromatograph.

2. The manual integration toolbar contains nine types of manual integration options. Four of the most commonly used options are **None** integration, **Drop** integration, **Based** integration, and **Rubber Band** integration.

3. To make a baseline ignore a peak use the None integration tool. After opening chromatogram 602.CHR and the manual integration toolbar, zoom in on the baseline of the solvent peak and the smaller unrecognized peak immediately to its right. Click on the **None** integration tool in the manual integration toolbar with the mouse cursor and then click on the valley between the two peaks where they meet the baseline. The area of the small peak is now added to the solvent peak.

4. To undo the changes made to a chromatogram at any time simply click on the **Undo** integration tool in the manual integration toolbar. After selecting this tool all integration changes made to the chromatogram will be undone.

5. Click on the **Undo** tool with your mouse cursor and select the **Drop** integration tool to enable the dropping of the baseline below the between the two peaks. After selecting the Drop tool click where the valley of the peaks meet the baseline with the cursor. The baseline should now be dropped below the base of the peaks and a line should extend from it to the baseline.
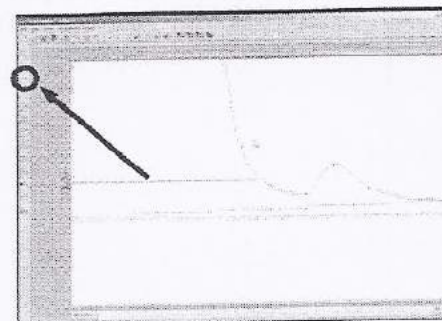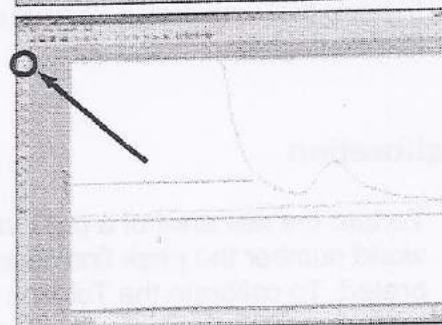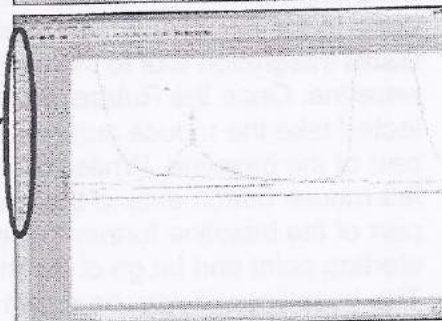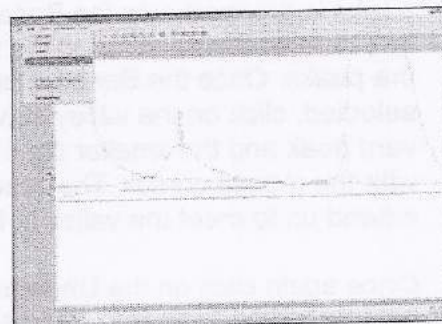
6. After the manual integration between the two peaks is dropped use the **Based** integration tool to raise the baseline to the valley between the peaks. Once the Based integration tool is selected, click on the valley between the solvent peak and the smaller peak to its right with the mouse cursor. The baseline will now extend up to meet the valley of the two peaks.
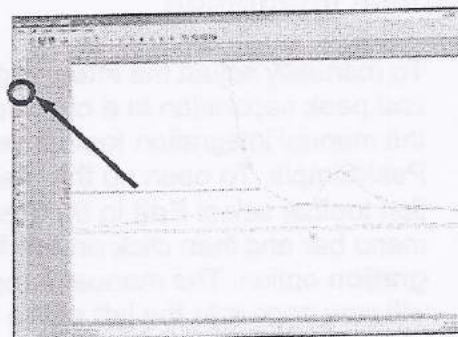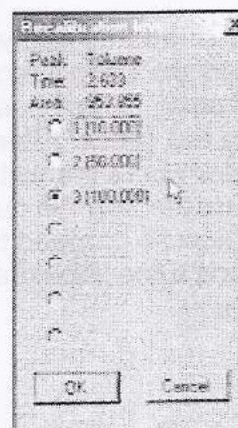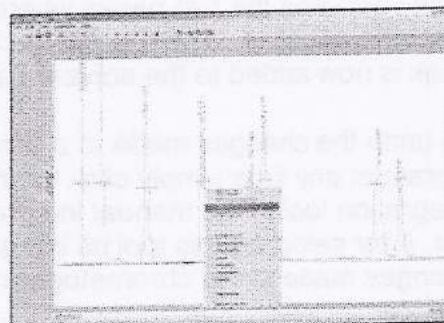
7. Once again click on the **Undo** tool in the manual integration toolbar to remove all changes done to the chromatogram. Select the **Rubber Band** integration tool to manually draw a baseline. Once the Rubber Band tool is selected take the mouse cursor and click on a part of the baseline. While holding down the left mouse button extend the line to another part of the baseline further to the right of the starting point and let go of the mouse button. The base line will now be drawn according to the line that was drawn using the Rubber Band integration tool.
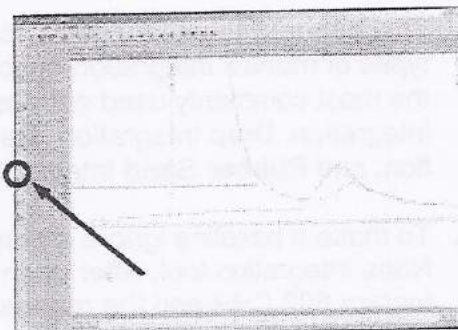
Calibration

1. To turn the raw area of a peak into a real-world number the peak first needs to be calibrated. To calibrate the Toluene peak in chromatogram 602.CHR, open up the file and then right click using the mouse on the Toluene peak. After right clicking on Toluene select **Calibrate Toluene** from the resulting menu.

2. From the Recalibration level window click on the third level radio button **3 (100.000)** and then select **OK** with your mouse cursor.

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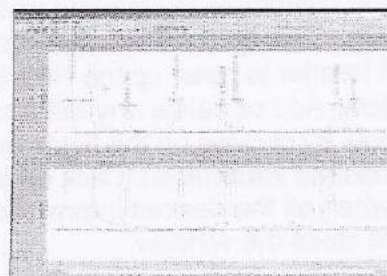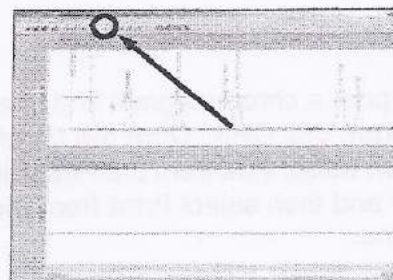Website NEW : www.chromalytic.net.au E-mail : info@chromtech.net.au Tel: 03 9762 2034 . . . in AUSTRALIA

3. After selecting OK from the Recalibration level menu the Calibration menu for Toluene will pop up. Check to make sure the flashing asterisk on the calibration curve is on level 3 and then click on the **Accept New** button to the right of the window.

4. Once the new data is accepted, click on the **Method** button immediately below the Accept New button. The Recalibration type window will now open allowing the user to select a method of calibration. By default the calibration type is set at Multiple Line Segments. Select the **Quadratic (Ax2+Bx+C)** radio button and then click on **OK** with the mouse cursor.

5. After changing the method of calibration click on **Statistics** in the upper right hand corner of the Calibration level window. The Calibration statistics window will pop up revealing the statistics for the calibration of Toluene. Click **OK** with the mouse cursor to close the Calibration statistics window and then select **Close** from the Calibration window to finish calibrating Toluene.
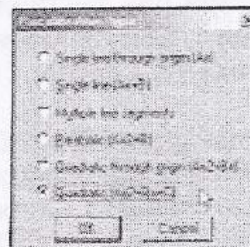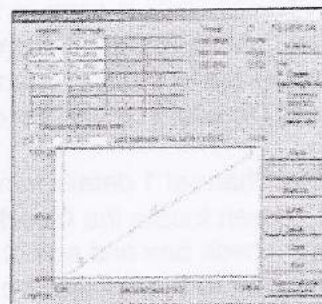
Overlay

1. To compare two or more chromatograms overlay them using PeakSimple. To overlay two chromatograms first open chromatogram 602.CHR and then click on the **2** button in the PeakSimple toolbar. A second chromatogram channel is now open in the PeakSimple window.

2. Once the second channel is open select **File** from the PeakSimple menu bar and then click on **Open**. The Load chromatogram file window will open up displaying a list of files to load. Select chromatogram **FID602.CHR** to load and then select the **2** channel radio button to load the chromatogram in the second channel.

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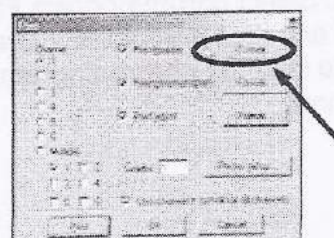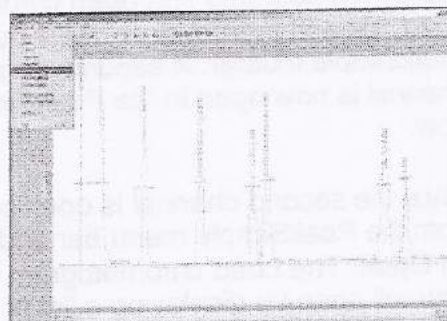Website NEW : www.chromalytic.net.au E-mail : info@chromtech.net.au Tel: 03 9762 2034 . . . in AUSTRALIA

3. Once FID602.CHR is open in the second channel right click using the mouse on the chromatogram in the first channel and select **Channel Details** from the list of options.

4. After the Channel 1 details window appears on the screen locate the **Overlay data in channel** check box and select it. Look to the dialogue box to the right of the Overlay data in channel check box and insert the number **2** in place of the 1. Click on **OK** with the mouse cursor to exit the Channel 1 details window.

5. The chromatogram FID602.CHR is now in place overlaid on top of chromatogram 602.CHR in channel 1. Chromatogram 602.CHR is in blue while FID602.CHR is in red.
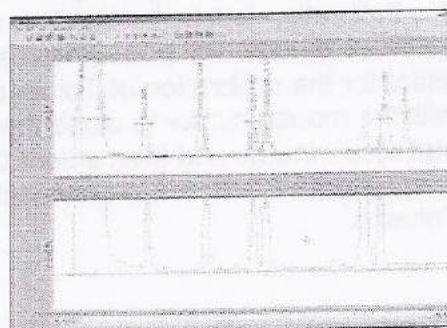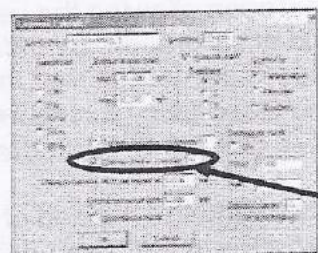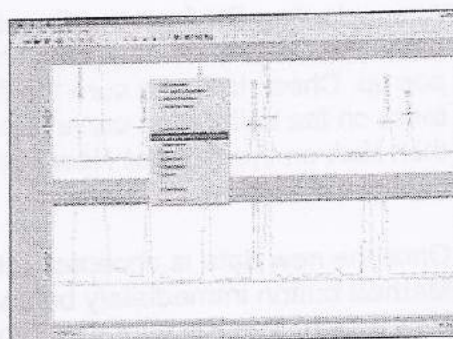
Printing a Chromatogram

1. To print a chromatogram first open chromatogram 602.CHR. Once the chromatogram is open select **File** from the PeakSimple menu bar and then select **Print** from the drop-down menu.

2. The Print window will open and will allow the user to customize the printing of a chromatogram. Click on the **Format** button for the Print header to open up the Header format window. Add or delete any information in the window by clicking on the fields and inserting the desired information. Click on the **OK** button when all the desired information is inputted to close the window.

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3. In the Print window click on the **Format** button for Print chromatogram to open up the Chromatogram format window. Locate the **Chart speed** dialogue box and insert the number of inches each minute on the chromatogram will take up when printed (for a nine minute run try **0.50** inches per minute). After the Chart speed is entered click on **OK** to exit the window.

4. In the Print window locate the Print report check box and click on the **Format** button to its right.

5. Once the Report format window is open click on **External** in the Available dialogue menu (on the left) and then click with the mouse cursor on the right facing arrow button to add External to the Selected dialogue box (on the right). After External is added to the Selected dialogue box click on **Units** with the mouse cursor and click on the right facing arrow button to add Units to the Selected dialogue box. Click on **OK** with the mouse cursor to exit out of the Report format window.

6. Select **Print** in the Print window to print the chromatogram or click on **OK** in the Print window to exit the window.
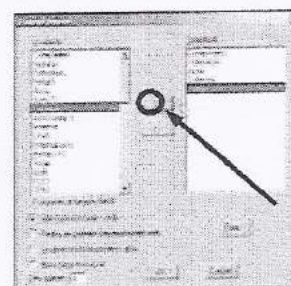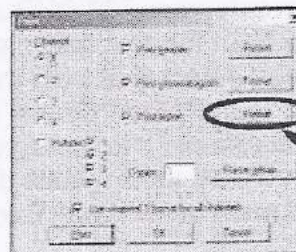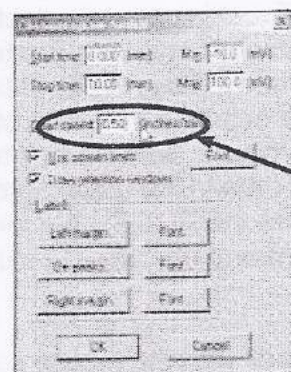
Exporting to Excel

1. In the PeakSimple toolbar click on the **Results** window button to open up the Results window. Once the Results window is open click on the **Copy** button to copy the results data to the Windows clipboard.

2. Make sure Microsoft Excel is loaded on the computer. If Excel is not loaded you can copy results data and chromatograms to Microsoft Word or PowerPoint. Open up Microsoft Excel by clicking with the mouse cursor on the **Start** button in the bottom left of the Windows screen and then **Programs** and then **Microsoft Excel** in the Windows Program menu.
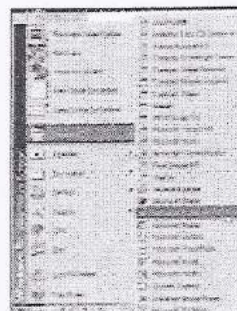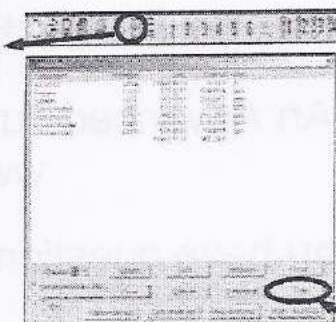
6. Once Excel is opened select **Edit** from the Excel menu bar and then **Paste** from the drop down menu. The results data is now placed into the columns and rows of Excel. Using the mouse cursor, select a box to the right of the results data in the Excel spreadsheet. Go back into the PeakSimple for Windows NT program and hit **Close** to exit the Results window.

7. Right click with the mouse cursor anywhere on chromatogram 602.CHR and select **Copy picture** from the resulting menu. Go back into Excel and select **Edit** from the Excel menu bar and then **Paste** from the drop down menu. The PeakSimple chromatogram will now be displayed next to its results data in the rows and columns of Microsoft Excel.
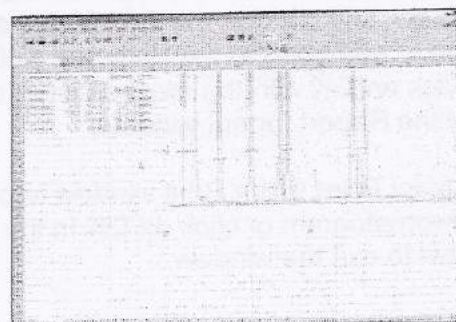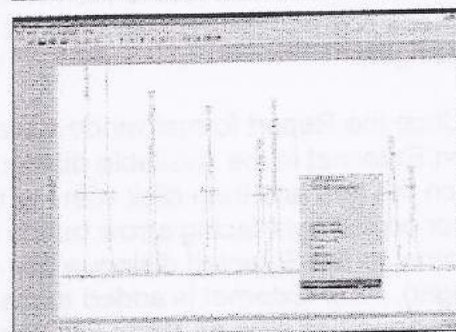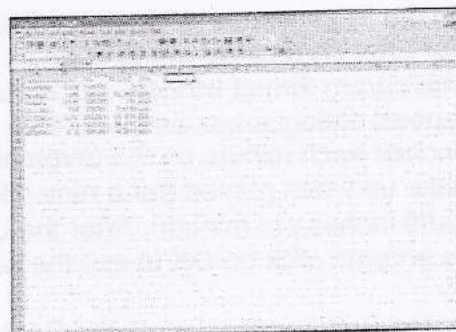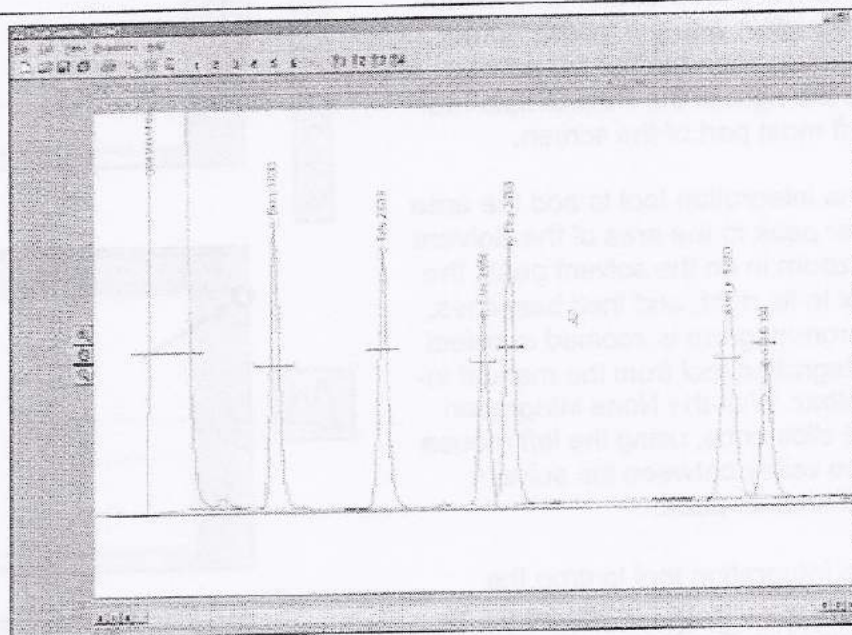
This concludes the PeakSimple 2000 Basic Tutorial

An Advanced Tutorial can be obtained by going to:
www.srigc.com online

If you have questions or would like to place an order call:
(310) 214-5092

PeakSimple 2000
Chromatography Integration Software

Advanced Tutorial

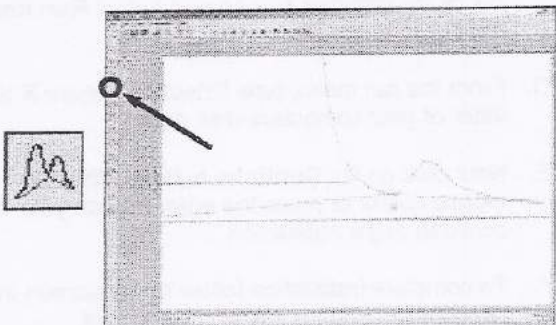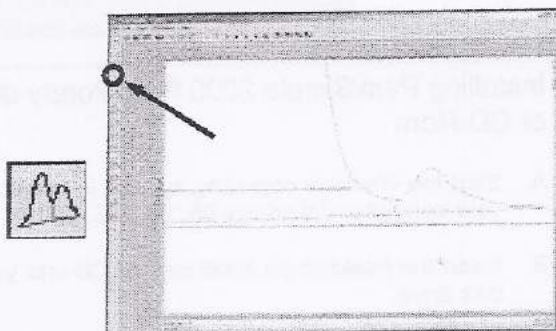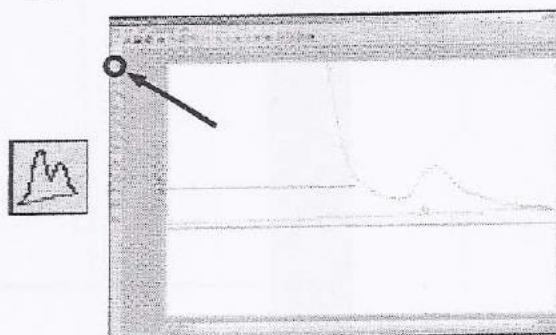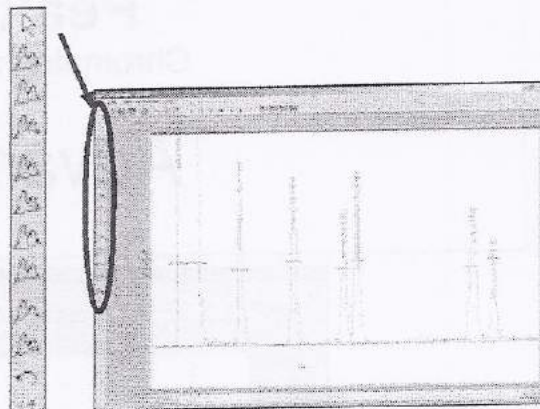

Installing PeakSimple 2000 from floppy disk or CD-Rom

A. Start the Windows operating system in use on your computer. (Windows 95, 98, ME, 2000)

B. Insert the PeakSimple 2000 disk or CD into your disk drive.

C. Go to the **Start** menu in the bottom left hand corner of the windows screen and select **Run** from the set of icons.

D. From the run menu, type **X:\setup** (where **X** is the letter of your computers disk drive).

E. Now click on the **Continue** button with your mouse cursor or press the enter key on your keyboard to begin installation.

F. To complete installation follow the onscreen instructions during the installation wizard.

Installing PeakSimple 2000 from software download

A. Start the Windows operating system and use an online browser to access www.srigc.com.

B. From the menu on the left hand side of the screen select **Download our Software** and then download PeakSimple 2000 from the following page.

C. Save the file to a temporary folder and then double click on it from My Computer to allow the program to self-extract.

D. Once all the files have been extracted successfully double-click the install file and press the **Continue** button when prompted.

E. Follow the onscreen instructions to complete the installation of PeakSimple.

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Manual Integration

1. To manually integrate the PeakSimple baseline in a chromatogram use the manual integration tools found in the manual integration toolbar. To open the manual integration toolbar first have chromatogram 602.CHR loaded and then select **Edit** from the PeakSimple menu bar. From the drop down menu select **Manual integration** with the mouse cursor. The manual integration toolbar will now be displayed to the right of the PeakSimple toolbar in the left most part of the screen.

2. Use the None integration tool to add the area of the smaller peak to the area of the Solvent peak. First, zoom in on the solvent peak, the smaller peak to its right, and their baselines. Once the chromatogram is zoomed in select the **None** integration tool from the manual integration toolbar. With the None integration tool selected click once, using the left mouse button, on the valley between the solvent peak and the smaller peak.

3. Use the Drop integration tool to drop the baseline from the valley of the two peaks to an existing baseline. To drop the baseline select the **Drop** integration tool from the manual integration toolbar. Using the mouse cursor, click on the valley between the solvent peak and the smaller peak to drop the baseline.

4. The Based integration tool raises the baseline to the valley between two specified peaks. With the baseline dropped, click on the **Based** integration tool button and then click on the valley between the solvent peak and the smaller peak to its right to raise the baseline to the valley.

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5. The Lead skim integration tool allows a peak's area to be skimmed off of the leading edge of another peak. To use the Lead skim tool first unzoom off of the solvent peak and the other smaller peak and then zoom in on the Chlorobenzene peak, the Ethylbenzene peak, and the baseline. After the chromatogram is zoomed click on the **Lead skim** integration tool button and then click on the valley between the two peaks with the mouse cursor.
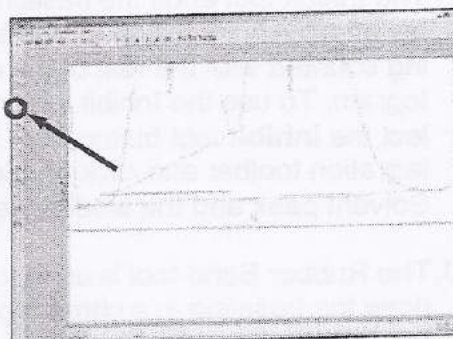
6. The Trail skim integration tool is similar to the Lead skim tool except a peak's area is now skimmed off of the trailing edge of another peak. Select the **Trail skim** tool button from the manual integration toolbar and then click on the valley between the Chlorobenzene and Ethylbenzene peaks with the mouse cursor to see the Ethylbenzene peak skimmed off of the Chlorobenzene peak.
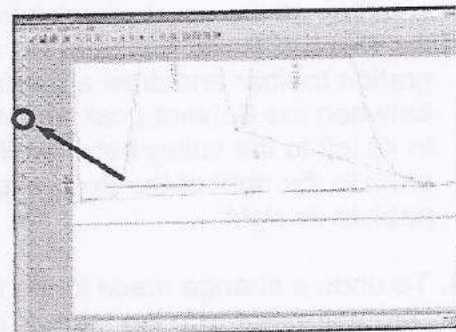
7. The Lead horizontal tool constructs the baseline horizontally for the leading peak while the trailing peak's baseline stretches from the horizontal line to the next valley. Unzoom off of the Chlorobenzene and Ethylbenzene peaks and instead zoom in on the Solvent peak, the smaller peak to its right, and the baseline. Click on the **Lead horizontal** integration tool in the manual integration toolbar and then click, using the left mouse button, on the valley between the solvent peak and the other smaller peak.
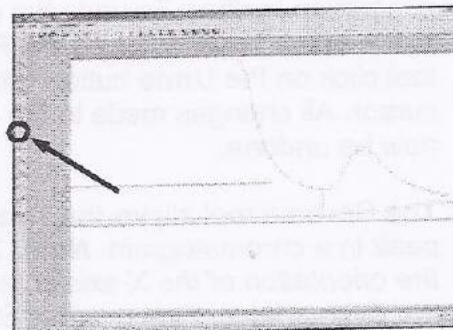
8. The Trail horizontal integration tool drops the baseline horizontally for the trailing peak while the lead peak's baseline stretches from the horizontal line to the previous valley in the chromatogram. After selecting the **Trail horizontal** tool in the manual integration toolbar click with the mouse cursor on the valley between the two zoomed in peaks.
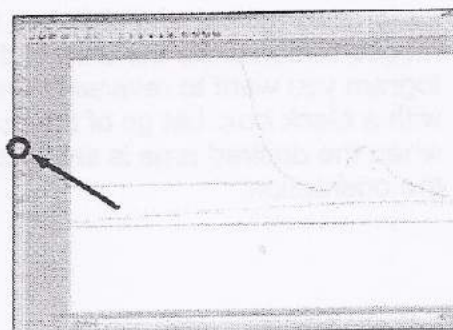
9. The Inhibit tool ends the baseline after a valley effectively inhibiting a peak's area from being counted with the rest of the chromatogram. To use the Inhibit integration tool select the **Inhibit** tool button from the manual integration toolbar and click on the valley of the Solvent peak and the smaller peak to its right.
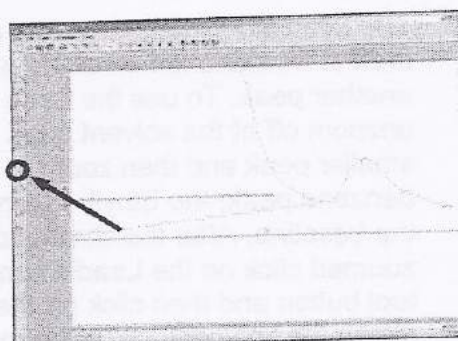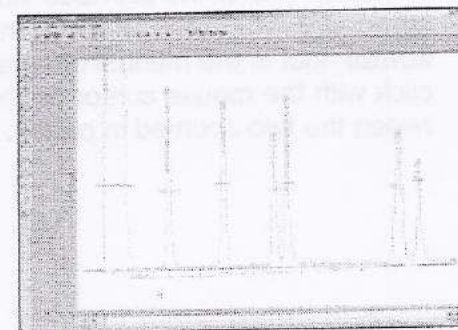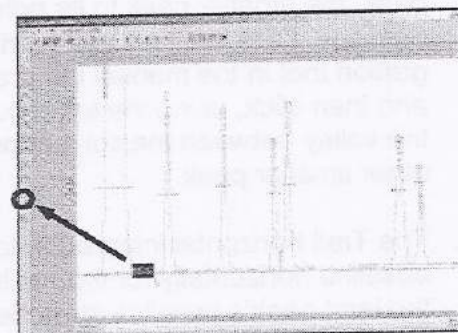
10. The Rubber Band tool is used to manually draw the baseline in a chromatogram. To use the Rubber Band tool first scroll the X-axis scrollbar all the way to the left to **0.000**. Select the **Rubber Band** tool from the manual integration toolbar and draw a line from the valley between the Solvent peak and the small peak to its left to the valley between the smaller peak to the right of the Solvent peak and the peak to its right.
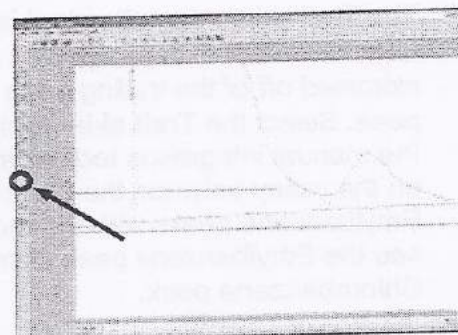
11. To undo a change made to the baseline of a chromatogram with the manual integration tools use the Undo button found in the manual integration toolbar. To undo the changes made to the baseline using the Rubber band tool click on the **Undo** button with your mouse cursor. All changes made to the baseline will now be undone.

12. The Reverse tool allows the inverting of a peak in a chromatogram. *Note: To reverse the orientation of the X-axis in real time go to the Events table*. First unzoom off of the Solvent peak and the smaller peak to its right and then select the **Reverse** tool from the manual integration toolbar and click and hold the left mouse button while the area of the chromatogram you want to reverse is dragged over with a black box. Let go of the mouse button when the desired area is selected to reverse the orientation.

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13. The Zero tool is used to set the value of the data line at a selected point and following in the chromatogram to zero. First undo the changes done to the chromatogram by the Reverse tool by reopening 602.CHR in the PeakSimple menu bar. *Note: Changes made to a chromatogram by the Reverse tool and the Zero tool cannot be undone with the Undo tool.* Once the file is reopened click on the **Zero** tool and click anywhere on the baseline between the Ethylbenzene peak and the two peaks to its right with the mouse cursor to set the data line at zero.
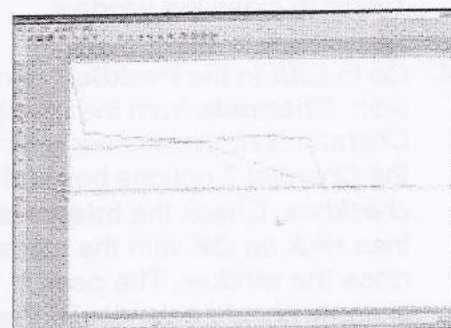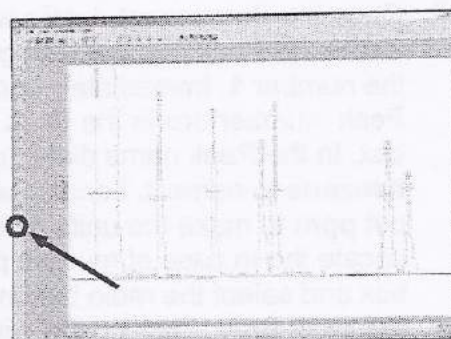
Creating Component Tables

1. To create a component table from scratch open up a second channel in the PeakSimple window by clicking on the Display Channel **2** button in the PeakSimple toolbar. Once the second channel is open click on **File** and then **Open** to get to the Load chromatogram file window. Select file **FID602.CHR** from the list of files and select the Channel 2 radio button to open the file in channel 2. Click **OK** with the mouse cursor to load the file.

2. In channel 2 locate the second tall peak from the left and right click on it with the mouse cursor. From the resulting menu select **Add component** to add a retention window bar to the peak. Once again right click on the peak and select **Edit component** from the menu to open up the Component details window.
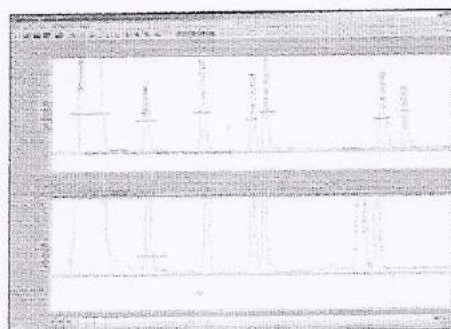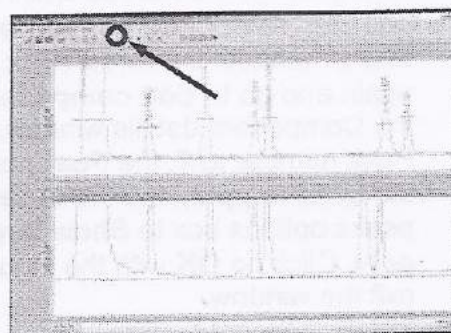
3. Once the Component details window is open locate the Peak number dialogue box and add the number **1**. Immediately underneath the Peak number box is the Peak name dialogue box. In the Peak name dialogue box input **benzene** to name it. Locate the Units box and put **ppm** to make the units parts per million. Locate the In case of multiple peaks options box and select the radio button for **Show largest peak only**. Click on **OK** with the mouse cursor to close the window.
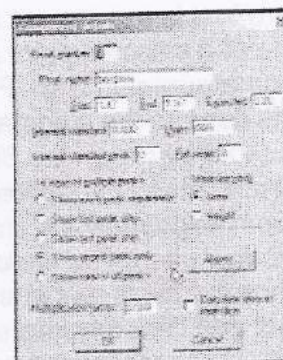
4. Go to **Edit** in the PeakSimple menu bar and then **Channels** from the resulting menu. The Channel controls window is now open. Locate the Channel 2 options box and the Integrate checkbox. Check the **Integrate** checkbox and then click on **OK** with the mouse cursor to close the window. The peak in the second channel should now identify itself as benzene.

5. Locate the large peak to the right of the benzene peak in the second channel. Right click and then select **Add component** to add a retention window bar to the peak. Right click again and go to **Edit component** to open up the Component details window. Change the Peak number to **2**, the Peak name to **toluene**, the Units to **ppm**, and the In case of multiple peaks options box to **Show largest peak only**. Click on **OK** with the mouse cursor to exit the window.
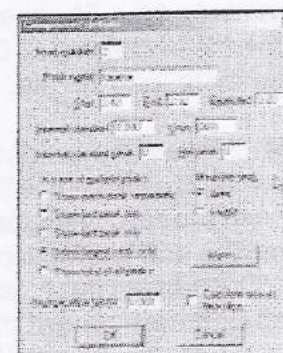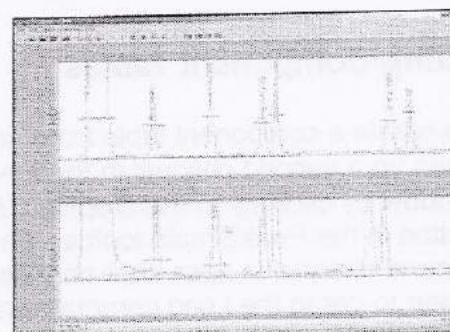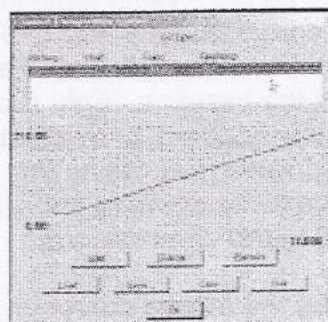
7. Right click anywhere on the second channel and select **Components** from the list of options. Once the Channel 2 components window is open make sure all the data is correct and then click on **Save** to save the Component data to disk. Name the file **Ctable** and then click on **OK** to close the window. An unlimited number of component windows may be added to the component table.
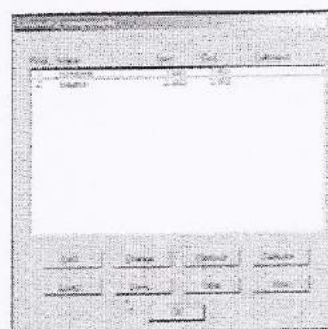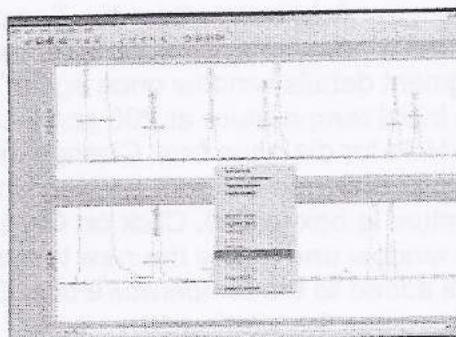
Temperature Programming

1. To modify the temperature programming in PeakSimple first open chromatogram 602.CHR and then right click anywhere on the chromatogram. From the drop down menu select **Temperature** to open up the Temperature control window.

2. In the Temperature control window click using the mouse cursor on the set of numbers in the box and select **Change** from the group of buttons below. The Temperature segment details window will open allowing the modification of the temperature programming. Locate the Hold for dialogue box and insert a **2** in the box. Click on **OK** to close the window and go back into the Temperature control window.

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3. Select the **Add** button from the Temperature control window to open up the Temperature segment details window once again. Leave the Initial temperature at 200 and insert a **1** in the Hold for dialogue box. Change the Then ramp at dialogue box to **5** and the Until temperature is box to **250**. Click on **OK** to close the window and to see the new temperature data added to the temperature box. Click on **OK** to close the window.
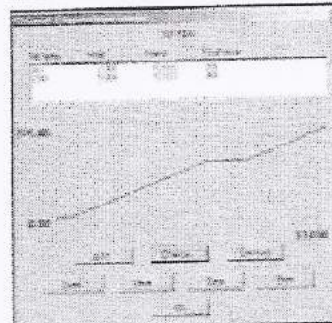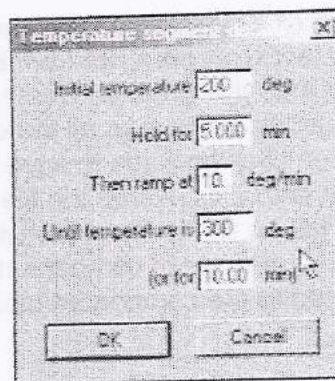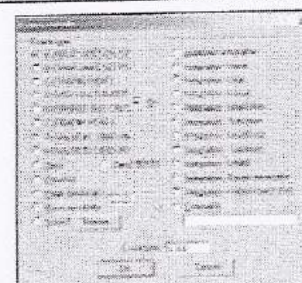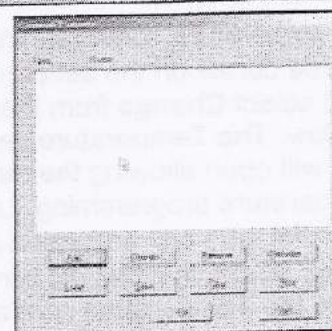
Events Table

1. To modify up the Events table in PeakSimple open up chromatogram 602.CHR and zoom in on the benzene peak, the smaller peak to its right, and the baseline. Right click anywhere on the chromatogram and select **Events** from the drop down menu. Doing this will open up the Events window where specific events can be added to the chromatogram.
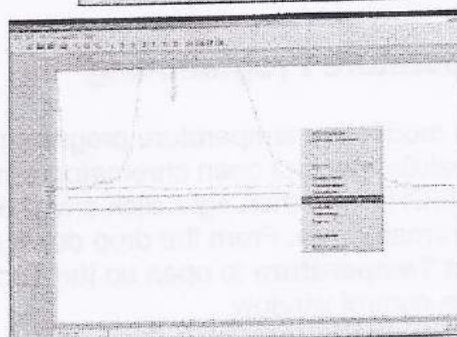
2. Click using the mouse cursor on the **Add** button to view the Event details window. A list of event types are available with their radio buttons to either select or deselect the event. *Note: The event types to the left of the window are real-time and thus will only affect the chromatogram when A/D hardware is connected. The event types to the right are concerned only with integration and their changes will be immediately evident after returning to the main screen and selecting **Reintegrate** from the **Edit** menu bar.*

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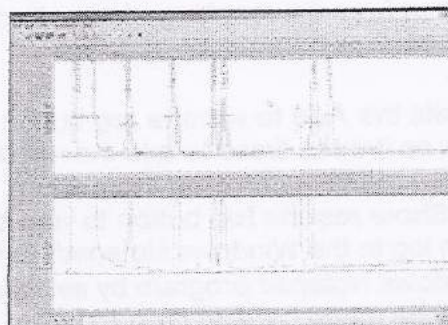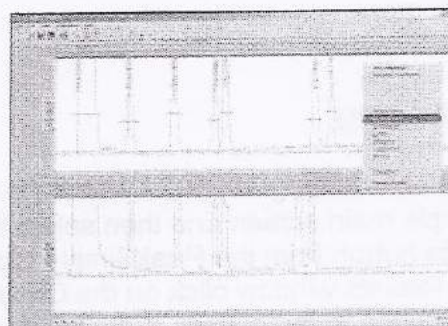*Importers & Manufacurers*
www.chromtech.net.au

3. In the Event details window locate and select the relay **G** radio button with the mouse cursor and then locate the Event time dialogue box and enter **.1** in the box. Click on **OK** to exit the window. *Note: The relay might be used to actuate a valve when hardware is connected.* The event type will now be added to the Events table. Select the **Add** button and now locate and select the **Zero** event type radio button. Leave the Event time box at 0.000 and once again click on **OK** to exit the window and add the event to the Events table. *Note: The Zero event auto-zeros the detector signal at the beginning of the run.* Click on the **Add** button again and select the **Integration-Based immediate** radio button in the Event details window and input **1.86** in the Event time dialogue box. Select **OK** to exit the window.

4. There are now three events in the Events table. Click on **OK** to exit the Events window and then hit the **Enter** button on the keyboard to reintegrate the baseline according to the events in the Events table. Notice that the baseline is connected to the data line at 1.86 minutes.
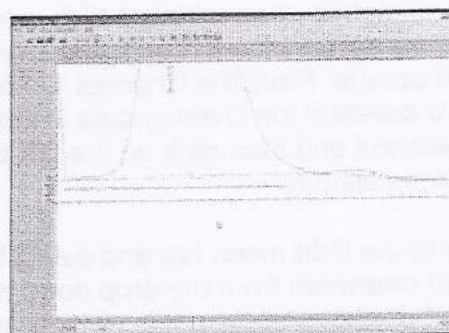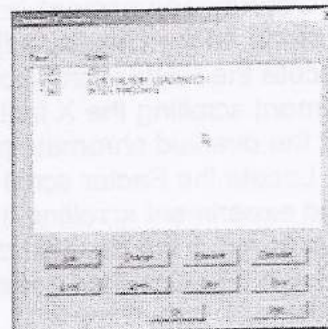
Overlay and Subtract

1. To overlay one PeakSimple chromatogram on top of another chromatogram open up a second channel in the main screen and load chromatogram 602.CHR in the first channel and chromatogram FID602.CHR in the second channel. Right click anywhere in the first channel and select **Channel details** from the drop down menu.

2. In the Channel 1 details window locate the Overlay data in channel checkbox and check it and then input a **2** in the dialogue box to the right. The chromatogram in channel 2 is now overlaid on top of the chromatogram in channel 1. The overlay appears in a different color.

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3. Right click anywhere on the first channel and select **Overlay adjustment** from the drop down menu. In the Overlay adjustment window locate the Factor scroll box in the X box. Experiment scrolling the X factor up or down to shift the overlaid chromatogram to its right or left. Locate the Factor scroll box in the Y box and experiment scrolling the Y factor up or down to move the overlaid chromatogram up or down. Click on the **Close** button to close the window.

4. To subtract a chromatogram in one channel from another channel, right click using the mouse cursor on channel 1 and select **Channel details**. From the Channel 1 details window deselect the Overlay data in channel checkbox and then click on the **OK** button to exit the window.

5. Go to the **Edit** menu bar and select **Subtract/ Add channels** from the drop down menu. In the Subtract/add channels window make sure the Subtract radio button is selected and that channel 2 is being taken from channel 1. Click on the **OK** button to make the changes take effect and have channel 2 subtracted from channel 1. The normal way to use this feature is to subtract a drifting baseline from a chromatogram.
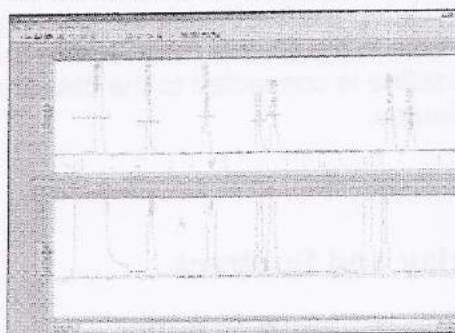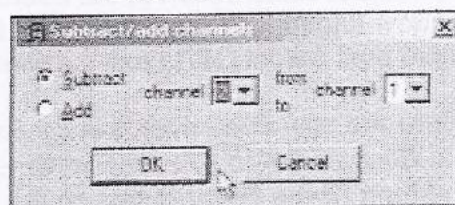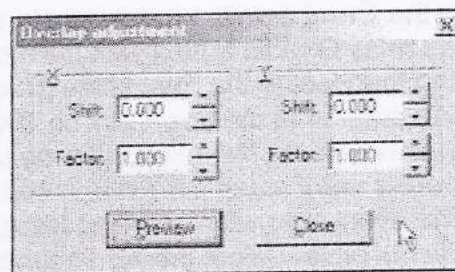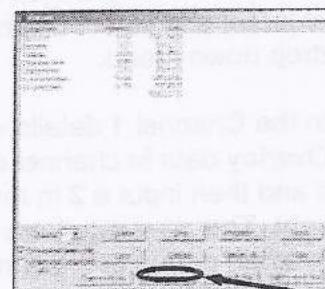
Results Log

1. Open chromatogram 602.CHR in the Peak-Simple main screen and then select the **Results** button from the PeakSimple toolbar. In the Results window click on the **Clear results log** button at the bottom of the window. Click on **Yes** from the resulting window to clear the results.

2. Locate the **Add to results log** button and click on it three times to add the results on the screen to the Results log three times. Click on the **Show results log** button to view the results log in the Windows Notepad. Exit the Windows Notepad program by selecting **File** from the menu bar and then **Exit**.

3. In the Results window locate the **Copy re-sults log** button at the bottom of the window and click on it with the mouse cursor (don't confuse the Copy button with the Copy results log button). Open up Microsoft Excel (or if Excel is not loaded Microsoft Word or Power-Point) and select **Edit** from the menu bar and then **Paste** to copy the results log to Excel.

4. Go back into PeakSimple and close the Results window by selecting the **Close** button. Right click using the mouse cursor on the chromatogram and select **Postrun** from the drop down menu to open the Post-run actions window. From the window locate the Add to results log checkbox and add a check to the box. By selecting the Add to results log checkbox all results from data analysis will automatically be added to the results log after the run is done. Click on **OK** to exit the window. In this way a summary of many analyses can be automatically created and then exported from PeakSimple.
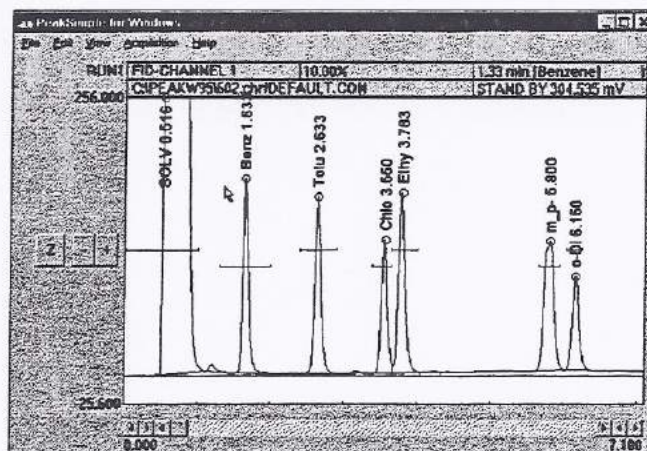
Copy results log
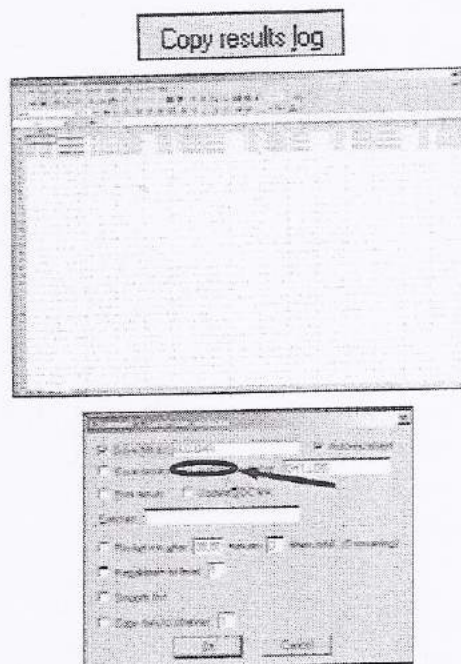
This concludes the PeakSimple 2000 Advanced Tutorial

Further documentation can be obtained by going to:
www.srigc.com online

If you have questions or would like to place an order call:
(310) 214-5092

PeakSimple for Windows Software and Chromatography Data System Validation Statement
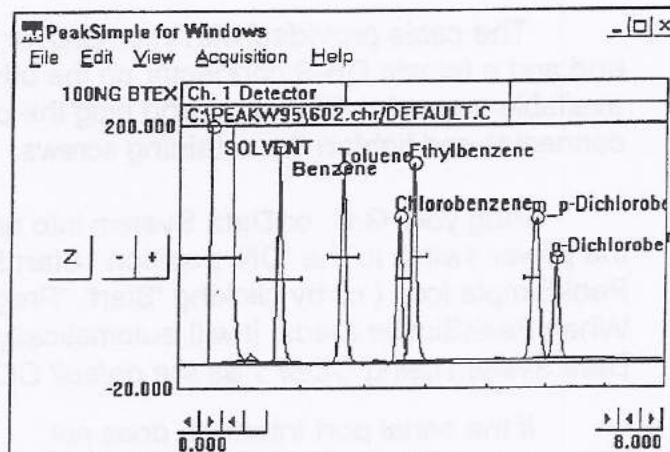

February 1, 1999

1)   PeakSimple for Windows software and Chromatography Data System (PeakSimple) is written, manufactured and maintained by SRI Instruments, Inc. a Nevada Corporation.

2)   PeakSimple for Windows software has been under continuous development since 1994. Periodic testing of the software is performed by SRI employees.

3)   PeakSimple software is designed to be self-validating to enable quick verification by customers that PeakSimple is functioning consistently, reliably and according to specifications under actual operating conditions.

4)   Self-validation is performed by configuring PeakSimple into the "Loopback mode" (see loopback instructions in manual). In this mode, an actual user generated chromatogram which is loaded into channel 4 is re-played (like a tape recorder) through the TP2 output channel, and then re-acquired and processed through any one of the remaining input channels. This is done 7 or more times to insure that data is being processed consistently and reliably.

The results from multiple loopback analyses are used to calculate the percent relative standard deviation (precision) of each peak in the chromatogram. Chromatographic data is highly variable, and the precision obtained is dependent on many factors including the peak shape, signal to noise ratio, interferences, co-eluting peaks, data acquistion rate and customer selected integration parameters. For this reason, self-validation is more valid than factory validation, since self-validation takes into account the exact chromatographic conditions and user specified parameters in effect for the particular application whereas factory validation can not.

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Getting Started

In this section, we will cover the basic information needed to set up proper communication with your G.C. or Data System hardware.

The Windows version of PeakSimple requires the use of the serial port interface that is built into most 8610-C and Model 310 gas chromatographs. This data acquisition and interface unit permits you to acquire up to four separate channels of data simultaneously without the need for additional hardware or acquisition boards.

```
PeakSimple for Windows                    _ □ ×
File  Edit  View  Acquisition  Help

100NG BTEX  Ch. 1 Detector
200.000      C:\PEAKW95\602.chr/DEFAULT.C

            SOLVENT   Toluene Ethylbenzene
                      Benzene
                              Chlorobenzene m p-Dichlorobe
  z   -  +
                                          o-Dichlorobe n

 -20.000

  ◄ ► ◄                                      ► ◄ ►
   0.000                                      8.000
```

The earlier IBM PC-compatible ISA expansion bus data acquisition cards (AD100 and AD110) used by PeakSimple II and PeakSimple III data systems are *not supported* by PeakSimple for Windows. However, all chromatograms acquired using DOS-based PeakSimple II and PeakSimple III continue to be compatible with this Windows version and may be imported as native files.

Identifying Your COM Port

Before attempting to establish communication between your G.C. and the serial data system interface, be sure to check that all the necessary electrical connections have been made, including the connection of any optional remote start cables.

Select an unused serial port on your PC and identify the COM port number assigned to it. It is important that this port NOT SHARE AN INTERRUPT with any other device used in your computer. Typical PCs are equipped with two COM (or serial) ports. COM 1 is typically used by the mouse or some other pointing device. COM 2 may be open (unused) or shared with another device, such as a fax modem, scanner or other peripheral. Determine which COM port you will use and remove any other device that may be in contention with that specific COM port number. Refer to your PC's hardware manual for instructions on changing COM port addresses and device drivers.

Most COM ports are provided with DB-9 connectors (nine pins configured in two rows - 5 pins over 4 pins - within a D-shaped plug or chassis connector). If your PC has a DB-25 serial port (25-pin connector), you will require a DB-25 to DB-9 adapter.
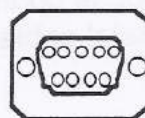
DB-9
Serial Port
Connector
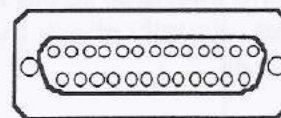
DB-25
Serial Port
Connector

Establishing Communication

The cable provided with your G.C. or data system has a male DB-9 plug on one end and a female DB-9 connector on the other end. Plug one end of this cable into your available computer COM port and plug the other end into the G.C. or Data System DB-9 connector and tighten the retaining screws.

Plug your G.C. or Data System into an approved GFCI protected outlet and turn the power switch to the 'ON' position. Start PeakSimple by double-clicking on the PeakSimple icon (or by clicking 'Start', 'Programs', 'PeakSimple' in Windows 95). When PeakSimple loads, it will automatically attempt communication with your G.C. or Data System using *COM 1 as the default COM port.*

If the serial port interface does not respond, you will see the following error messages appear on the screen: **"Can't wake- check power and cable"** followed by the message **"Acquisition system is not functioning"**.

These messages indicate that your computer cannot communicate with your G.C. or Data System through the default COM port, COM 1. You will need to set up the correct COM port in PeakSimple. To do this, click on the EDIT pull-down menu and select **OVERALL**. Change the **PORT NUMBER** to the COM port into which you chose to plug your interface cable. Click **O.K.**.

If at anytime you wish to force PeakSimple to reinitialize communication, click on the **ACQUISITION** pull-down menu and select **RE-INITIALIZE**. If the COM port information is correct and communication errors still appear when the computer attempts to activate the serial port interface, check the serial port connections at both ends of the interface cable for loose connections. Also, visually check the serial cable for nicks or cuts.
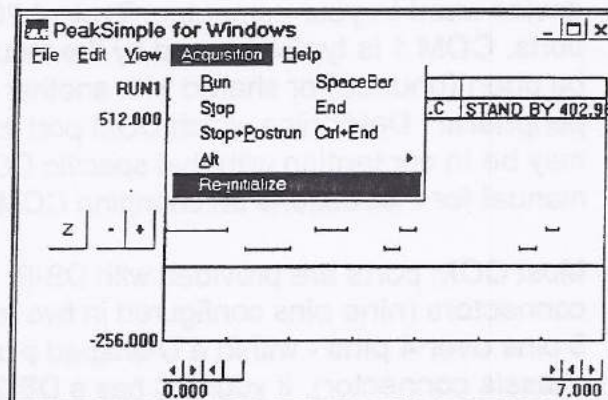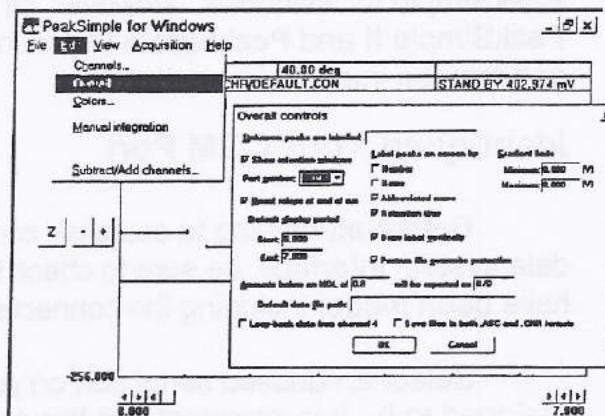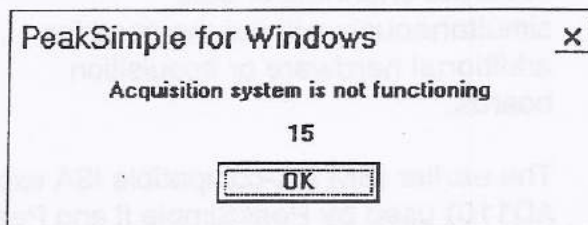
It is important to understand that in order for PeakSimple to communicate with your G.C. or Data System, at least ONE channel must be **ACTIVE**. To determine which channels are active, click on the **EDIT** pull-down menu and select **CHANNELS**. A channel is active if the box next to **ACTIVE** is marked with a checkmark. The **EDIT-CHANNELS** menu is described in greater detail in the **EDIT** section in this manual.

Chapter: PeakSimple

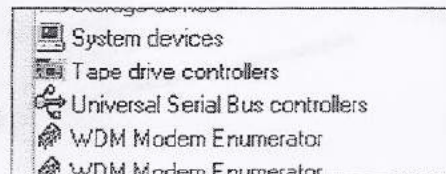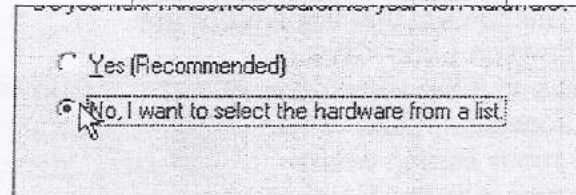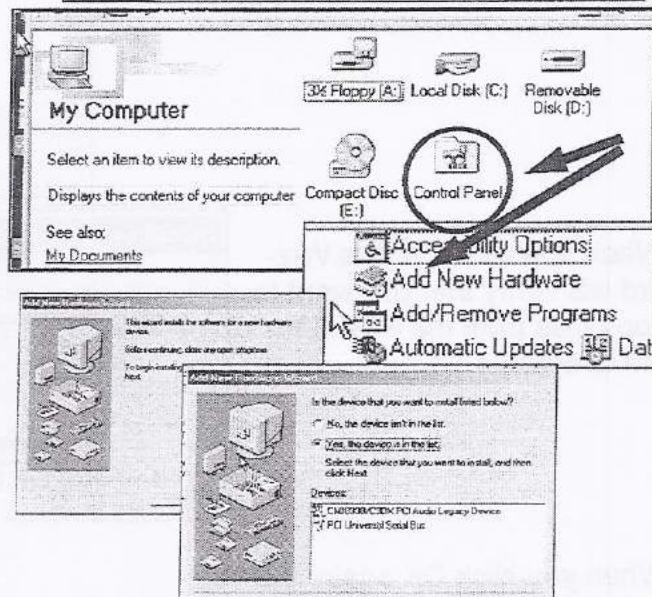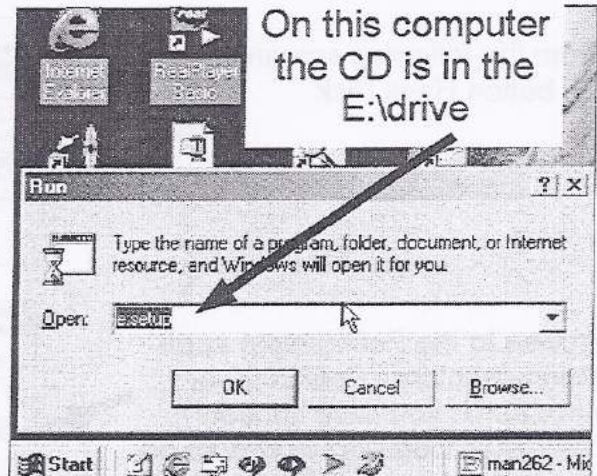## Topic: Installation of USB drivers for use with Model 302 USB data system

Install PeakSimple software from the installation CD or from a download from SRI's website http://www.srigc.com. The download is a zip file which must be un-zipped using PKZip or Win-Zip. From the Windows RUN command click on setup.exe in either the CD drive or the directory where the download was saved.

On this computer the CD is in the E:\drive

At the conclusion of the PeakSimple software installation, there will be 3 important files saved to the application directory. The application directory could be c:\Peak2000 or it could have another name which you specified during the installation. The 3 import files are named LL_USB.inf, LL-USB.sys and LL_USB2K.sys. These files are required to make Windows recognize the A/D board connected to the USB port of the computer.

Click on the Windows My Computer icon and then Control Panel and then on Add New Hardware.

Click on the NEXT button several times until you get to the screen which allows you to select hardware from a list.
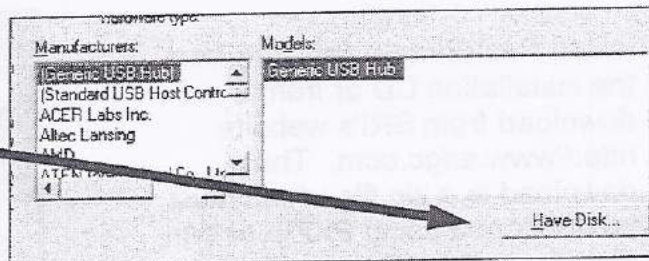
Scroll down the list until you can click on Universal Serial Bus controllers
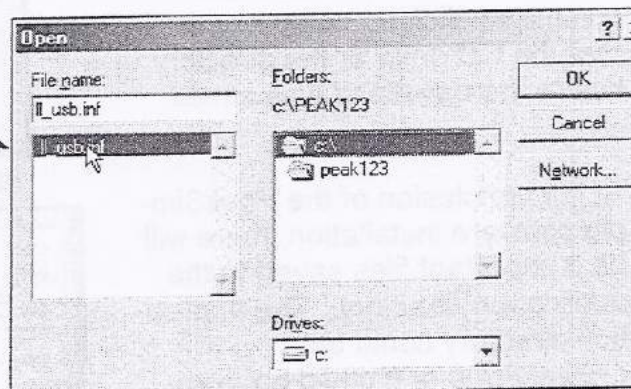
Chapter: PeakSimple

## Topic: Installation of USB drivers for use with Model 302 USB data system

From the following screen click on the button Have Disk

Browse to the PeakSimple application directory (Peak2000 or other name you have chosen). The Installation Wizard should be able to find the LL_USB.inf file.

When you click OK, the Wizard will verify that you want to copy files from the PeakSimple directory.
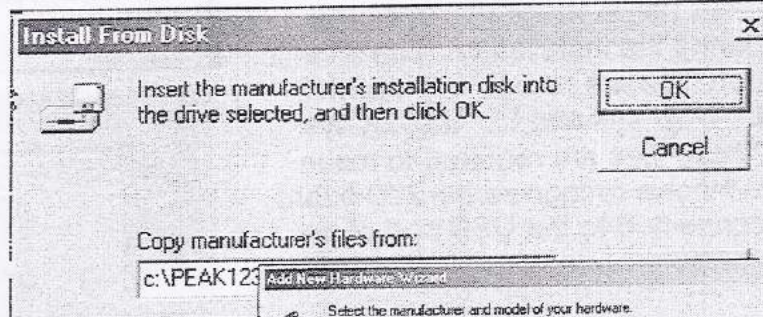
When you click OK again, the Wizard will confirm that the drivers are for Lawson Labs. Click Next and if the installation is successful, there will be one more screen where you can click the Finish button.
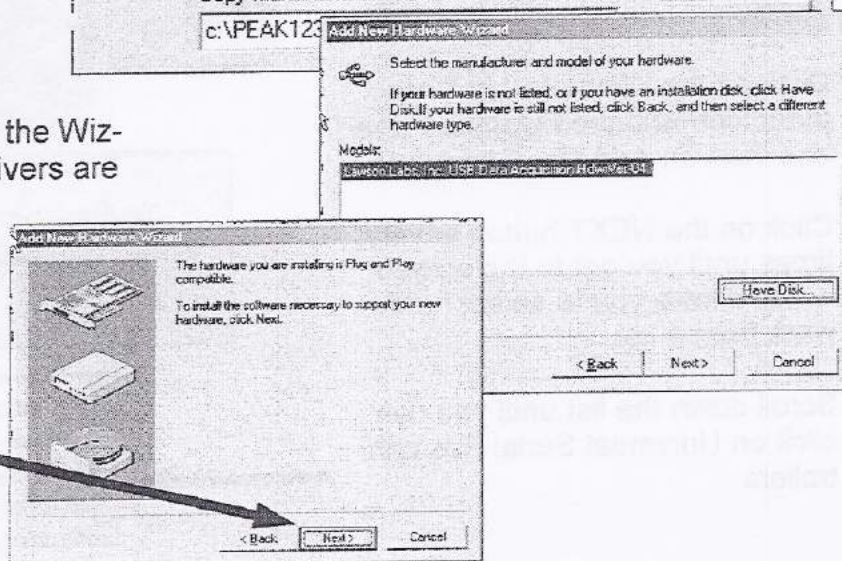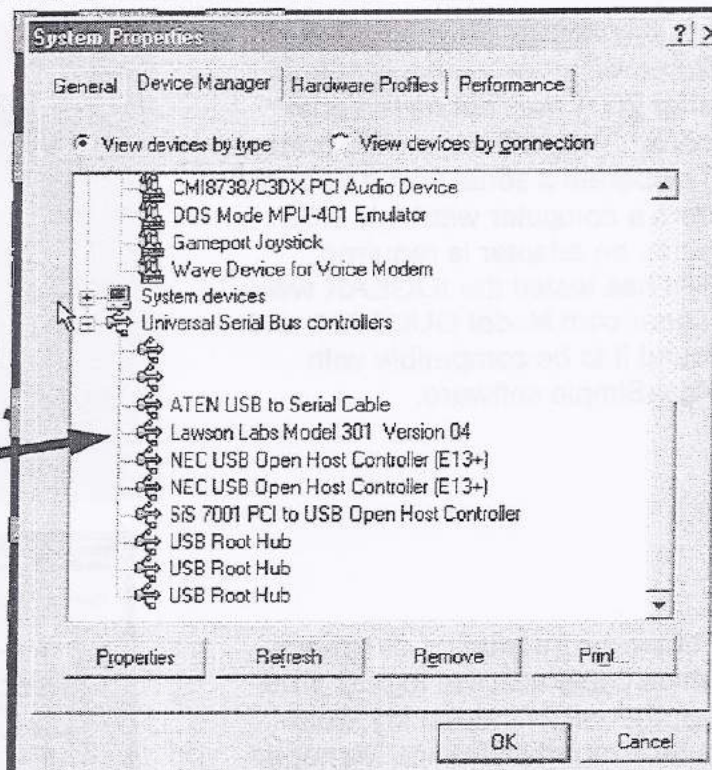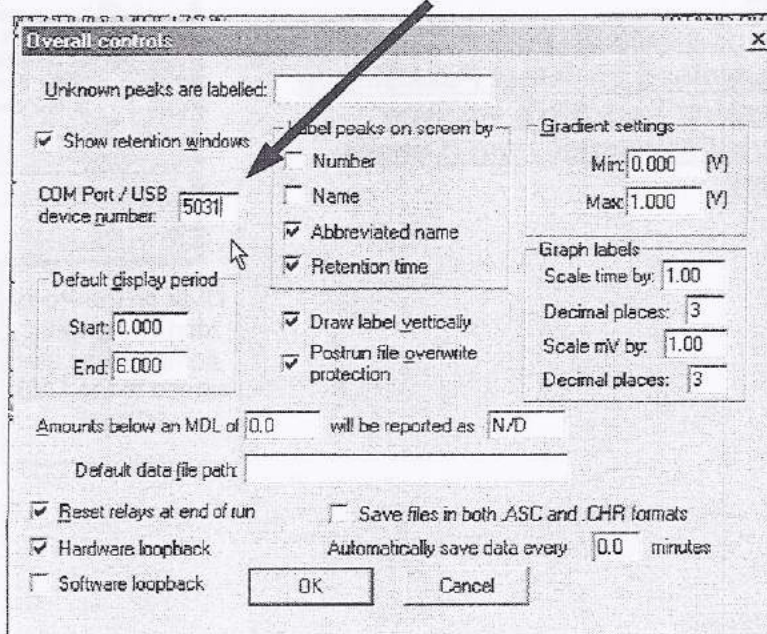
You must RESTART the COMPUTER before the drivers will work.

Once the computer has restarted right click on My Computer and examine the System/Device Manager Screen. If the USB drivers have been successfully installed, the Universal Serial Bus Controllers section will list Lawson Labs Model 301 version 04.

System Properties

General | Device Manager | Hardware Profiles | Performance

○ View devices by type ○ View devices by connection

- CMI8738/C3DX PCI Audio Device
- DOS Mode MPU-401 Emulator
- Gameport Joystick
- Wave Device for Voice Modem
- System devices
- Universal Serial Bus controllers
  - ATEN USB to Serial Cable
  - Lawson Labs Model 301 Version 04
  - NEC USB Open Host Controller (E13+)
  - NEC USB Open Host Controller (E13+)
  - SiS 7001 PCI to USB Open Host Controller
  - USB Root Hub
  - USB Root Hub
  - USB Root Hub

Properties | Refresh | Remove | Print...

OK | Cancel

You can now launch PeakSimple. Each USB data system has a unique USB device number which must be entered into the Edit\Overall window in PeakSimple. The USB device I.D. is listed on the back of each GC or stand-alone data system and will be a 4 digit number starting with 5 (5031, 5032, etc.) Once you enter the number Peak-Simple will attempt to wake up the data system. Don't forget to click the SaveAll icon so you don't have to re-enter the USB device I.D. again.

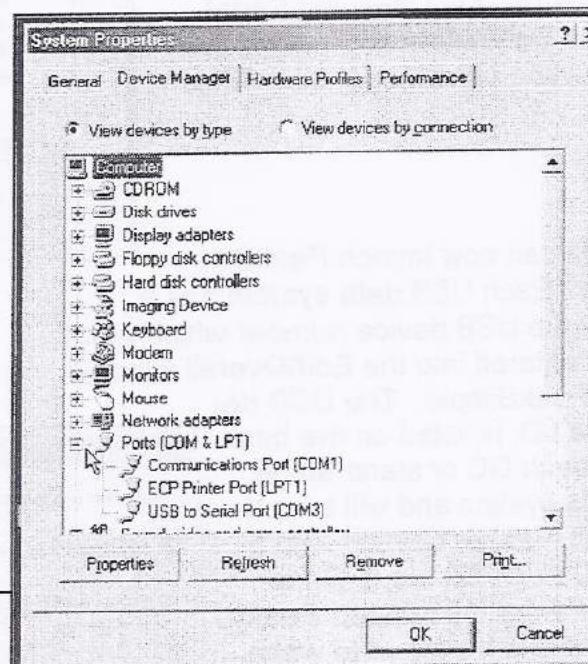Enter the USB device I.D. number in PeakSimple's Edit/Overall screen.

Overall controls

Unknown peaks are labelled: [   ]

☑ Show retention windows

Label peaks on screen by:
- ☐ Number
- ☐ Name
- ☑ Abbreviated name
- ☑ Retention time
- ☑ Draw label vertically
- ☑ Postrun file overwrite protection

COM Port / USB device number: [5031]

Default display period
Start: 0.000
End: 6.000

Gradient settings
Min: 0.000 [V]
Max: 1.000 [V]

Graph labels
Scale time by: 1.00
Decimal places: 3
Scale mV by: 1.00
Decimal places: 3

Amounts below an MDL of 0.0 will be reported as N/D

Default data file path: [   ]

☑ Reset relays at end of run
☑ Hardware loopback
☐ Software loopback

☐ Save files in both .ASC and .CHR formats
Automatically save data every 0.0 minutes

OK | Cancel

Some Windows computers sold
after 2001 may not have serial
ports. They will have USB ports.
To operate a serial port A/D board
from a computer with only USB
ports, an adapter is required.
SRI has tested the IOGEAR www.
iogear.com Model GUC232A and
found it to be compatible with
PeakSimple software.

This is the
adapter cable

Follow the installation directions
which come with the Iogear prod-
uct, then examine the My Com-
puter/Properties\Device Manager
screen and click on the Ports icon.
The serial port may be assigned to
Com 5 which is only supported by
version 2.83 and later PeakSim-
ple.

**Download the latest PeakSimple
version from SRI's website
http://www.srigc.com if neces-
sary**

Click on the Ports
icon to see what
comport was as-
signed to the USB
adapter

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Operation of Menu Bar Pull-Down Menus

All PeakSimple for Windows features may be accessed from pull-down menus. When you click on a menu bar item, a pull-down group menu will open to permit navigation to specific group features. These pull-down menus may also be opened by pressing the **<ALT>** key and the letter key corresponding to the **underlined** letter in the menu bar item name. For example, to open the **EDIT** menu press **<ALT>** and the letter "E" (This is not case sensitive).
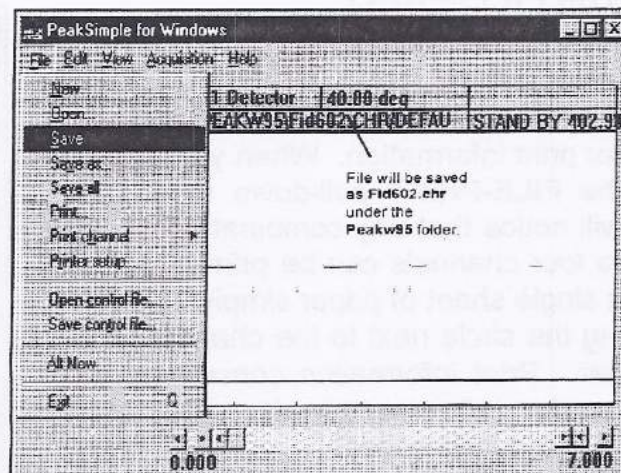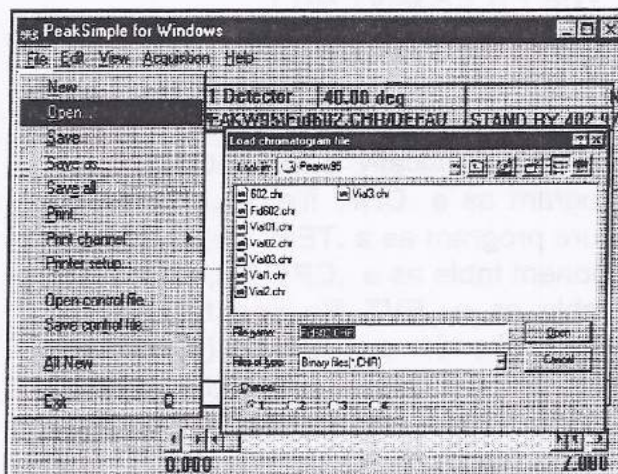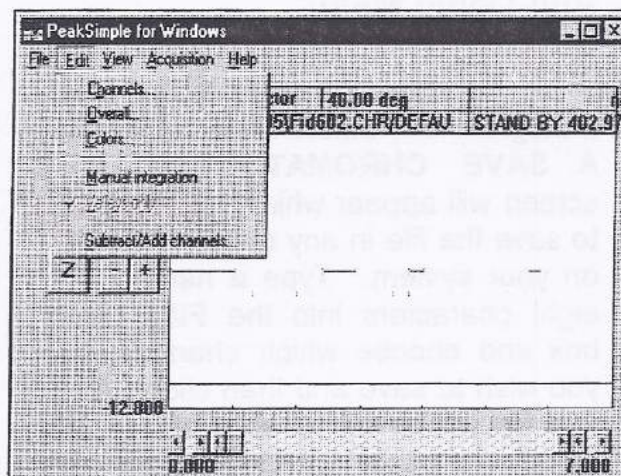
The FILE Pull-Down Menu

The **FILE-NEW** feature will clear the display of all active channels in the **Main** timebase without starting a new chromatographic run.

To open a previously saved chromatogram file, select **FILE-OPEN**. A **LOAD CHROMATOGRAM FILE** screen will appear which will allow you to select any file from any directory (folder) on your system. Choose the channel (1-4) in which you wish to display your saved chromatogram and than click **OPEN**.
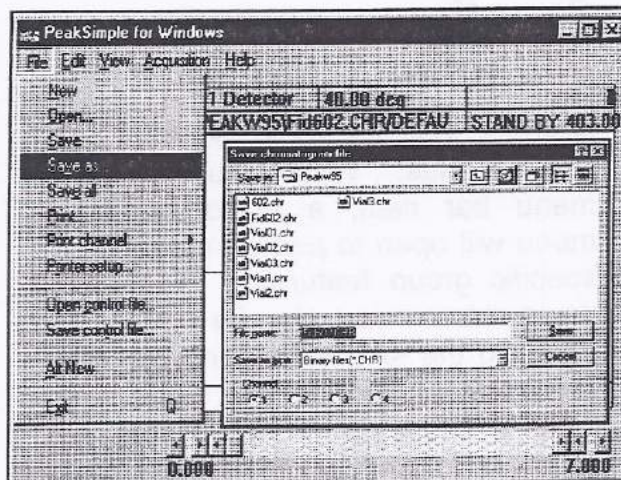
FILE SAVE

The **FILE-SAVE** feature saves the displayed chromatograms of all active channels. The name given to the file(s) is the same name that is displayed in the Data Boxes below the menu bar and will be given the default . **CHR** extension. This file name is editable by the user by changing information in the **EDIT-CHANNELS-POSTRUN** pull-down menu. See the **EDIT** section for more information.

The FILE-SAVE-AS
Pull-Down Menu

To save a newly created chromatogram file, select **FILE-SAVE AS**. A **SAVE CHROMATOGRAM FILE** screen will appear which will allow you to save the file in any directory (folder) on your system. Type a name up to eight characters into the **File** Name box and choose which channel (1-4) you wish to save and than click **SAVE**. The file will be saved as a **binary file** by default, with a **.CHR** extension. You may also select to save the file in AS-CII format with a **.ASC** extension.

The FILE-SAVE-ALL
Pull-Down Menu

The **FILE-SAVE-ALL** feature will automatically save your chromatogram as a **.CHR** file; your temperature program as a **.TEM** file; your component table as a **.CPT** file; your event table as a **.EVT** file and then saves them all under a control file (**.CON** file). **DEFAULT.CON** will be used if no other name for the **control file** is specified using the **SAVE-CONTROL FILE** feature. All print information is also saved when you save a **control file**.
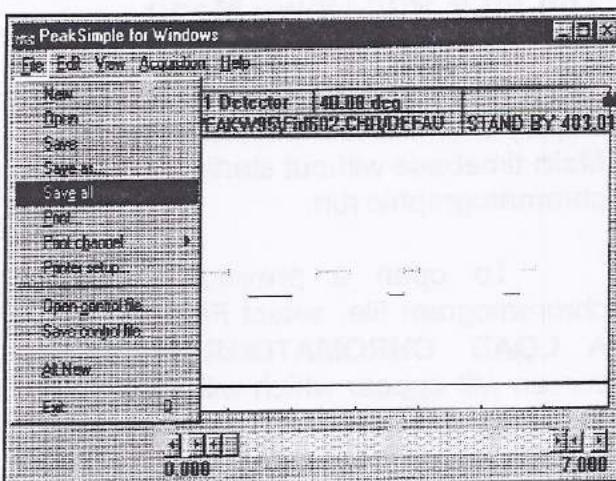
The FILE-PRINT
Pull-Down Menu

Numerous fields are available for print information. When you access the **FILE-PRINT** pull-down menu you will notice that any combination of one to four channels can be printed out on a single sheet of paper simply by marking the circle next to the channel number. Print information concerning the **header, chromatogram** and **report** can be easily edited.
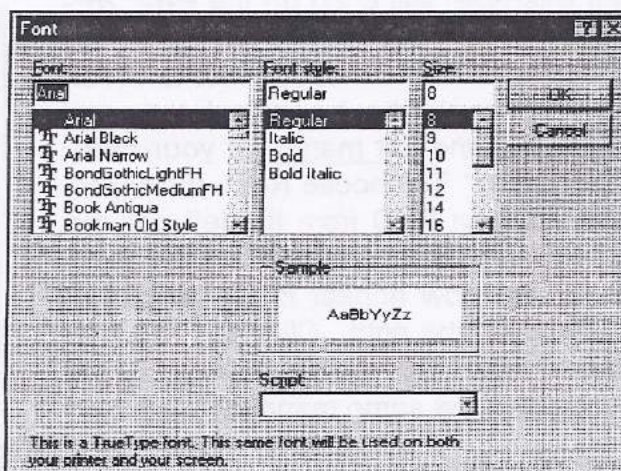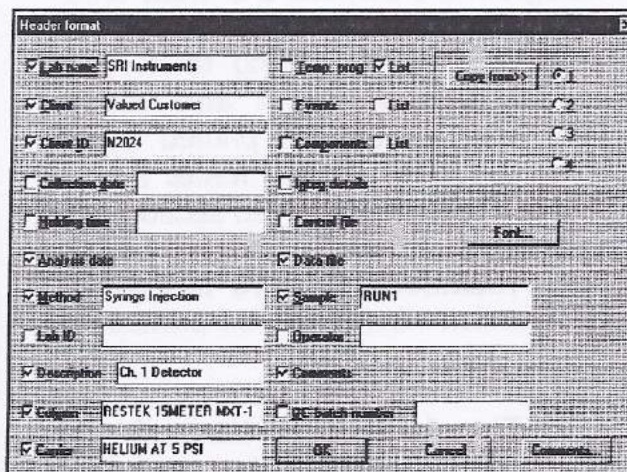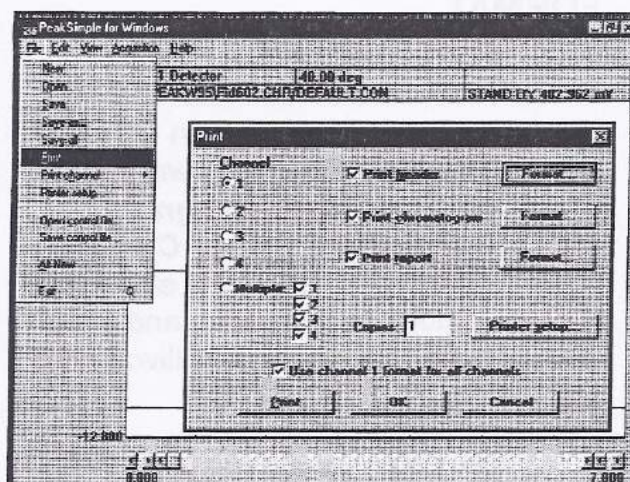
Pw-pg11.pub
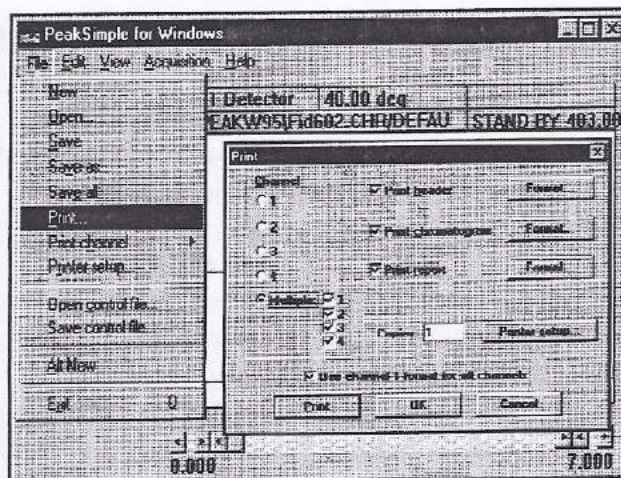
The FILE-PRINT
Pull-Down Menu (CHANNEL 1)

When you access the FILE-PRINT pull-down menu you will notice that you can select to print any combination of **multiple** channels by clicking on the circle next to the word **multiple**. You may also choose to print individual channels by clicking on the circle next to the desired channel. **Click on Channel 1** to edit the **Channel 1** information in the **Print Header, Print Chromatogram and Print Report Format** fields. Rather than enter unique information for all four channels, you may wish to check the **Use channel 1 format for all channels** box.

PRINT HEADER FORMAT

Clicking on the **Print Header FORMAT** button will allow you to customize the appearance of your printed chromatogram header. Input your **Laboratory name, Analysis method, Sample type, Column, etc** and check the box next to each field. **Analysis date** prints the date in your PC's BIOS.

Print out **Temperature Programs, Events and Components** file names by checking their boxes; or click on **List** to print the complete Temperature Program, Event Table or Component List. **Copy from:** selects which channel will provide the **List** information. Check the **Comments** box and click on **Comments...** to enter customized information about your analysis. You can change the **Font,** style and size of your printed text by clicking on the **Font** box. Select a size that will provide readable text while still leaving room for your chromatogram and report.

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                                        www.chromtech.net.au
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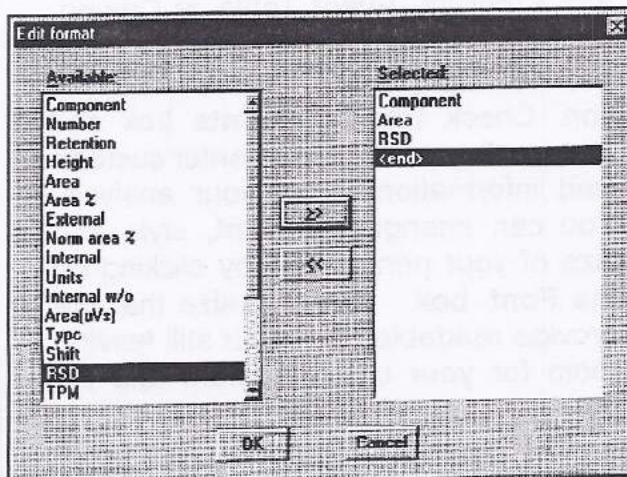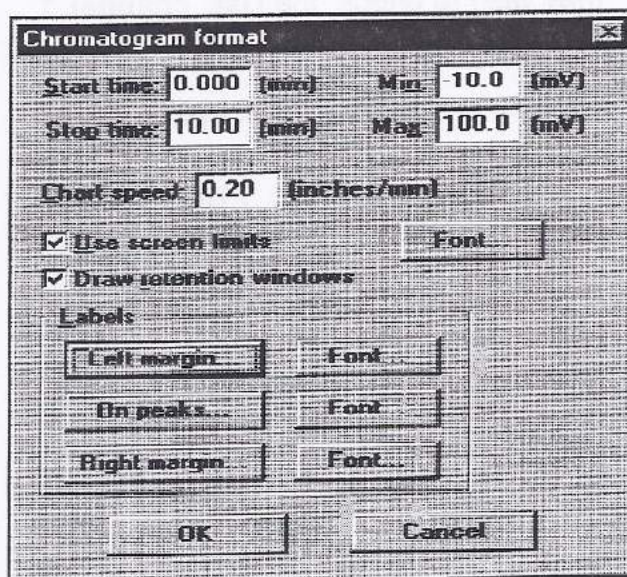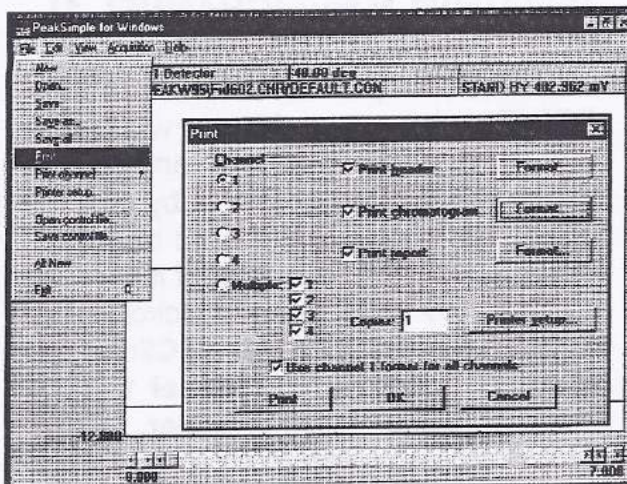
PRINT CHROMATOGRAM FORMAT

You can also edit the chromatogram print parameters when you access the **FILE-PRINT** pull-down menu. Check the **Print Chromatogram** box and select **Format**. The **Chromatogram format** screen allows editing of the chromatogram **Start time** and **Stop time** and the **Min** and **Max** millivolt levels.

The **Chart speed** setting will determine the size of the chromatogram section of your printout. A setting of **1.0 inches/minute** for a 5 minute chromatogram will produce a **5 inch** chromatogram print. You may need to experiment with this setting to fit your **header, chromatogram and report** information all on one printed page. When the **Use screen limits** box is checked only the displayed section of a chromatogram will be printed. The **Draw retention windows** box allows for retention windows to be printed as well.

The **Labels** section of the screen lets you select what useful information will be printed along the borders of the chromatogram, and above the peaks. Clicking on **Left margin**, for example, will bring up the **Edit format** screen which will allow you to select from a list of measurements which will automatically be calculated and printed in the left margin of your chromatogram. To choose **RSD**, for example, click on **RSD** from the left column and then click on the right arrows (>>). **RSD** will now appear in the **selected** column on the right. Click **OK** to close the window. Edit **On peaks** and **Right margin** in the same manner.

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PRINT REPORT FORMAT

A report may be printed along with your chromatogram to summarize component retention time, area counts or other data. Clicking on the **View** pull-down menu and selecting **Results** will show a preview of your report.

Click on the **Print Report** box and select **Format**. The **Report Format** screen will appear which will allow you to select from a list of measurements which will automatically be calculated and printed on the bottom of your chromatogram. To choose **AREA**, for example, click on **AREA** from the left column and then click on the right arrows (>>). **AREA** will now appear in the **Selected** column on the right.

Clicking on the box next to **Recognized peaks only** will place a check mark in the box and only those peaks which integrate properly within named retention windows will be printed in the report. Checking the **Peaks on printed chromatogram only** box will allow the report to show only those peaks defined by the **Chromatogram format– Start time and Stop time.** This feature allows you to set up your report to ignore all peaks that appear outside your window of interest.

Checking the **Undetected components also** box will report information about all named peaks even if no peak is present within the retention window. Checking **New page for report** will print all report information on a separate page. Click **OK** to close the **Report format** window. You may print out as many Chromatogram **Copies** as you need by entering a number in the **Copies** box and selecting **Print**.
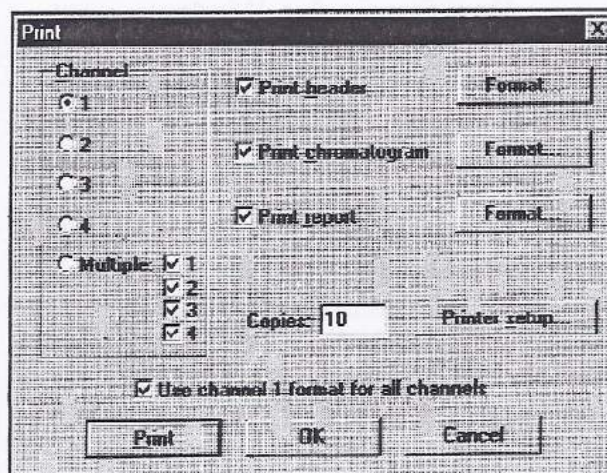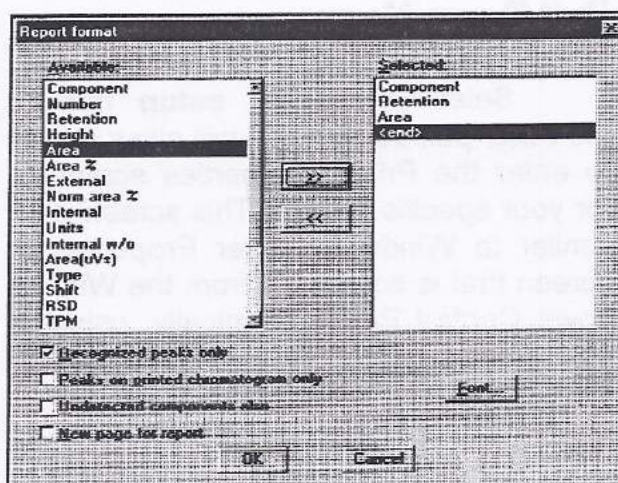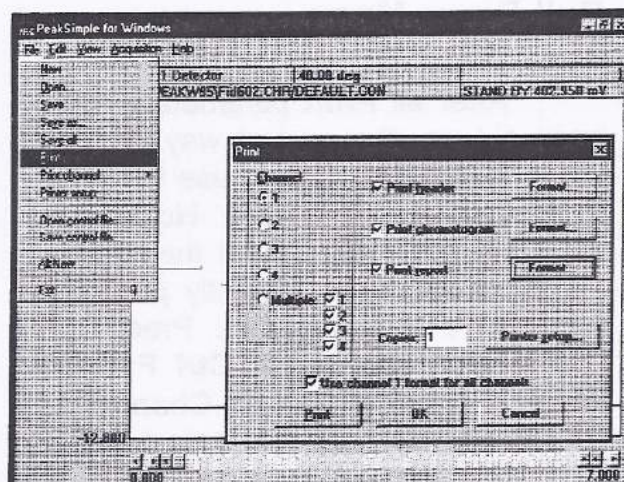
The FILE-PRINT CHANNEL Pull-Down Menu

After all **Print** parameters have been set up, the easiest way to print out a chromatogram is to use the **File-Print Channel** quick keys. Hold down the **Ctrl** (control) key and then press **F9** (function #9) to instantly print the **Channel 1** chromatogram. Press **Ctrl F10** to print **Channel 2, Ctrl F11** for **Channel 3** or **Ctrl F12** for **Channel 4**. Of course you may also select these commands from the pull-down menu.
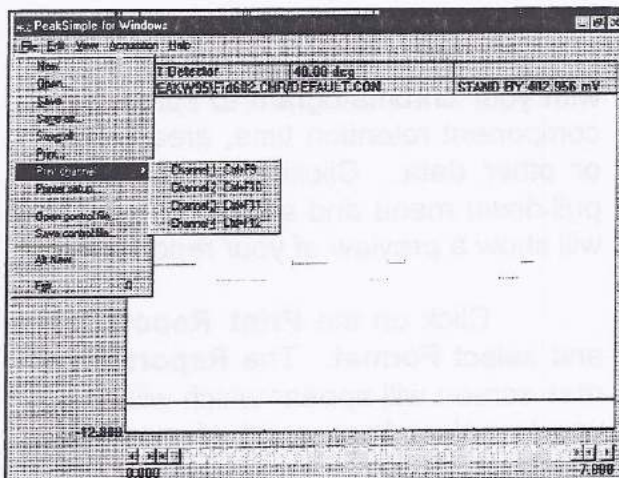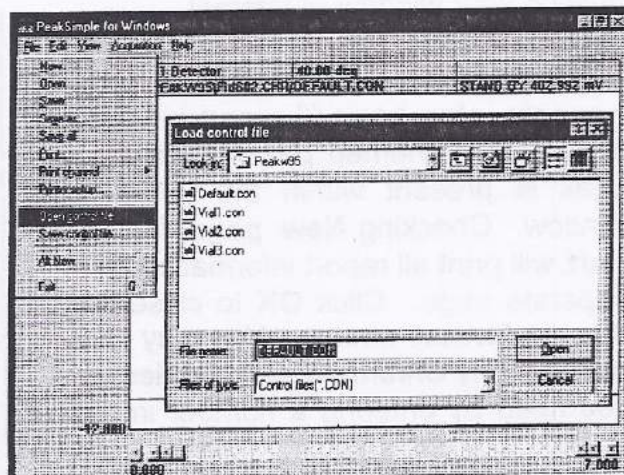
The FILE-PRINTER SETUP Pull-Down Menu

Selecting **Printer setup** from the **FILE** pull-down menu will allow you to enter the Printer Properties screen for your specific printer. This screen is similar to Windows Printer Properties screen that is accessible from the Windows Control Panel. Typically, using your printer default settings with **portrait** orientation will produce a visually appealing printout.
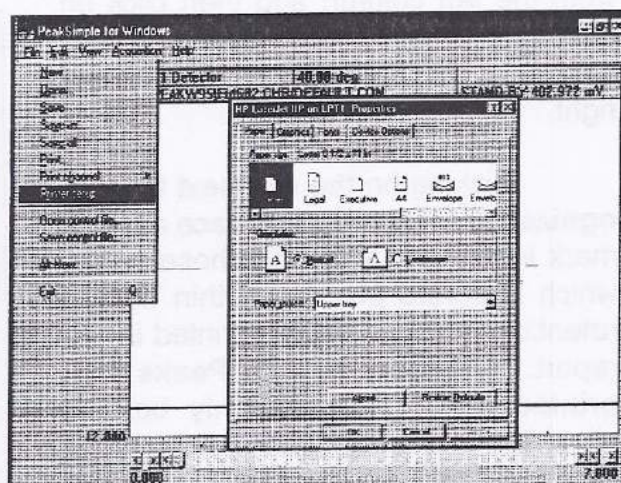
The FILE-OPEN CONTROL FILE Pull-Down Menu

PeakSimple for Windows uses **Control Files**, identified with the **.CON** extension, to save the operating settings of specific methods. To load a **Control File**, drop down the **FILE** menu and select **OPEN CONTROL FILE**. A window will open which will allow you to use standard Windows navigating tools to select from a list of **.CON** files, located on the **Drive** or **Directory** of your choice. Click on the desired **File Name** and then click **O.K.**

The FILE-SAVE CONTROL FILE Pull-Down Menu

Once you have set up all of the user-definable parameters within Peak-Simple for Windows that meet the requirements of your system and/or your specific analytical method, it is wise to save these settings for future use. PeakSimple uses **control files**, identified with a **.CON** extension, to save the operating settings of specific methods, this includes the event table, temperature program, component table, print information, calibration table, etc.

A **control file** is like a photo-copy of your operating settings that you can reload for use at any time. When using **control files**, you only need to set analysis parameters once and then save them using a descriptive file-name, followed by the **.CON** extension, (for example, **BTEX.CON**). To save the **control file**, drop down the **File** menu and select **Save control file**. Enter the name for your file in the **File name** box and click **O.K.**. If you want these current settings to be loaded by default each time you start PeakSimple, name the control file **Default.con**.
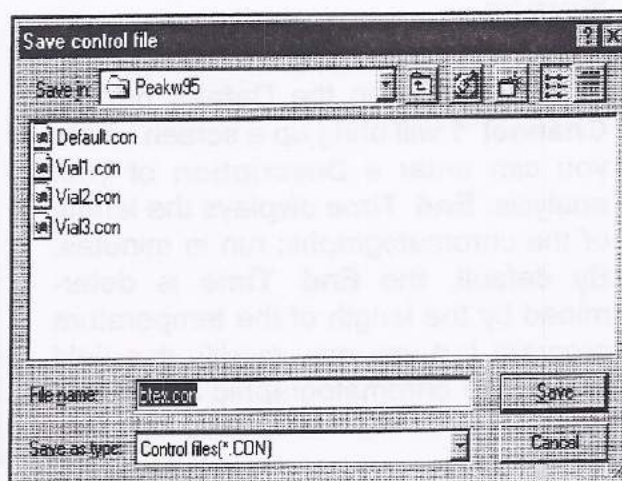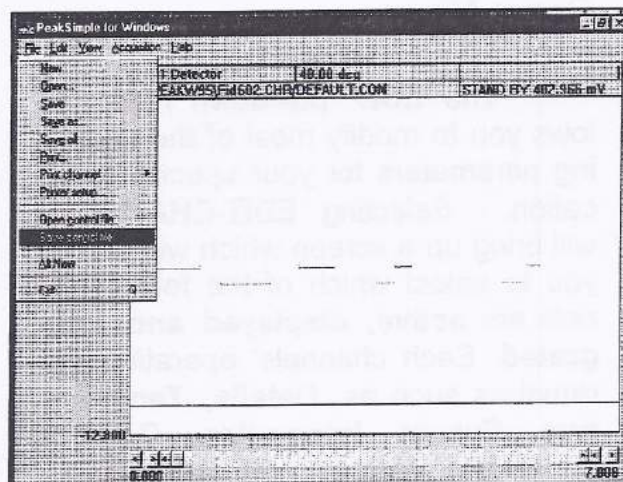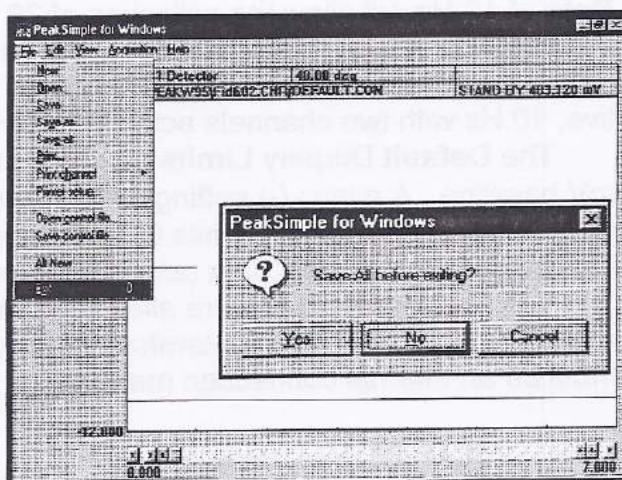
FILE-ALT NEW

The **FILE-ALT NEW** feature will clear the display of all active channels in the **Alternate** timebase without starting a new chromatographic run.

FILE-EXIT

Exits PeakSimple for Windows. Click **Yes** to save any changes made to your **control file** parameters.

The EDIT-CHANNELS Pull-Down Menu

The **EDIT** pull-down menu allows you to modify most of the operating parameters for your specific application. Selecting **EDIT-CHANNELS** will bring up a screen which will enable you to select which of the four channels are **active, displayed and integrated**. Each channels' operating parameters such as **Details, Temperature, Events, Integration, Components and Postrun** information can be easily modified.
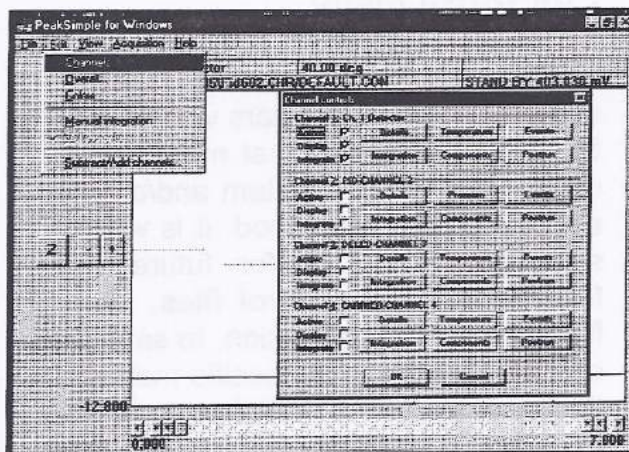
The EDIT-CHANNELS-DETAILS Screen

Clicking on the **Details** box for **Channel 1** will bring up a screen where you can enter a **Description** of your analysis. **End Time** displays the length of the chromatographic run in minutes. By default, the **End Time** is determined by the length of the temperature program but you may modify this field to end the chromatographic run at any time.
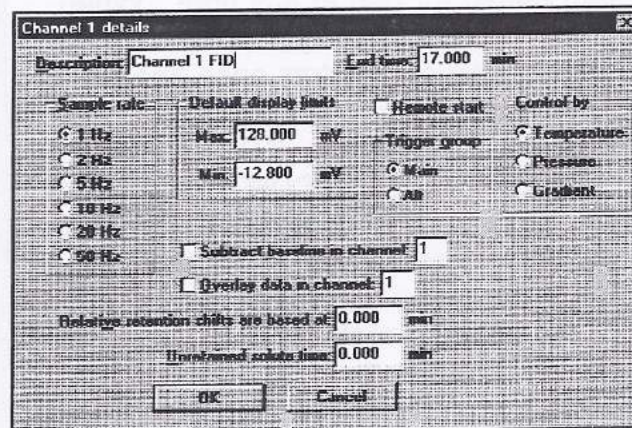
The **Sample Rate** should be set to a rate sufficient to ensure that 20 data samples are collected for each peak. For example: A **Sample Rate** of 1 Hz will allow the collection of 20 data points from a peak 20 seconds wide from base to base. And a **Sample Rate** of 10 Hz will allow the collection of 20 data points from a peak 2 seconds wide from base to base. The analog to digital converter is limited in its ability to sample high rates when many channels are active. The limits are: 50 Hz with one channel active, 10 Hz with two channels active and 5 Hz with three or four channels active.

The **Default Display Limits** can be adjusted to view data above and below the 0 mV baseline. A minus (-) setting for **minimum** will display negative going peaks. The ratio of **min./max.** display limits is maintained when you click on the Display minus and plus buttons in the main data acquisition screen.

The **Remote Start** feature allows the user to start a chromatographic run using an external signal such as a footswitch. Check the box to enable **Remote Start**. (There must be an internal connection made to the A/D board in order for this option to work.)

The EDIT-CHANNELS-DETAILS Screen (continued)

Trigger Group

The **Trigger Group** selection assigns the channel to the **Main** or **Alt** trigger group. The picture at right shows the **Channel 1 Details** screen with the **Main Trigger Group** selected.

Any **Channel** with the **Main** trigger group selected will start running when the SPACEBAR is pressed and end when the END key is pressed. Any **Channel** with the **Alt** trigger group selected will start running when the + (plus) key is pressed and end when the − (minus) key is pressed. When acquiring <u>four</u> detector signal inputs from <u>one</u> gas chromatograph; verify that all four channels' **Trigger Group** is set to **Main**. This ensures that all four channels are acquiring data synchronously by using the same timebase. If two channels of data are coming from an SRI gas chromatograph, and you also wish to acquire two channels from an external input device such as an HPLC, then select the **Alt** trigger group for channels 3 and 4. This allows for asynchronous data collection.

Subtract Baseline In Channel "X"

Checking **Channel 1's** box for **Subtract Baseline In Channel "X"**, where **"X"** is 1,2,3 or 4, will cause the chromatogram in **Channel 1** to subtract the baseline stored in **Channel "X"**, while running in <u>real-time</u>. Load the baseline to be subtracted into an <u>inactive</u> channel to ensure that the data is not deleted by the start of a new run on that channel. (Uncheck the **active** box, see **Edit-Channels**). Baseline subtraction can also be performed using PeakSimple's **Edit-Subtract/Add Channels** feature, however, this is not a real-time function, but a <u>post-run</u> function, done at the end of the chromatographic run.

Overlay Data In Channel "X"

Checking **Channel 1's** box for **Overlay Data In Channel "X"**, where **"X"** is 1,2,3 or 4, will overlay the data stored on **Channel "X"** onto **Channel 1** using contrasting colors. The channel selected for overlay can be either an active or inactive channel. When the overlay channel is active then the overlay will be seen in real-time.

Relative Retention Shifts Are Based At "X" Minutes

Relative Retention Shifts Are Based At "X" Minutes. Enter into this box the time, in minutes, that the sample is actually injected onto the column. This is done to ensure that relative retention times are correctly calculated. See the **EDIT-CHANNEL-COMPONENTS** section of this manual for more details.

The EDIT-CHANNELS-DETAILS Screen (continued)

Unretained Solute Time

If resolution has been selected to be printed in the chromatogram report, then a **Unretained Solute Time** value needs to be entered to ensure correct resolution calculations. Enter the number of minutes an **Unretained Solute** takes to pass through the column. This value is used in the determination of peak resolution statistics.

Control By

The A/D Board that is built into the SRI gas chromatograph includes two digital-to-analog converters or DACs. DAC1 is primarily used to control the <u>column oven #1</u> **temperature** ramp by introducing 10mV / °C to the oven heating circuit and is programmable by editing the **Channel 1–Temperature** control window. DAC2 is primarily used to control the <u>column oven #2</u> **temperature** ramp. Carrier gas E.P.C. pressure is also programmable by editing the **Channel 2–Temperature/Pressure** control window. The DACs may be used to control **Pressure** by following the procedure described below and then selecting **Pressure** in the **Control By** window of the **Edit-Channels-Details** screen.

To avoid startup difficulties, the Carrier E.P.C. is shipped disabled. To enable the use of the DACs to set up a **Pressure Program**, only a single wire needs to be moved inside the G.C.. Unplug the G.C. and remove the six screws which secure the bottom cover. Tilt the G.C. onto its back and remove the bottom cover. The A/D Board is green in color and is mounted on the right-hand side of the G.C. chassis. Locate the Green wire with a White stripe on the A/D Board. This is the

Carrier Program wire. Normally this wire is connected to a ground (GD) screw terminal. Unscrew the **Carrier Program** wire and connect it to the temperature/pressure #2 (TP2) screw terminal also on the A/D Board. Re-attach the bottom cover, connect power and re-establish communication between the G.C. and the computer. Select **Pressure** in the **Control By** window of the **Edit-Channels-Channel 2-Details** screen. A pressure program ramp set up in **Channel 2** will now control the Carrier Gas E.P.C. pressure by introducing 10mV for every P.S.I.. Turn the Carrier 1 **Local Setpoint** to zero. This is necessary since the Local setpoint is added to the programmed E.P.C. input in determining the Carrier 1 **total setpoint**.
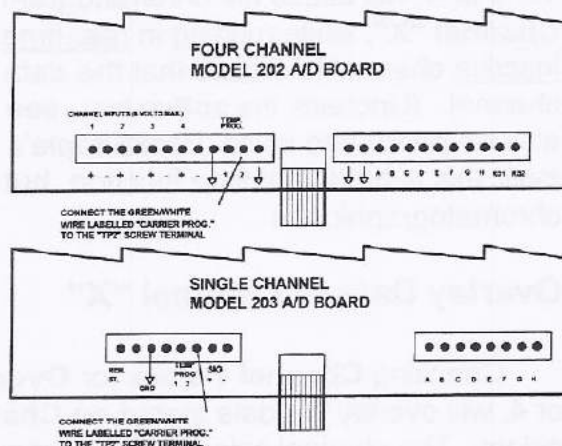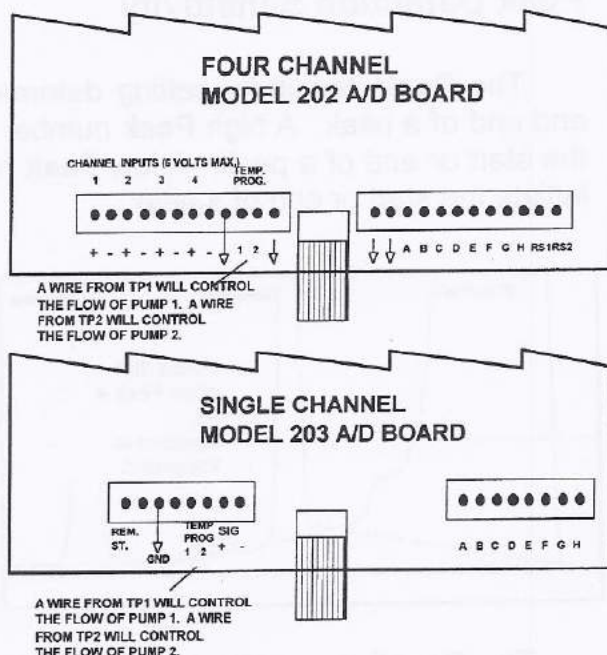
The EDIT-CHANNELS-DETAILS Screen (continued)

Setting Up Gradients for Liquid Chromatography

Data System users may wish to use the A/D Board DACs for setting up an HPLC solvent gradient. PeakSimple for Windows allows the user to control the flow of two pumps, provided they operate from a zero to five volt (0-5V) ramp input.

To operate the pumps, several internal connections must be made between the HPLC and the Data System. Unplug the Data System and remove the two screws which secure the top cover. Route the Pump A and Pump B control wires from the HPLC to the Data System and connect the Pump A control wire to TP1 and the Pump B control wire to TP2. Re-attach the top cover, connect power and re-establish communication between the Data System and the computer.

Set up the **Gradient** ramp on **channel one** (TP1) to control the flow of Pump A into the system (10mV / %) and the **Gradient** ramp on **channel two** (TP2) to control the flow of Pump B. Modifying the **Gradient** ramp program on **Channel 1** to rise from 10% to 90% will automatically create a **Gradient** ramp program on **Channel 2** that decreases proportionately from 90% to 10%.

Gradient Limits Zero and **Span** may be scaled in the **Edit-Overall** screen to account for any offsets. PeakSimple allows for a voltage offset and scaling factor in these fields to calibrate the voltage output to match the pump's requirements.

FOUR CHANNEL MODEL 202 A/D BOARD

CHANNEL INPUTS (5 VOLTS MAX.)

A WIRE FROM TP1 WILL CONTROL THE FLOW OF PUMP 1. A WIRE FROM TP2 WILL CONTROL THE FLOW OF PUMP 2.

SINGLE CHANNEL MODEL 203 A/D BOARD

A WIRE FROM TP1 WILL CONTROL THE FLOW OF PUMP 1. A WIRE FROM TP2 WILL CONTROL THE FLOW OF PUMP 2.

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The EDIT-CHANNELS-INTEGRATION Screen

PeakSimple for Windows allows you to define specific integration parameters necessary for the proper analysis of your sample data, such as peak and baseline sensitivity and area reject. Any of the **Integration** parameters described below may be modified either before or after data collection. Pressing the **ENTER** key will update the report and the results of the chromatogram currently being displayed.
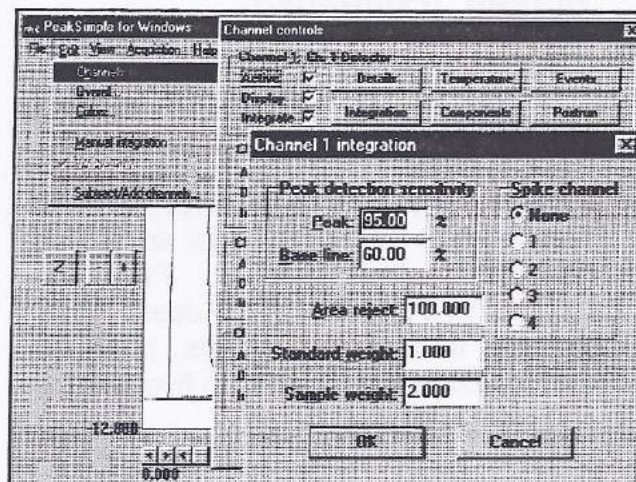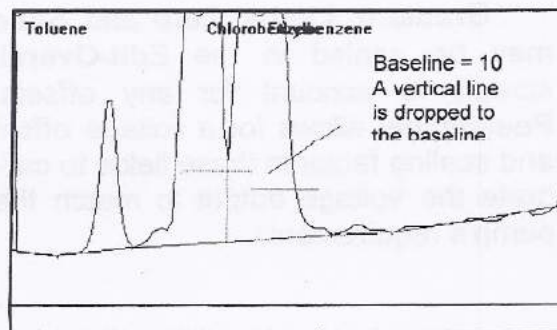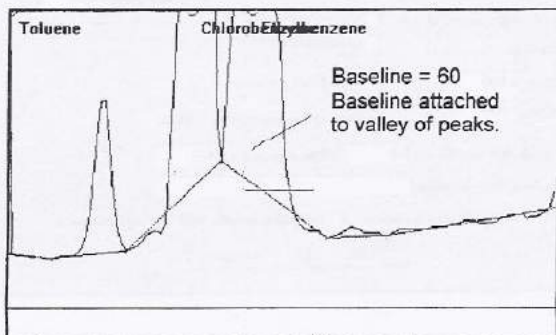
Peak Detection Sensitivity

The **Peak** sensitivity setting determines how PeakSimple detects the beginning and end of a peak. A high **Peak** number requires only a small slope change to initiate the start or end of a peak. A low **Peak** number requires a very large slope change to initiate the start or end of a peak.
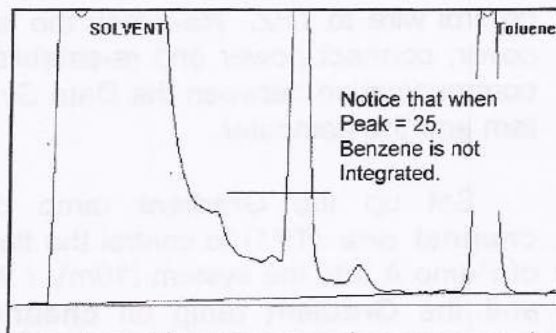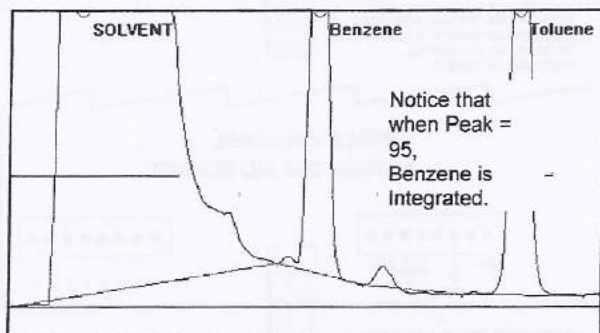
The **Baseline** sensitivity setting determines how PeakSimple attaches the baseline to the data line. The larger the **Baseline** number; the more likely PeakSimple will draw the baseline to a valley between two peaks. The smaller the **Baseline** number; the more likely PeakSimple will drop a vertical line from a valley to a horizontally constructed baseline below the peak.

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The EDIT-CHANNELS-INTEGRATION Screen (continued)

Area Reject

If a chromatogram contains peaks whose area counts fall below the threshold defined by the **Area Reject** for that channel, the peak will be ignored and no integration will occur. If the peak area is of interest, you can lower the **Area Reject** value until the peak in question is integrated. Integrated peaks are marked with a circle at the top of the peak.

Standard Weight

PeakSimple for Windows determines the internal or external standard results by the ratio of the STANDARD divided by the SAMPLE.

The **Standard Weight** setting may be changed to adjust the channel's quantification, affecting internal or external peak results by the factor entered. For instance: A setting of 2.000 will double the weight of the <u>standard</u> thereby doubling the internal or external standard results. (Increased to 20.000 in the example shown.)

Sample Weight

The **Sample Weight** setting may also be changed to adjust the channel's quantification, affecting internal or external peak results by the factor entered. For instance: A setting of 2.000 will double the weight of the <u>sample</u> thereby halving the internal or external standard results. (Decreased to 5.000 in the example shown.)

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Channel 1 integration
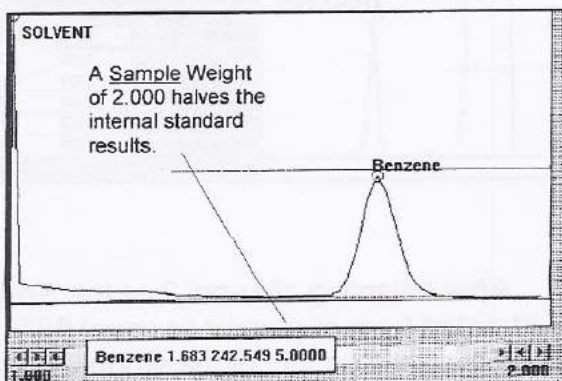
Peak detection sensitivity
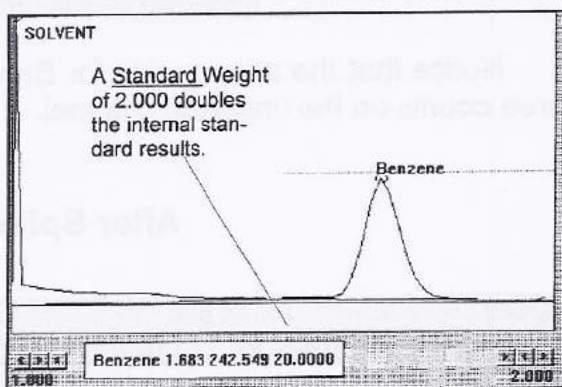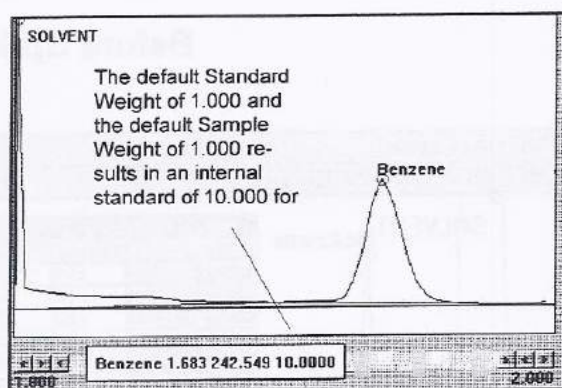
Peak: 95.00 %

Base line: 60.00 %

Area reject: 100.000

Standard weight: 1.000

Sample weight: 1.000

Spike channel
- None
- 1
- 2
- 3
- 4

OK   Cancel

SOLVENT

The default Standard Weight of 1.000 and the default Sample Weight of 1.000 results in an internal standard of 10.000 for

Benzene

Benzene 1.683 242.549 10.0000

1.000   2.000

SOLVENT

A <u>Standard</u> Weight of 2.000 doubles the internal standard results.

Benzene

Benzene 1.683 242.549 20.0000

1.000   2.000

SOLVENT

A <u>Sample</u> Weight of 2.000 halves the internal standard results.
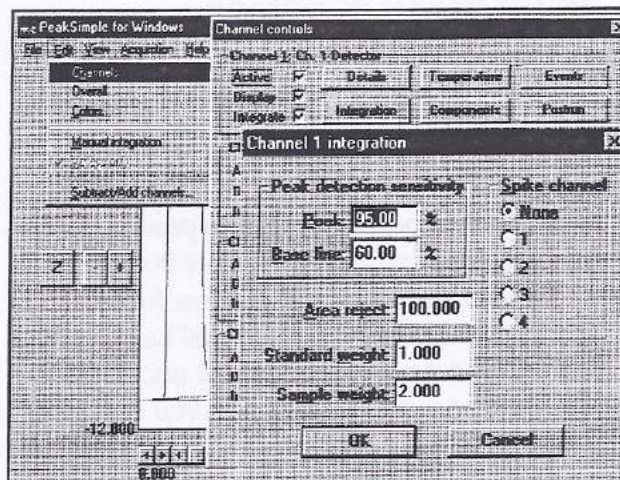
Benzene

Benzene 1.683 242.549 5.0000

1.000   2.000

The EDIT-CHANNELS-INTEGRATION Screen (continued)

Spike Channel

Another feature of PeakSimple for Windows allows you to display the results of a matrix **Spike Channel** subtraction. The example shown below demonstrates the peak area counts of a unspiked channel being subtracted from the area counts of a spiked channel.
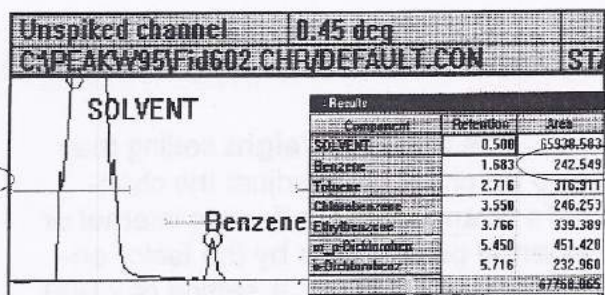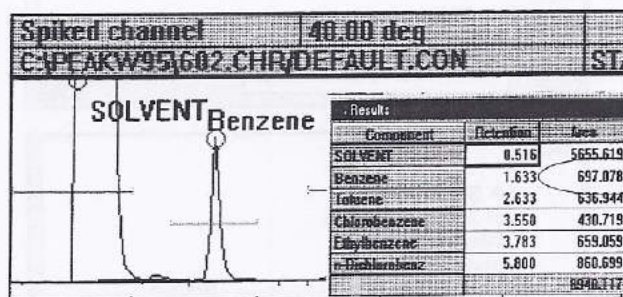

Before Spike Channel Subtraction
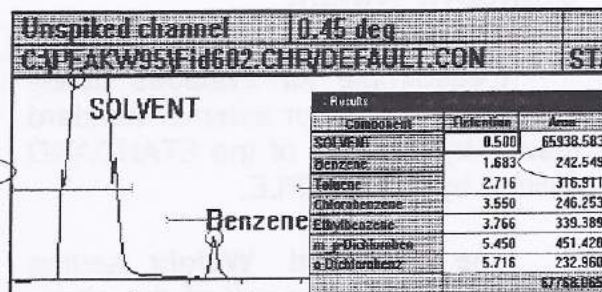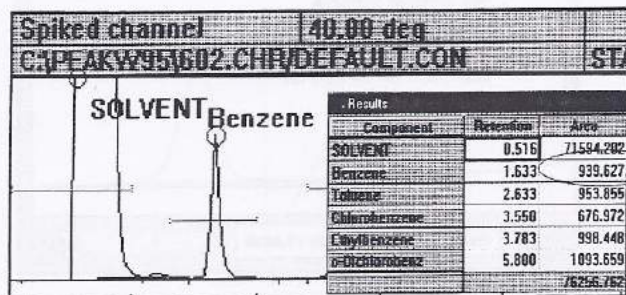
Notice that the area counts for Benzene are 939 on the spiked channel, and 242 area counts on the unspiked channel.

After Spike Channel Subtraction

After selecting channel 2 as the **Spike Channel**, the area counts for channel 2 are subtracted from channel 1 to equal 697, (939 – 242 = 697). The difference of 697 indicates the area counts of the amount of sample spiked into channel 1.

The EDIT-CHANNELS-TEMPERATURE Screen

PeakSimple for Windows features temperature-programming of the G.C 's column oven(s). Access the **Edit-Channel 1-Temperature screen** to specify the temperature parameters to be used during the analytical run. The temperature program is capable of executing an unlimited number of temperature ramp and hold periods during the analysis as well as maintaining a single temperature throughout the run for isothermal operation.
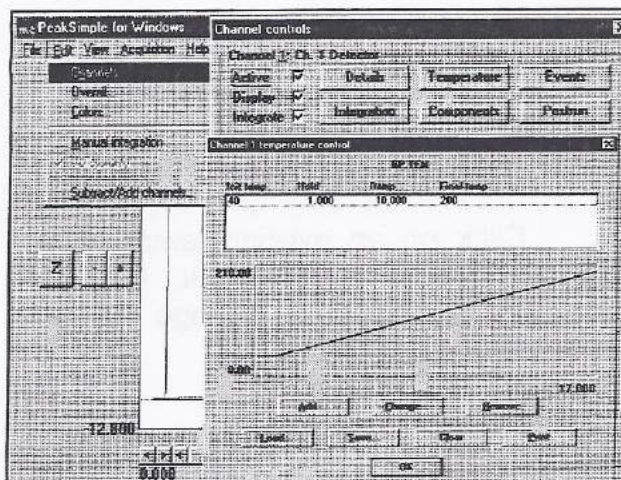

The Temperature Segment Details Screen

The **Add** Button

Click on the **Add** button from a blank **Channel 1 temperature control** window to create a new temperature program for Column oven #1. (Use the **Edit-Channel 2-Temperature** screen for controlling column oven #2). Type in the required data in the following fields; **Initial temperature**, the **Hold** period in minutes, the **Ramp** rate in °C / min, and the final **Temperature**, or the duration of the Ramp.

The length of the run is automatically calculated by PeakSimple based on the information provided in these fields, and is also displayed in the **Edit-Channels-Details End Time** field. Additional ramp segments may be added by clicking the **Add** button again.

In isothermal operation, the **Initial** and the final temperature are the same, so a **Ramp** rate of 0.000 is entered. The **Hold** period determines the length of the analytical run.
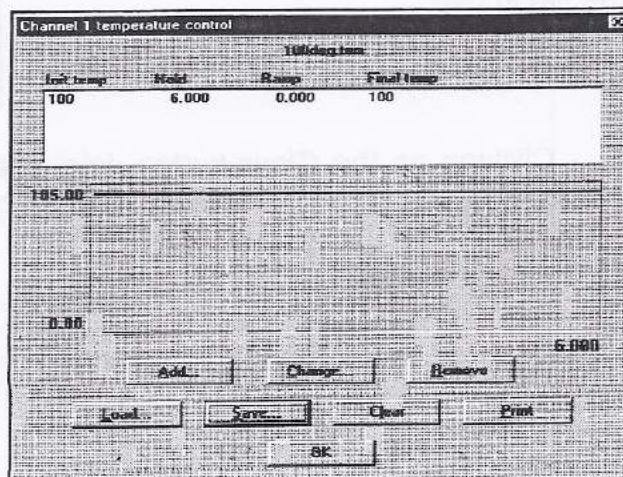
The EDIT-CHANNELS-TEMPERATURE Screen (continued)

The Change Button

Click on an existing temperature program segment to select it. Click on the **Change** button to change the parameters of the segment.
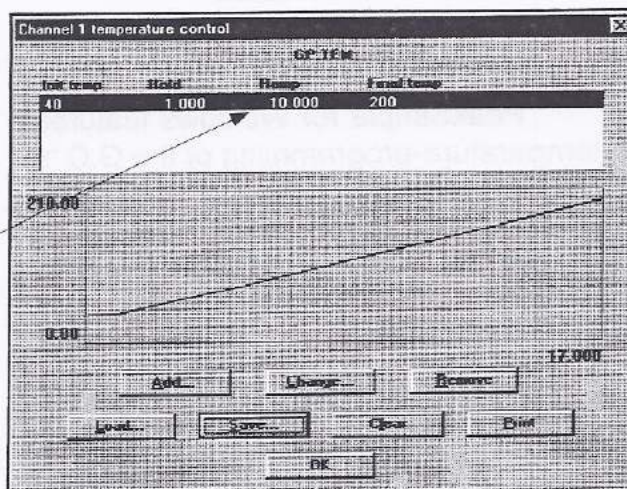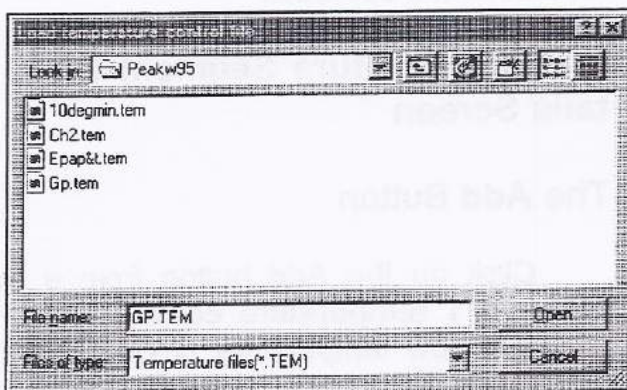
The Remove Button

Click on the **Remove** button to remove the segment from the current program.

The Load Button

Click on the **Load** button to load an existing temperature control file, designated with the **.TEM** file extension.

The Save Button

Click on the **Save** button to save a new temperature control file, or to update an existing one. Remember to use the **.TEM** extension when naming the temperature control file. The saved file name appears at the top of the temperature control window indicating the file in use.
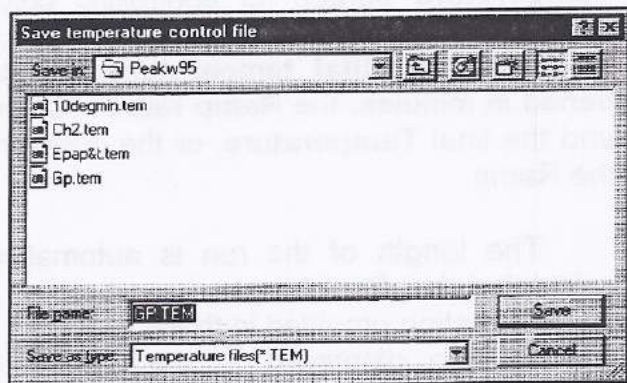
The Clear Button

Clicking on the **Clear** button deletes all temperature data from the temperature control window. The temperature program name is also removed.
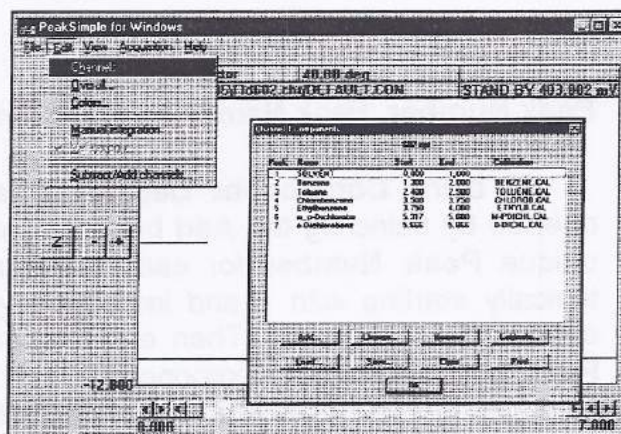
The Print Button

Clicking on the **Print** button sends the file data and temperature program profile to the printer.

Pw-pg25.pub

The EDIT-CHANNELS-COMPONENTS Screen

PeakSimple for Windows can identify and quantify sample components through the use of a component table. The component table enables PeakSimple to recognize each peak by its retention time and compare the area counts against the calibration curve to produce actual concentration data. The user can edit the component table for each channel by accessing the **Edit-Channels-Components** screen.

When a component table is loaded, the table will show each component by its peak number, peak name, the start time for the retention window, the stop time for the retention window, and the associated calibration file name. Different component tables may be used for each active channel and any component table can be saved as a component file for future use. Component files are designated with a **.CPT** extension. The component file-name appears at the top of the **Components** screen.

COMPONENT DETAILS

Select **Add** to add a new component to a blank or existing component table. The Component Details screen will open allowing the user to input specific peak parameters. As a minimum, enter the **Peak Number**, **Peak Name**, **Start** time and **End** time. Other optional parameters are the **Expected** peak time, the concentration **Units** to be reported, any **Internal Standard** or **Reference** peak information, peaks measured by **Area** or **Height**, handling of **Multiple Peaks**, the **Multiplication Factor** and **Alarm** parameters.

The EDIT-CHANNELS-COMPONENTS-DETAILS Screen (continued)

Peak Number, Peak Name, Start and End

A blank **Component Details** screen is opened by selecting the **Add** button. Enter a unique **Peak Number** for each component, typically starting with **1** and incrementing for each additional peak. Then enter a unique **Peak Name** for each component. **Start** and **End** define the beginning and ending of the retention windows, which is used to identify the peak. The width of the retention window should be set wide enough so that small fluctuations in the peak's retention time will still allow for proper integration.

Internal Standard and Units

Internal Standard calculations are used to correct for injection size variations, or to compensate for changes in detector sensitivity. An internal standard peak is added to the sample prior to injection at a known concentration. The internal standard peak is calculated just like any other peak using a calibration curve, typically a single point calibration. The known concentration of the internal standard peak is entered into the **Internal Standard** dialog box of the **Component Details** screen. In the example shown below, Benzene has been chosen as the internal standard peak. The known concentration of Benzene is entered as **100**, and **ppm** is entered in the **Units** dialog box. When a chromatogram is integrated and a report is produced, the external calculation yields a result which is the **peak area x calibration factor** (slope of the calibration curve) = **external standard result**.

The **internal standard** calculation yields a result which is the **external result times the ratio of the known concentration of the internal standard peak divided by the external result for the internal standard peak**. As shown in the example to the right, note that while the external result for Benzene yields 104.95, the internal result yields exactly 100 (the known concentration) as a result of the calculation **104.95 x 100/104.95**. In the same way, the internal result for every analyte peak which is referenced to Benzene is calculated as **external result x 100/104.95 = internal standard result**.

Component	Retention	Area	External	Internal	Units
SOLVENT	0.516	71594.202	0.00	0.0000	%
Benzene	1.633	939.027	104.95	100.0000	ppm
Toluene	2.633	953.855	106.73	106.7319	ppm
Chlorobenzene	3.550	676.972	72.12	72.1215	ppm
Ethylbenzene	3.783	998.448	112.31	112.3059	ppm
m-p Dichlorobenzene	5.800	1093.659	124.21	124.2074	ppm
o-Dichlorobenzene	6.150	536.767	54.60	54.5959	ppm
		76753.529	574.92	569.9626	

Internal Standard Peak

PeakSimple allows any peak to be referenced to any other peak for internal standard calculations. Typically all analyte peaks will be referenced against a single **Internal Standard Peak** (Benzene [peak #2] in the example shown below). To reference other peaks to Benzene, the number **2** must be entered in the **Component Details** screen dialog box labeled **Internal Standard Peak** for each analyte peak. Notice that the **Results** screen, (**View-Results**), will reflect the new value for all the peaks' internal results.

Reference Peak

A **Reference Peak** is used to shift the retention windows of other peaks. In the example below, ethylbenzene eluted prior to its retention window so therefore it was not integrated. By entering a value of **4** in the **Reference Peak** box, ethylbenzene's retention windows are referenced to chlorobenzene, [peak #4]. Ethylbenzene's retention window is then shifted by a percentage equivalent to chlorobenzene's distance from the middle of _its_ retention window. This shift in the ethylbenzene retention window allows ethylbenzene to be integrated.
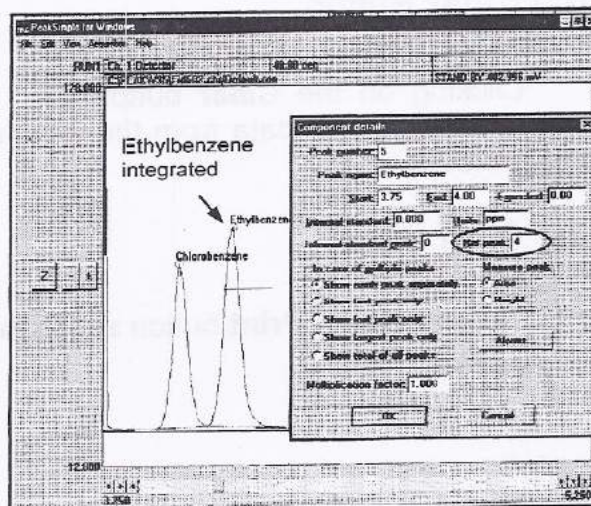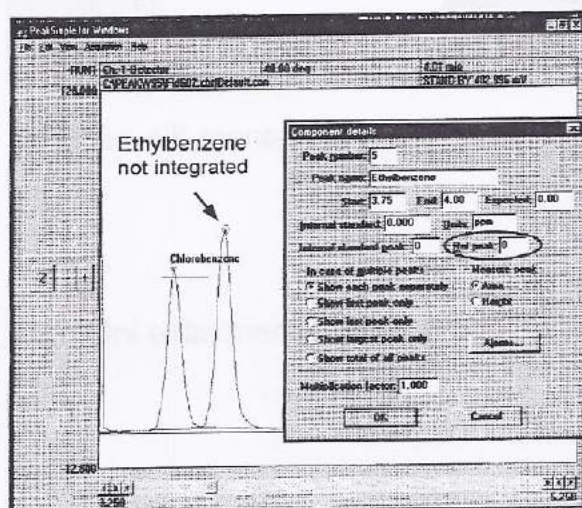
The EDIT-CHANNELS-COMPONENTS Screen (continued)

The **Change** Button

Click on an existing component to select it. Click on the **Change** button to change the parameters of the component.

The **Remove** Button

Click on the **Remove** button to remove the component from the component table.

The **Load** Button

Click on the **Load** button to load an existing component file, designated with the **.CPT** file extension.

The **Save** Button

Click on the **Save** button to save a new component file, or to update an existing one. Remember to always use the **.CPT** extension when naming the component file. The saved file name appears at the top of the components window indicating the file in use.

The **Clear** Button

Clicking on the **Clear** button deletes all component data from the component window. The component file name is also removed.

The **Print** Button

Clicking on the **Print** button sends the file data and the component table information to the printer.
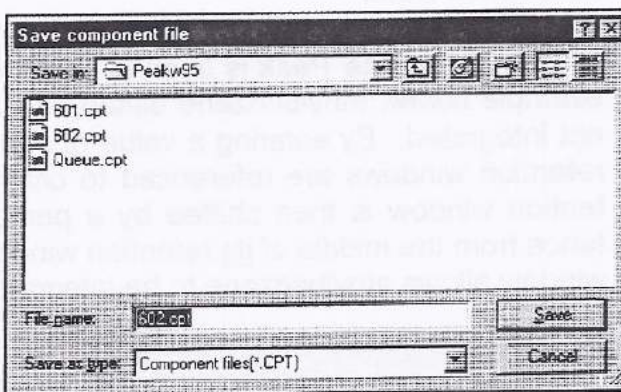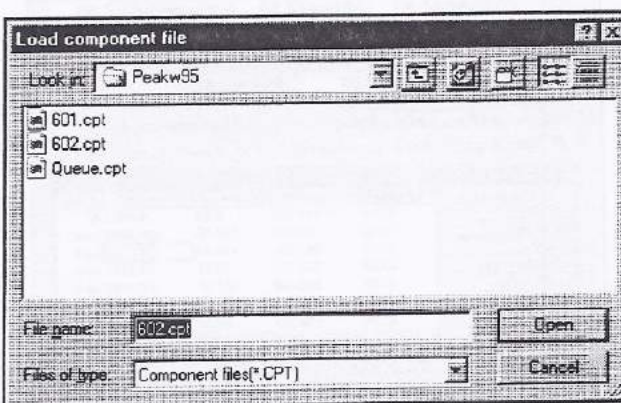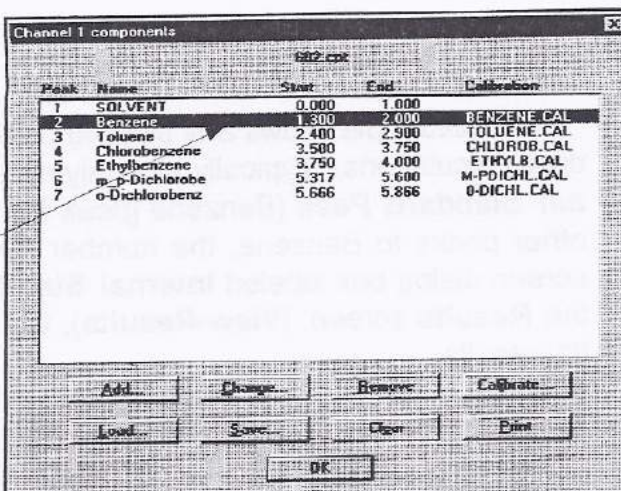
The EDIT-CHANNELS-COMPONENTS Screen (continued)

The Calibrate Button

After creating a component table, each component in the table will need to be calibrated. This allows PeakSimple for Windows to not only identify each analyte peak, but also to quantify each peak using a calibration curve. The calibration curve is calculated from user-generated results obtained at several different concentrations that span the expected range to be encountered in actual samples.

Inject a standard containing a known concentration of the component you wish to calibrate. Use a concentration higher than what you would expect to encounter in your analyses. Another few samples should be run at lower levels, using precise dilutions of your standard. Make note of the area counts or peak height at each concentration or use the shortcut method described in the next section.

The Calibration Window

In the **Edit-Channels-Components** screen, highlight the component to be calibrated and select **Calibrate**. If this is the first time calibrating a component, an error message will appear which says "**Not enough data points**". This is simply a warning to inform you that PeakSimple currently does not have enough data points for the calibration method in use. Once enough data is entered for the calibration curve, this message will no longer appear. Click **OK** to bypass the error message and continue to the calibration window.
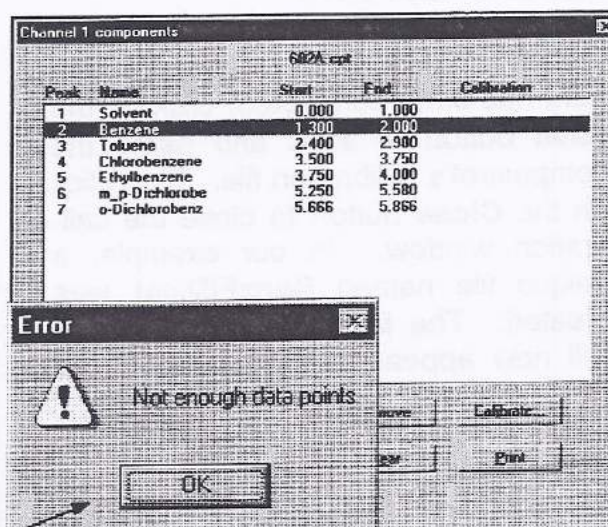
The **Calibration** window will open and allow you to enter the raw data that you previously obtained. In the example shown, data is entered into the table in the upper left corner of the calibration screen, beginning with the lowest concentration and ending with the highest concentration. If you wish to enter the data in descending order, check the **Show list in descending order** box. When entering data into the table, first enter the concentration injected, then the area count or peak height obtained.
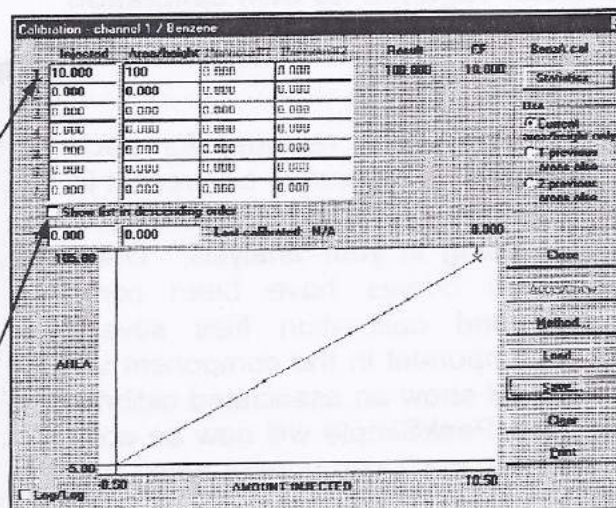
The Calibrate Button (continued)

As data is entered for each concentration, a data point will be added to the calibration curve displayed in the lower section of the window. You may use as many as seven concentration levels for your calibration curve. In the fictitious example to the right, a Benzene standard was injected in concentrations of 10 ppm, 50 ppm, and 100 ppm. The area counts from the FID detector were 100, 500 and 900, respectively. Notice the three corresponding data points on the newly created calibration curve.
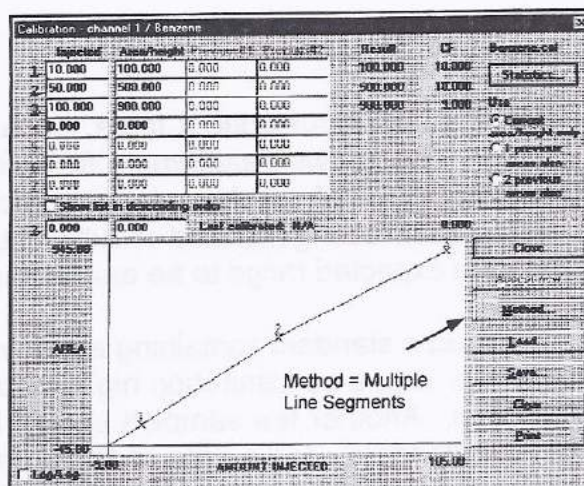
When calibration for each component has been completed, click on the **Save** button to save and name the component's calibration file. Then click on the **Close** button to close the calibration window. In our example, a unique file named **BenzFID.cal** was created. The **BenzFID.cal** file name will now appear in the **Components** window next to Benzene.

WARNING:
Do not use the same calibration curve file name for two different channels or detectors since each detector requires its own calibration curve. (ie Benz<u>FID</u>.cal; Benz<u>PID</u>.cal; etc)

Calibration is required for each component you expect to be present in your sample, and for each detector you will be using in your analysis. Once calibration curves have been completed, and calibration files saved, every component in the component table should show an associated calibration file. PeakSimple will now be able to quantify each component when actual samples are injected.

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Calibration Screen Shortcuts

As an added convenience, PeakSimple for Windows offers shortcuts to commonly used screens. These shortcuts may be accessed by pointing to the desired channel and **clicking once on the <u>right</u> mouse button**. The following pages describe the shortcuts available to set up calibration tables and calibrate components.

After a known standard has been run and the peaks have been identified, a new component table may be constructed by simply positioning the mouse pointer over a peak and clicking once on the right mouse button, ("right-clicking"). The shortcut menu will appear. Select **Add component** from the menu. A retention window will be drawn horizontally across the peak. Right-click again over the peak and select **Edit component**. The **Component Details** screen will open allowing the peak to be named and numbered. The example below shows Benzene as peak #2. The component has been named F-benzene to avoid confusion with a benzene peak from another detector such as a PID.

Note: It is important that you choose the component name carefully since the <u>calibration file name</u> is derived from the <u>first eight letters</u> of the <u>component name</u>. The F-benzene calibration file would be named F-benzen.cal.
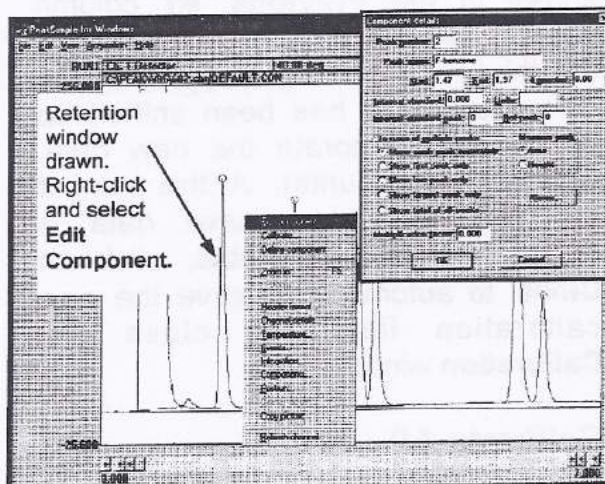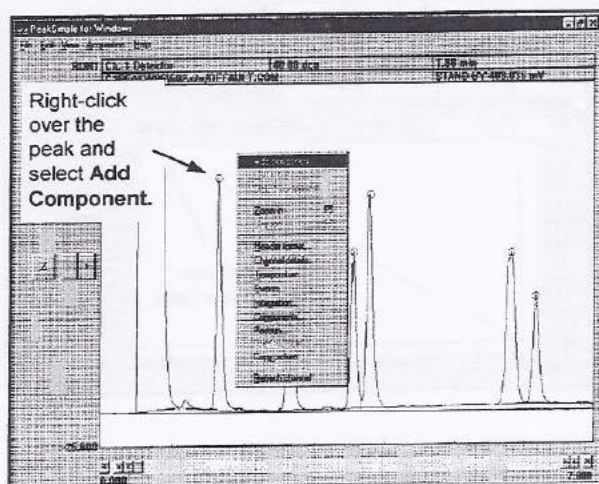
Right-click over the peak again and select **Calibrate**. If no calibration curve exists for the peak, a window will open asking if you would like to use a calibration file. PeakSimple offers a template calibration file aptly named TEMPLATE.CAL. Click yes to use the default TEMPLATE calibration file or select your own by clicking **Browse**. This example uses the template calibration file. Another window will open asking you to select the **Recalibration Level**. Select **100** for 100 ppm standards, **50** for 50 ppm, etc.
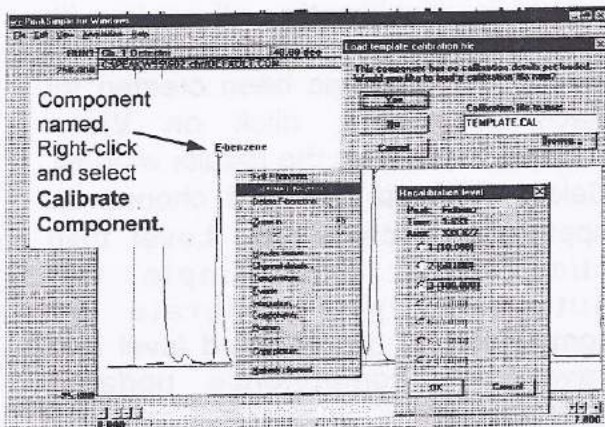
Calibration Screen Shortcuts (continued)

Click **OK** to accept the **Recalibration Level**. The Calibration screen will open and a flashing asterisk (*) will appear along the existing calibration curve depicting the new data point. Notice that the calibration curve has been named **F-benzen.CAL**. If the new caliabration data point is acceptable, click **Accept New** to update the calibration curve data.
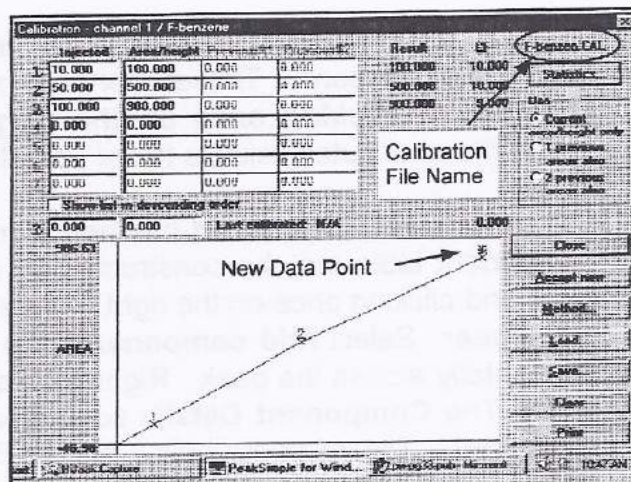
In the example to the right, the updated **F-benzen.CAL** calibration table reflects the new area count of **939.627** at the concentration level of **100** ppm. (The previous calibration data of 900 area counts at 100 ppm is shown in the **Previous #1** column which is 'grayed out'). Notice also that the third data point (100 ppm) in the calibration curve has been shifted up slightly to incorporate the new data, (939.627 area counts). At this point, if the new calibration curve data is deemed to be acceptable, click on **Close** to automatically save the new calibration file, and close the **Calibration** window.
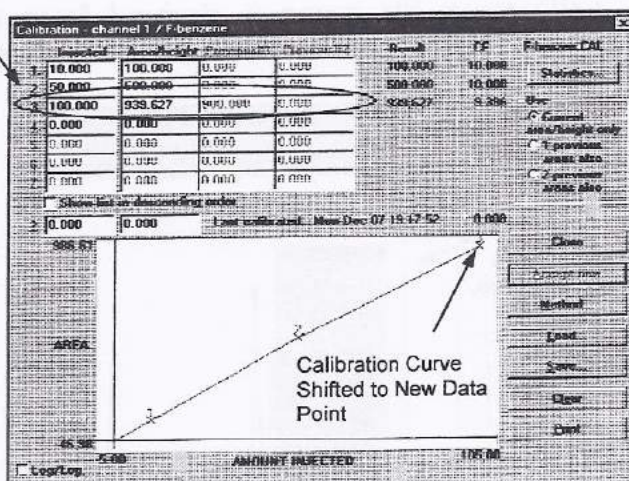
Calibrate All

PeakSimple offers a time-saving feature for **recalibrating all peaks** with just one mouse click. After a calibration curve has been created for each component, click on **View-Results** to bring up the results window. Select **Calibrate All** and choose an appropriate **Recalibration Level**, then click **OK**. PeakSimple will automatically recalibrate all components at the selected level and save each component's updated calibration file.

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Calibration Screen – Use and Statistics Radial Buttons

To improve the calibration accuracy, chromatographers may prefer to average the areas of 1, 2 or 3 replicate injections. The **Use** radio button allows the user to select how many injections are used in the calculation of calibration factors, (CF). Calibration Factors are used to construct the calibration curve using the formula: CF = area count divided by the amount injected. The example to the right shows the calibration data at the 100 ppm concentration level, (circled), with the **Use** button set to the default setting of **Current Area / Height Only**. This setting uses only the latest calibration data to calculate the calibration factor for the #3 data point. (CF = 939.627 / 100 = 9.396)
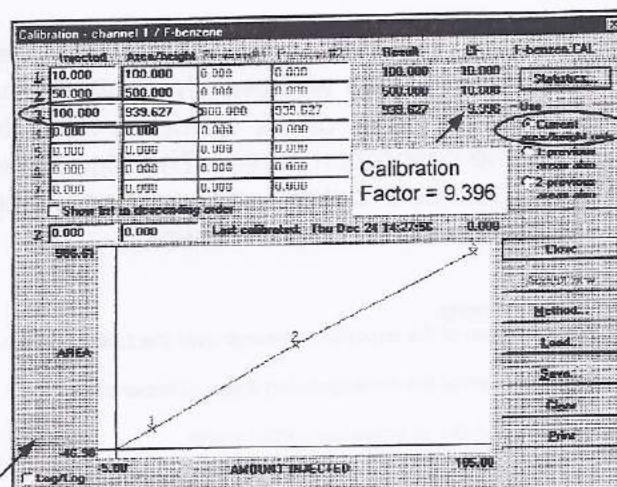
This next example shows how the calibration curve is changed when the **Use** button is set to **1 Previous Areas Also**. This setting averages the last two areas to derive the average calibration factor. Notice that the calibration factor is now 9.198 when the two area counts are averaged together. (939.627 + 900.000 / 2 = 919.814 average area counts. The CF is calculated as: CF = 919.814 / 100 = 9.198)
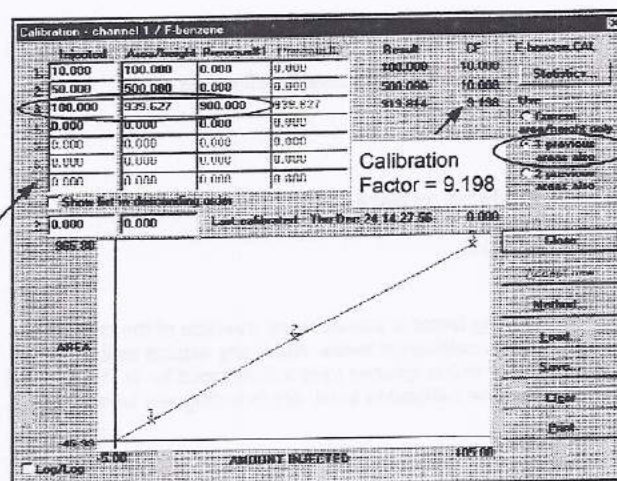
Setting **Use** to **2 Previous Areas Also** will average the last three areas to derive the calibration factor.

The **Calibration Statistics** screen shows calibration curve details such as the **Average Slope of the Curve**, the **Y Intercept**, the **Linearity** of the curve, the **Number of (calibration) Levels**, the **Standard Deviation and Relative Standard Deviation of Calibration Factors**, the **R2** and the **Formula** used which is based on the **Method** selected.

Calibration Factor = 9.396

Calibration Factor = 9.198
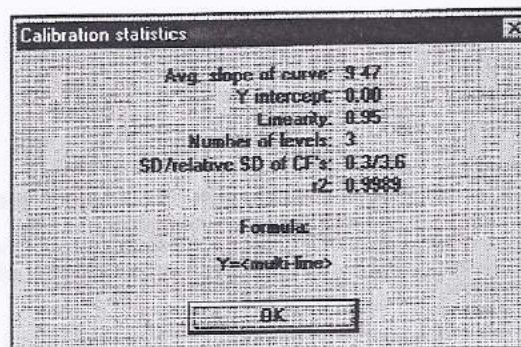
Calibration Window– Methods

The **Method** button opens the **Recalibration Type** window which allows the selection of one of six formulas used to draw the calibration curve. The algorithms are described below and corresponding calibration statistics are shown.

In the following:
X is the sum of the external measures over the calibration levels
Y is the sum of the corresponding areas at those calibration levels
n is the number of active calibration levels
Several other sums are used, for instance:
X2 is the sum of the squares of the external measures
Y4 is the sum of the (area to the 4th power)
XY is the sum of the (external measure * area)
X2Y is the sum of (external measure squared * area)
Y|X is the sum of the (area / external measure) etc.

Single line through origin:

The resulting calibration curve is defined as

$y = Ax$

where:
    x is external measure
    y is area
    $A = (Y|X)/n$

Notes:
The resulting factor is therefore the average of the calibration factors at the calibration levels. Note: any explicit calibration level point at x=0 is ignored (and n is reduced by 1). There must be at least one calibration level, not including any level at x=0.

Single line:

The resulting calibration curve is defined as

$y = Ax + B$

where:
    x is external measure
    y is area
    $A = ((XY * n) - (X * Y)) / D$
    $B = ((X * Y2) - (XY * X)) / D$
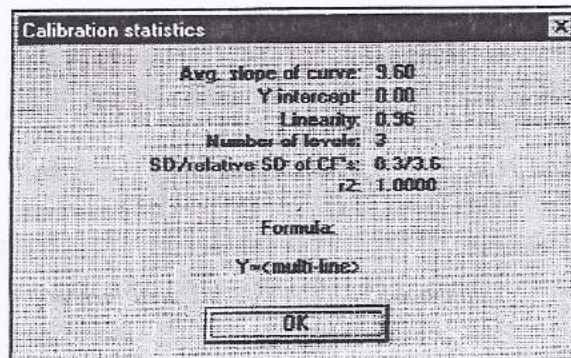 $D = ((X2 * n) - (X * X))$

Notes:
This is a least squares fit algorithm over the calibration levels. A point at (0,0) is also assumed (by incrementing n) unless there is already a value at x=0, or if [Statistics]R2IncludeZero is set to 0 in the PEAKWIN.INI file. There must be at least 2 calibration levels.

EPA rules allow the use of Single Line Fit provided that the standard deviation of calibration factors is <20%.

Multiple line segments:

There is no resulting formula here, just interpolation between the levels, and the origin. There must be at least one calibration level.

Recalibration type

- ● Single line through origin [Ax]
- ○ Single line [Ax+B]
- ○ Multiple line segments
- ○ Parabolic [Ax2+B]
- ○ Quadratic through origin [Ax2+Bx]
- ○ Quadratic [Ax2+Bx+C]

[OK]   [Cancel]

Calibration statistics

Avg. slope of curve: 9.80
Y intercept: 0.00
Linearity: 1.00
Number of levels: 3
SD /relative SD of CF's: 0.3/3.6
r2: 0.9986

Formula:

Y=9.7988X

[OK]

Calibration statistics

Avg. slope of curve: 9.31
Y intercept: 16.83
Linearity: 1.00
Number of levels: 3
SD /relative SD of CF's: 0.3/3.6
r2: 0.9986

Formula:

Y=9.3072X+16.8253

[OK]

Calibration statistics

Avg. slope of curve: 9.60
Y intercept: 0.00
Linearity: 0.96
Number of levels: 3
SD /relative SD of CF's: 0.3/3.6
r2: 1.0000

Formula:

Y=<multi-line>

[OK]

Calibration Window– Methods (continued)

Parabolic:

The resulting calibration curve is defined as

$$y = Ax^2 + B$$

where:
- x is external measure
- y is area
- $A = ((X2Y * n) - (Y * X2)) / D$
- $B = ((Y * X4) - (X2Y * X2)) / D$
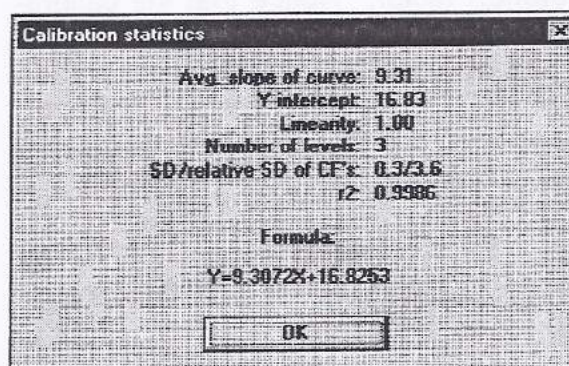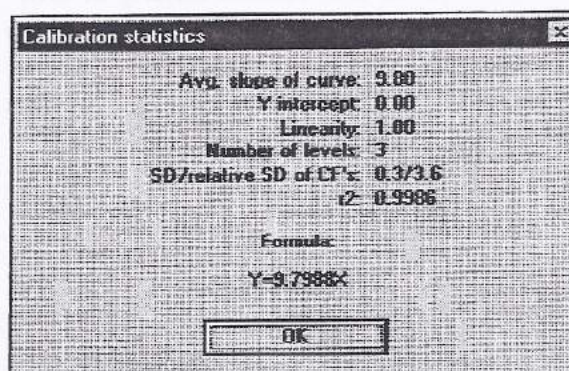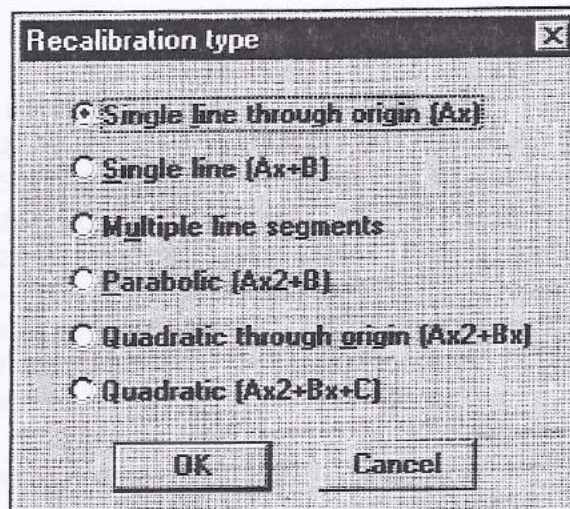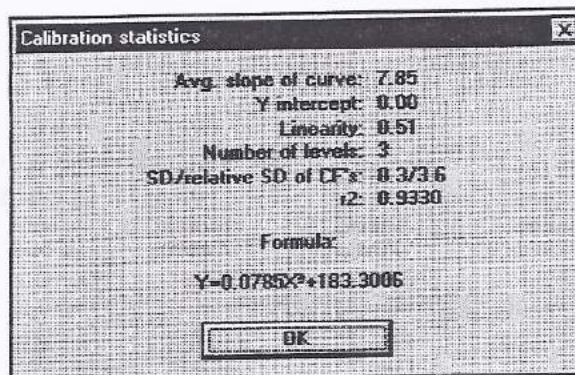- $D = ((X4 * n) - (X2 * X2))$

Notes:
This is a least squares fit algorithm over the calibration levels. A point at (0,0) is also assumed (by incrementing n) unless there is already a value at x=0, or if [Statistics]R2IncludeZero is set to 0 in the PEAKWIN.INI file. There must be at least 2 calibration levels (3 if the origin is not assumed).

Calibration statistics

Avg. slope of curve: 7.85
Y intercept: 0.00
Linearity: 0.51
Number of levels: 3
SD/relative SD of CF's: 0.3/3.6
r2: 0.9330

Formula:

$Y = 0.0785X^2 + 183.3006$

OK

Quadratic through origin:

The resulting calibration curve is defined as

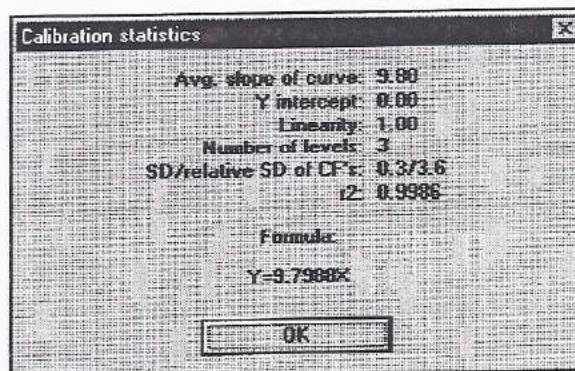$$y = Ax^2 + Bx$$

where:
- x is external measure
- y is area
- $A = ((XY * X3) - (X2Y * X2) / D$
- $B = ((XY * X4) - (X2Y * X3)) / D$
- $D = ((X3 * X3) - (X4 * X2)$

Notes:
This is a least squares fit algorithm over the calibration levels. There must be at least 2 calibration levels.

Calibration statistics

Avg. slope of curve: 9.80
Y intercept: 0.00
Linearity: 1.00
Number of levels: 3
SD/relative SD of CF's: 0.3/3.6
r2: 0.9986

Formula:

$Y = 9.7988X$

OK

Quadratic:

The resulting calibration curve is defined as

$$y = Ax^2 + Bx + C$$

where:
- x is external measure
- y is area

$A = ((XY*X-Y*X2)*(X2*X2-X*X3) - (X2Y*X2-XY*X3)*(X*X-X2*n)) / D$

$B = ((XY*X2-Y*X3)*(X2*X3-X*X4) - (X2Y*X3-XY*X4)*(X2*X2-X*X3)) / E$

$C = ((XY*X2-Y*X3)*(X3*X3-X2*X4) - (X2Y*X3 - X*X4)*(X2*X2-X*X3)) / F$

$D = ((X3*X-X2*X2)*(X2*X2-X*X3) - (X4*X2-X3*X3)*(X*X-X2*n))$

$E = ((X2*X2-X*X3)*(X2*X3-X*X4) - (X3*X3-X2*X4)*(X*X2-X3*n))$

$F = ((X*X2-X3*n)*(X3*X3-X2*X4) - (X2*X3-X*X4)*(X2*X2-X*X3)$

Notes:
This is a least squares fit algorithm over the calibration levels. A point at (0,0) is also assumed (by incrementing n) unless there is already a value at x=0, or if [Statistics]R2IncludeZero is set to 0 in the PEAKWIN.INI file. There must be at least 2 calibration levels (3 if the origin is not assumed).

Calibration statistics

Avg. slope of curve: 9.31
Y intercept: 16.83
Linearity: 1.00
Number of levels: 3
SD/relative SD of CF's: 0.3/3.6
r2: 0.9986

Formula:

$Y = 9.3072X + 16.8253$

OK
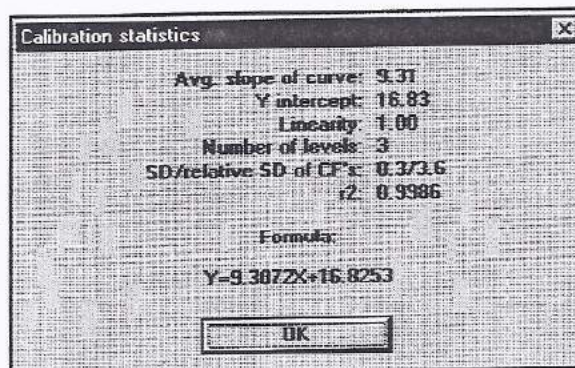
The Calibration Window (continued)

The **Accept New** Button

If the new calibration data is acceptable, Click **Accept New** to update the calibration curve data.

The **Close** Button

Automatically saves the new calibration file and closes the Calibration window.

The **Load** Button

Click on the **Load** button to load an existing calibration file, designated with the .CAL file extension.

The **Save** Button

Click on the **Save** button to save a new calibration file, or to update an existing one. Remember to always use the .CAL extension when naming the calibration file. The saved file name appears at the top of the calibration window indicating the file in use.

The **Clear** Button

Clicking on the **Clear** button deletes all calibration data from the calibration window. The calibration file name is also removed.

The **Print** Button

Clicking on the **Print** button sends the file data and the calibration curve information to the printer.
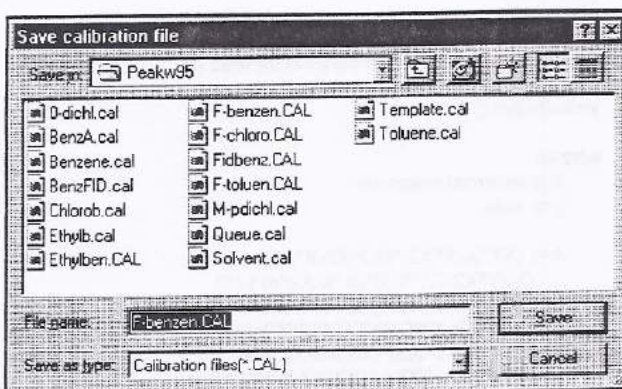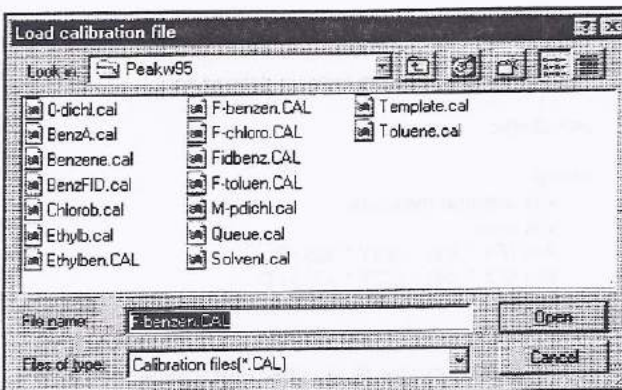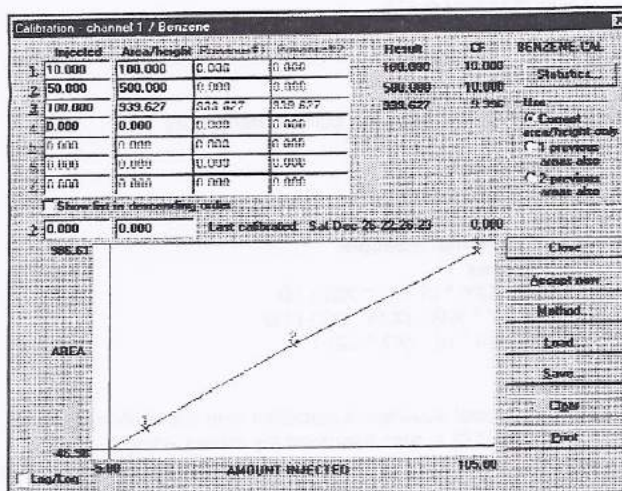
The Edit-Channels-Postrun Window

The Postrun Screen is used to determine all the actions that are to be done in PeakSimple after a chromatogram run. Clicking on the **Postrun** box for channel 1 in the Channel controls window will open up the Channel 1 post-run actions window.

Save file as "X"

The Save file as checkbox, when selected, automatically saves a chromatogram file to disk after a run is completed. The file will be saved under the file name and path entered in the information field to the right of the checkbox.

Auto-increment

When selected, the Auto-increment checkbox will incrementally add a numerical digit to the entered filename after each run. For example, a chromatogram run saved as RUN.CHR would be saved as RUN1.CHR after the second run and RUN2.CHR after the third run.

The **Save results** checkbox when selected will save the data in the results screen to disk after a chromatogram run (*Note: This is not the raw data but instead is the ASCII results*). The **Add to results log "X"** checkbox adds the results of a run to the results log specified in the information field to its right. It will be saved under the same filename as the raw data but with the extension .RES, for example 602.RES. The **Print results** checkbox will print whatever is specified to be printed in the Print format window, this might include the chromatogram and its results data. The **Update DDE link** checkbox when selected will automatically update the Dynamic Data Exchange link once the run is completed.

Execute "X"

The Execute information field opens any executable file (.exe, .bat, .bas) after the chromatogram run is completed. *Note: Be sure to include the full filename and path for the executable file.* Control is returned to PeakSimple when the called application closes.

Restart run after "X"

The Restart run after "X" checkbox and information field restarts a chromatogram run after an inputted delay time. The delay time is inputted in minutes and can be repeated as many times as is entered into the times total information field. *Note: If 0 is entered into the times total information field then the run will be restarted an infinite number of times.*
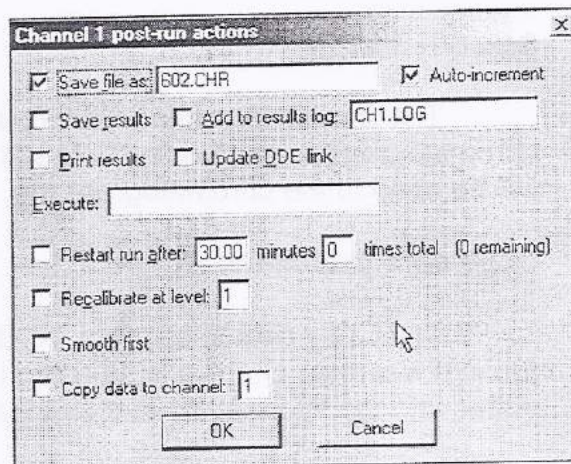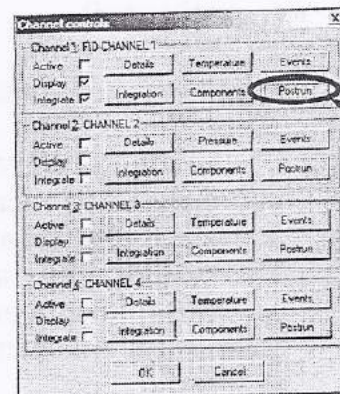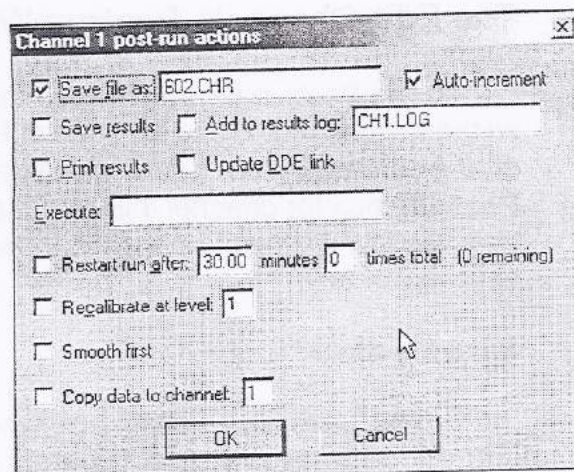
The Edit-Channels-Postrun Window (continued)

Recalibrate at level "X"

The Recalibrate at level "X" checkbox and information field recalibrates all identified peaks at the end of a run at a given level from 1 to 7. This feature is normally implemented as part of an autosampler queue. Detailed instructions are given in the Autosampler queue documentation section.

Smooth first

The Smooth first checkbox runs the smoothing algorithm as it was last applied to the chromatogram before the final integration is done. If the box is left unchecked no smoothing will be done to the chromatogram run.

Copy data to channel "X"

The Copy data to channel "X" checkbox and information field inputs the chromatogram run into whatever channel is selected in the information field. Only the values 1 to 4 can be inputted into the information field as there are four chromatogram channels in PeakSimple.

The Edit-Overall Window

The Overall controls window is used to define and control many of the options in PeakSimple. Clicking on **Edit** in the PeakSimple menu bar and then **Overall** from the drop down menu will open up the Overall controls window.

Unknown peaks are labeled "X"

The Unknown peaks are labeled information field, when filled out, labels all unknown peaks the value that is in the information field. If the word Peak was entered into the information field then all unknown peaks would be labeled Peak.

The **Show retention windows** checkbox is checked by default and thus retention windows are visible in PeakSimple; unchecking the Show retention windows checkbox removes the retention windows from sight. The **COM Port / USB device number "X"** information field specifies the COM port or USB device number that is to be used for the connection between PeakSimple and hardware. The COM port number is typically 1 or 2 while the USB device number is typically between 5000 and 9999.

Label peaks onscreen by

The Label peaks onscreen by options box enables a peak to be labeled by as many as four options. The **Number** checkbox labels all peaks with their peak number. The **Name** checkbox labels all peaks with their full name. The **Abbreviated name** checkbox labels all peaks with a shorter, four character abbreviated name while the **Retention time** checkbox labels peaks with their retention times. The **Draw label vertically** checkbox specifies whether peaks should be labeled horizontally or vertically on the chromatogram screen. When the box is checked the peaks labels will be drawn vertically when it is deselected they will be drawn horizontally.
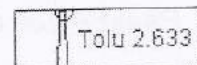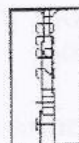
Gradient settings

Gradient settings are only used when PeakSimple is controlling an SRI HPLC Pump. The **Min** and **Max** voltage settings are used to calibrate the Pump.
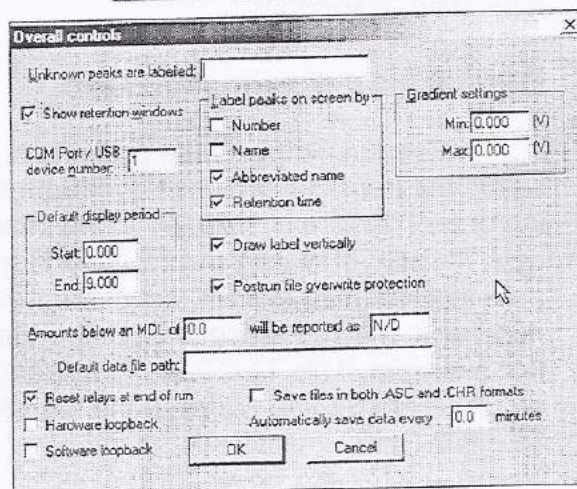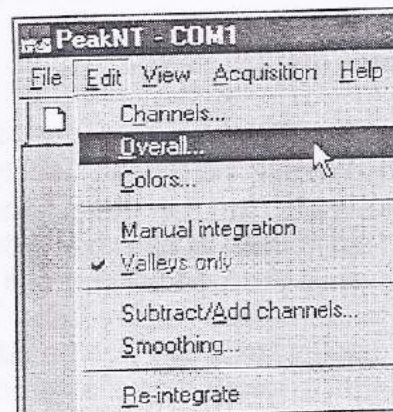
The Edit-Overall Window (continued)

Default display period

The default display period options box is used to define the default display limits for a PeakSimple chromatogram. The **Start** information field is used to specify the default beginning limits while the **End** field is used to specify the end to the default display limits. The start and end display limits can also be adjusted by the left and right arrows below the chromatogram in the main display window.

Postrun file overwrite protection

Postrun file overwrite protection protects a saved file from being written over when the auto-increment feature is selected in the Postrun window. Instead of writing over a used filename an auto-incremented run will select the next unused number in the sequence to save the file to disk. For example, if file TEST02.CHR already exists on disk PeakSimple will save the file as TEST03.CHR.

Amounts below an MDL of "X" will be reported as "Y"

Peaks with a value below a specified Minimum Detection Level or MDL will be reported as whatever is specified in the second information field, typically N/D or not detected. The number that is below the MDL will not be reported, only the entry in the second information field will be seen.

Default data file path

Typically all PeakSimple files are saved to the PeakSimple directory but by entering a full directory path into the Default data file path information field another directory can be selected to save files to. *Note: It is recommended that users save all PeakSimple files to the PeakSimple directory. If necessary export files to a different directory after saving them to the PeakSimple directory.*

Reset relays at end of run

The Reset relays at end of run checkbox when selected turns off all relays (A-H) at the end of a chromatogram run. If the box is left unselected the relays will not be shut off after a chromatogram run.

Hardware loopback and **Software loopback** are used for system validation and will be discussed in further detail in the Loopback test section.

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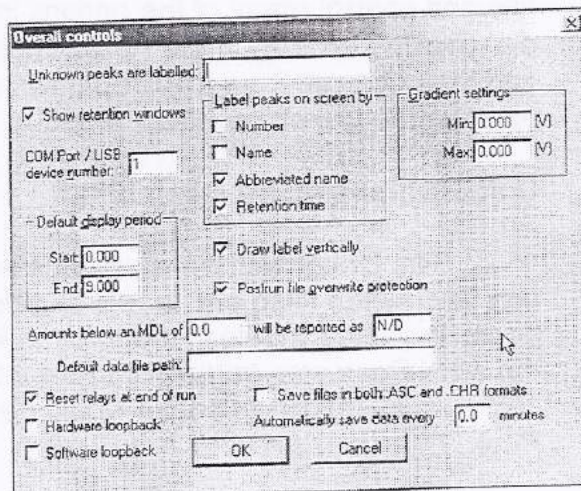
The Edit-Overall Window (continued)

Save files in both .ASC and .CHR formats

The Save files in both .ASC and .CHR formats checkbox when selected saves files in the .ASC format (ASCII) and the .CHR format (chromatogram). If the checkbox is not selected files will be saved only in the .CHR format.

Automatically save data every "X" minutes

The Automatically save data every "X" minutes checkbox and information field when selected saves the data during a chromatogram run at intervals specified by the information in the information field. This feature is useful for runs where power outages are frequent and data cannot be lost.

The Edit-Colors Window

The Colors window determines the color schemes that are to be used throughout PeakSimple. Open the Colors window by selecting **Edit** from the PeakSimple menu bar and then **Colors** from the list of options.

Selecting the **Background** button with the mouse cursor opens up the Background color window. The background color can be chosen from a set of 48 colors by selecting a color and then affirming the choice by clicking on the OK button.

The Graph background window is opened up by selecting the **Graph background** button in the Colors window. The graph background color is changed by selecting a color and then clicking on the OK button to make the color change.

The color of the labels controls the color of the words that belong to the peaks. The color of the labels is changed by selecting the **Labels** button to open up the Labels color window. In the Labels color window select a color and then press on the OK button to make the change to the labels color.

The peak color is the color of the circle at the top of each identified peak and is determined by the Peak color window which is opened up by selecting the **Peak** button in the Color window. Select the desired peak color and then click on the OK button to close the window and affirm the change.

The color of the zero axis is chosen by clicking on the **Zero axis** button and then selecting a color from the Zero axis color window. Clicking on the OK button closes the window and makes the change to the color of the zero axis. Don't set the Zero axis color to the same color as the Graph background because they won't be distinguishable from each other.

The baseline is the line that runs along the bottom of the peaks and its color is changed by selecting the **Baseline** button and then choosing a color from the Baseline color window. The change is made once the OK button is selected and the window is closed.

The data line is the signal line that makes up the peaks in PeakSimple and its color is defined by selecting the **Data line** button in the Colors window and then selecting a color from the Data line colors window. Once the desired color is selected apply the color change by clicking on the OK button to close the window.

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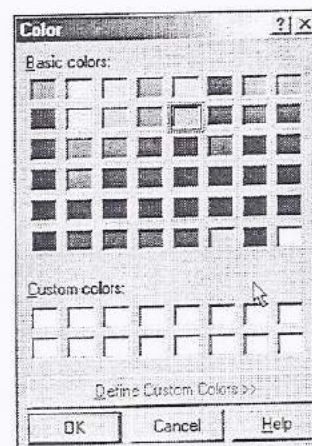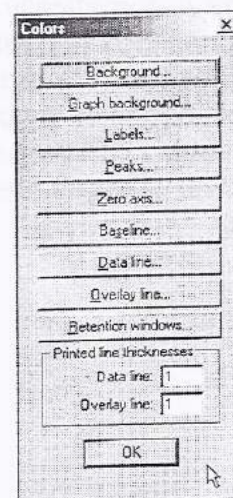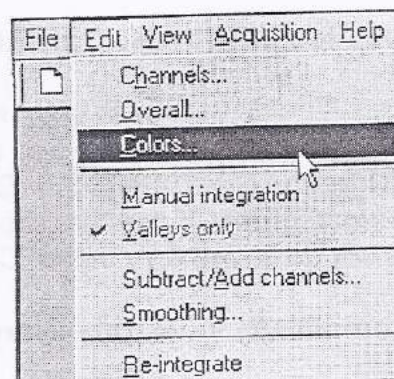
The Edit-Colors Window (continued)

The overlay line is a data line from a chromatogram that has been overlaid on top of an existing chromatogram and its color is changed by selecting the **Overlay line** button in the Colors window and then selecting a color with the mouse cursor in the Overlay line colors window. The color changes are made once the OK button is selected and the window closes.

Retention windows are the horizontal bars that appear onscreen and their color can be changed by clicking on the **Retention windows** button in the Colors window and then selecting the desired color in the Retention windows colors window. To apply the color changes click on the OK button to close the window.

Printed line thickness

The thickness of the Data line and the Overlay line when a chromatogram is printed is determined by the **Data line** information field and the **Overlay line** information field. The thickness of the Data line is determined by the numerical value in the Data line information field, larger numerical values will result in thicker lines. The thickness of the Overlay line is also determined by the numerical value in its information field. Larger numbers in the information field will result in a thicker overlay line.
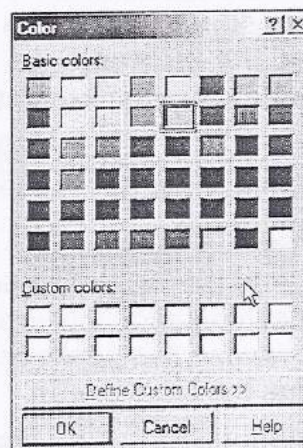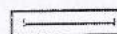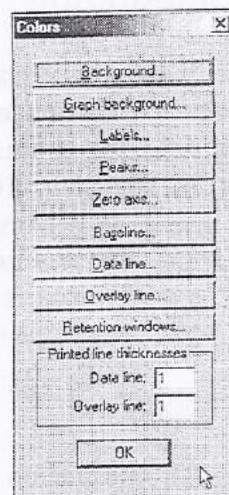
Manual Integration

The manual integration tools are used to manually draw in the baseline in a PeakSimple chromatogram. The manual integration toolbar is opened up by selecting **Edit** from the PeakSimple menu bar and then clicking on the **Manual integration** option. The manual integration toolbar appears to the right of the PeakSimple toolbar in the upper right hand corner of the screen.

Off Integration Tool

The Off integration tool or the mouse cursor is used to end a manual integration mode once it has been selected. When the mouse cursor icon is selected no more changes to the baseline of a chromatogram can be performed until another manual integration tool is selected.

None Integration Tool

The None integration tool adds the area of one peak to the area of an adjacent peak. Once the None integration tool is selected click on a valley between two peaks with the mouse cursor to change the baseline.
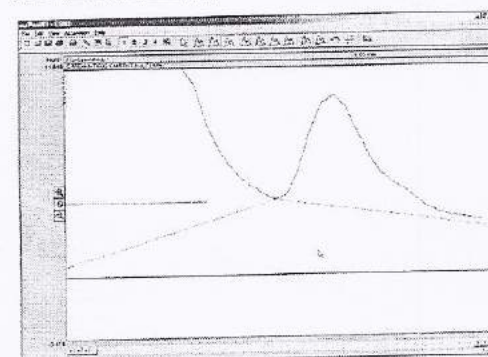
Drop Integration Tool

The Drop integration tool drops the baseline between two peaks straight down onto an existing baseline. The Drop integration tool is used by selecting the Drop tool in the manual integration toolbar and then clicking on a valley between two peaks to change the baseline.

Based Integration Tool

The Based integration tool raises the baseline to a valley between two specified peaks. To change the baseline select the Based tool and click on a peak with the mouse cursor to raise the baseline up to the valley.
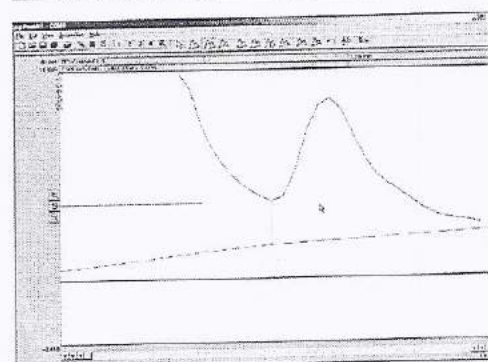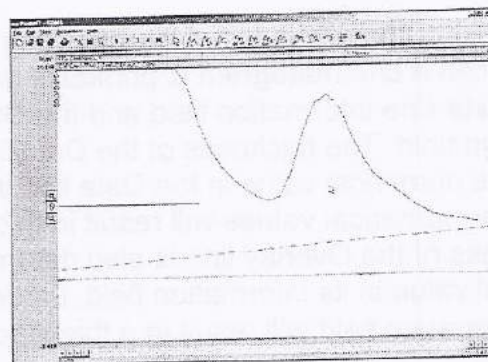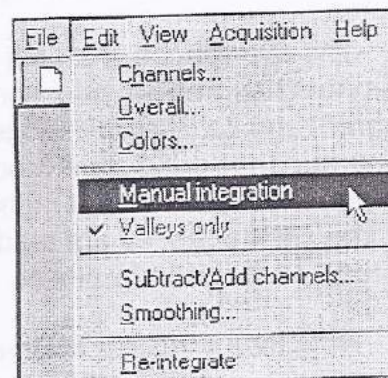
Manual Integration (continued)

Lead Skim Integration Tool

The Lead skim integration tool skims a peak's area off of the leading edge of an adjacent peak. To skim a peak off of the leading edge of another peak select the Lead skim tool from the manual integration toolbar and then click on the valley between the two specified peaks with the mouse cursor.

Trail Skim Integration Tool

The Trail skim integration tool skims a peak's area off of the trailing edge of another, adjacent peak. To skim a peak off of the trailing edge of another peak select the Trail skim tool and click on a valley between two peaks with the mouse cursor to make the change.

Lead Horizontal Integration Tool

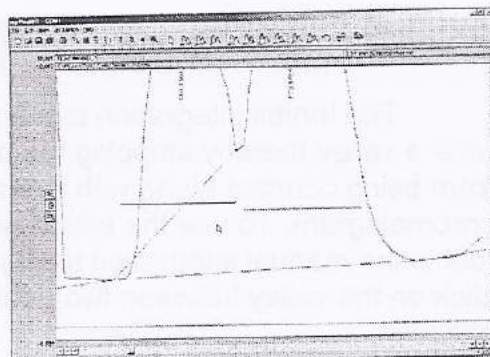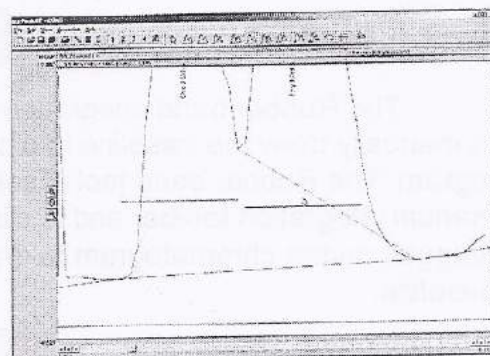The Lead horizontal integration tool draws the baseline horizontally for the leading peak while the trailing peak's baseline stretches from the horizontal line to the next valley. The Lead horizontal tool is selected in the manual integration toolbar and once a valley is selected the change to the baseline is made.
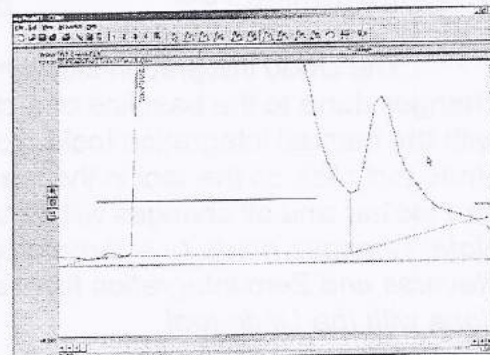
Trail Horizontal Integration Tool

The Trail horizontal integration tool draws the baseline horizontally for the trailing peak while the leading peak's baseline stretches from the horizontal line to the previous valley in the chromatogram. The Trail horizontal tool is used by selecting the Trail horizontal tool in the manual integration toolbar and then clicking on a valley with the mouse cursor to make the change.

Manual Integration (continued)

Inhibit Integration Tool

The Inhibit integration tool ends a baseline after a valley thereby stopping the peak's area from being counted along with the rest of the chromatogram. To use the Inhibit tool select the tool in the manual integration toolbar and then click on the valley between two peaks to end the baseline.

Rubber Band Integration Tool

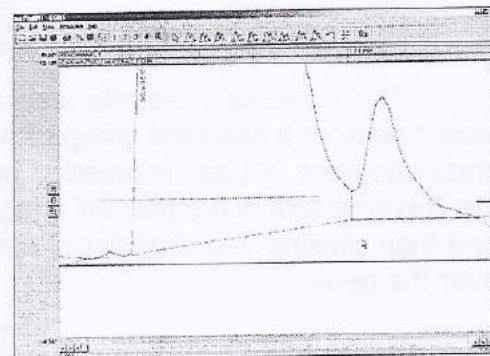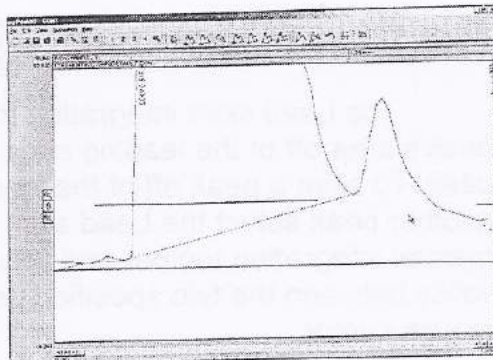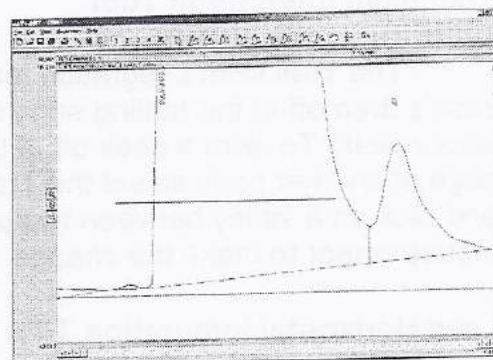The Rubber band integration tool is used to manually draw the baseline in a chromatogram. The Rubber band tool is selected in the manual integration toolbar and is clicked and dragged on the chromatogram to draw in the baseline.

Undo Integration Tool

The Undo integration tool removes all changes done to the baseline of a chromatogram with the manual integration tools. To use the Undo tool click on the tool in the manual integration toolbar and all changes will be undone.
Note: Changes made to a chromatogram with the Reverse and Zero integration tools cannot be undone with the Undo tool.
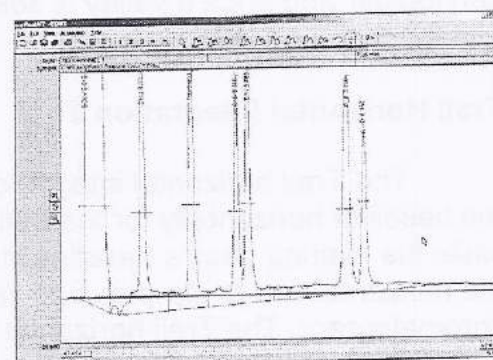
Reverse Integration Tool

The Reverse integration tool inverts a selected peak or a selected group of peaks in a chromatogram. A peak is inverted by selecting the Reverse tool in the manual integration toolbar and then clicking and dragging the mouse cursor over the peak.
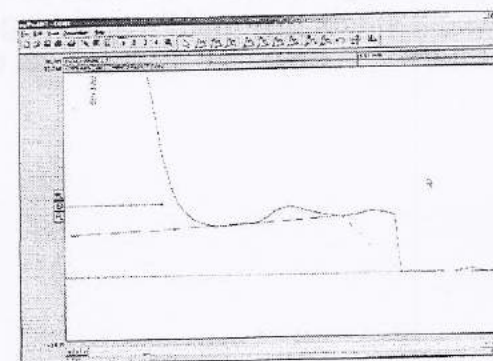
Zero Integration Tool

The Zero integration tool sets the value of the data line at zero starting at a selected point. To zero the data line at a given point select the Zero tool from the manual integration toolbar and click on the data line with the mouse cursor.

PeakNTdoc

The Edit-Valleys Only Option

The Valleys only option is available only when the Manual integration toolbar is open in PeakSimple. The Valleys only option can be selected by opening up the Manual integration toolbar in the Edit menu and then selecting the Valleys only option immediately below Manual integration in the drop down menu. When the Valleys only option is selected all changes made to the baseline of a chromatogram will snap only to the valleys of the chromatogram. When the Valleys only option is turned off changes made to the baseline of a chromatogram will go to wherever the mouse cursor was clicked.

The Edit-Subtract/Add Channels Menu

The Subtract/Add channels menu removes or adds the analog data signal from/to one channel in PeakSimple from/to another channel. The S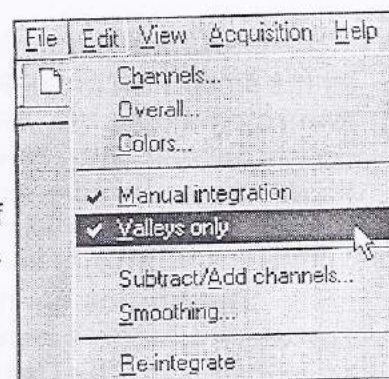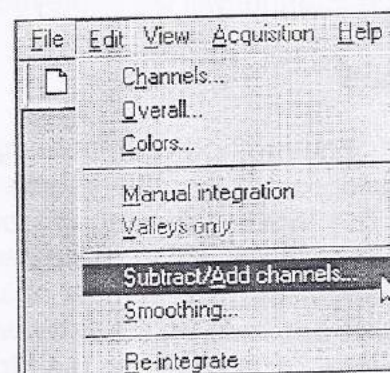ubtract/Add channels menu is opened by selecting the Edit menu and then by clicking on Subtract/Add channel in the drop down menu.

Subtracting a Channel

To subtract one channel from another channel click on the Subtract radio button with the mouse cursor and select the channel that is to be taken away in the first dialogue box. In the second dialogue box select the channel that is to have the first selection taken away from. Click on OK with the mouse cursor to effect the changes.
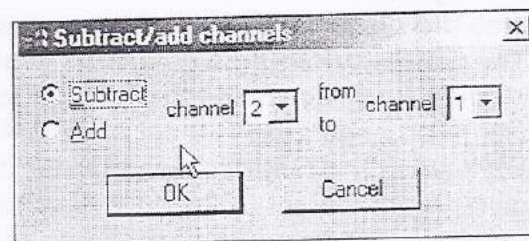
Adding a Channel

To add one channel to another channel select the Add radio button in the Subtract/Add channels menu. Select the channel that is to be added by selecting a number in the first dialogue box and then choose the channel that it is to be added to by selecting a number in the second dialogue box. All changes are made once the OK button is selected.
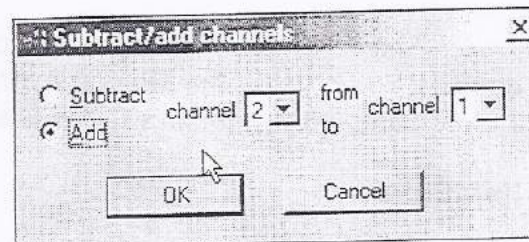
The Edit-Smoothing Window

The Data smoothing window determines all the smoothing options that are to be performed on a data line. The Data smoothing window is opened up by selecting Edit from the PeakSimple menu bar and then selecting Smoothing from the list of options.

The **Source channel** dialogue box specifies which channel the data line that is to be smoothed is in. The **Destination channel** is the channel that the smoothed data line from the source channel will be displayed in.

Method

The method of smoothing is determined by the smoothing algorithm selected in the Method box. The **Moving Average** algorithm sets each sample to the average of the samples around it including itself. The number of samples taken into account depends on the Filter width. The **Olympic** algorithm is similar to the Moving Average but the highest and lowest values in the set of samples are discarded before the average is taken. The **Savitsky-Golay** algorithm is similar to the Moving Average but each of the samples is weighted according to a set of weighting factors. Increasing the number in the **Order** dialogue box gives more weight to the central samples when using the Savitsky-Golay method.

Filter Width

The Filter width dialogue box controls the number of samples that are to be taken into account when using the Moving Average smoothing method. A filter width of 2 means that 2+1+2 samples are taken while a filter width of 5 means that 5+1+5 samples are taken.

Iterations

The Iterations dialogue box controls the number of times a smoothing method is to be applied to a chromatogram peak. Every iteration smoothes the data line more than the previous iteration eventually making the data line flat.
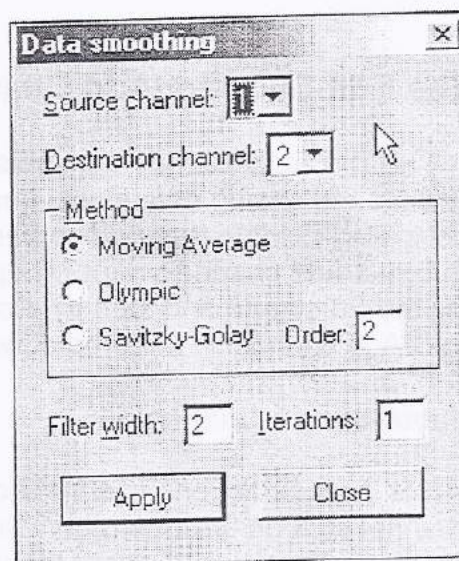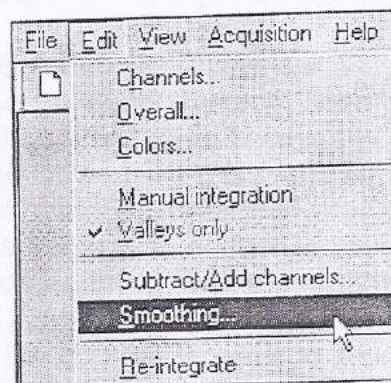
The Re-Integrate Option

The Re-integrate option is used to fully re-integrate a baseline in PeakSimple. When changes are made to a baseline often a partial integration will occur, selecting Re-integrate will perform a full integration on the baseline. The Re-integrate option can be selected by clicking on Edit in the PeakSimple menu bar and then Re-integrate from the list of options.

The View-Results Window

The Results window displays the results of the chromatogram runs performed in PeakSimple. The Results window is opened up by clicking on View in the PeakSimple menu bar and then selecting Results from the list of options.

The **Channel** option scrollbar specifies which of the four channels the results data should be displayed for. When the **Recognized peaks only** checkbox is selected only the results for named peaks will be displayed. The **Undetected components also** checkbox displays the results for the undetected components as well as the detected components in the chromatogram run when the option is selected.

Update

The Update button in the Results window updates the DDE link between the Results data and the DDE host program (typically Excel).

Save

Selecting the Save button in the results window opens up the Save results file window. In the Save results file window the results file is saved with a .res extension. The file is an ASCII file and not the raw chromatogram data.

Integration

As a convenience the integration button in the results window opens up the same Integration window that can be accessed in the Channels window. For more information on the Integration window consult the Channels-Integration portion of this manual.

PeakNTdoc-13.pub
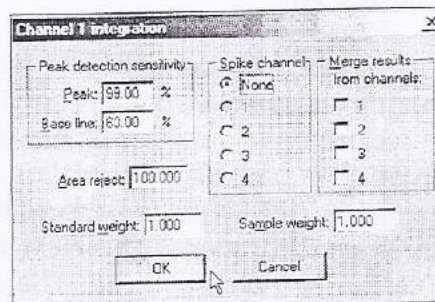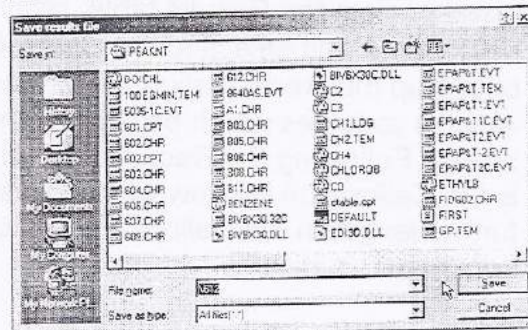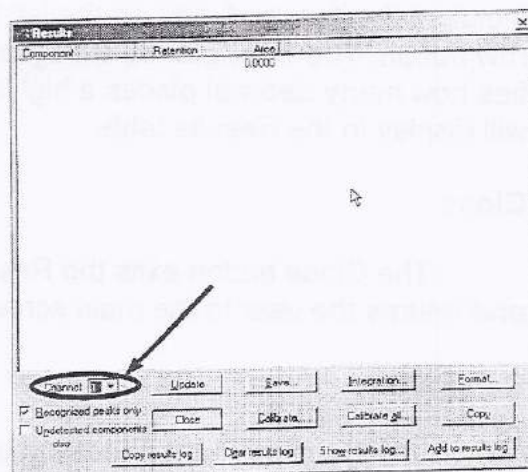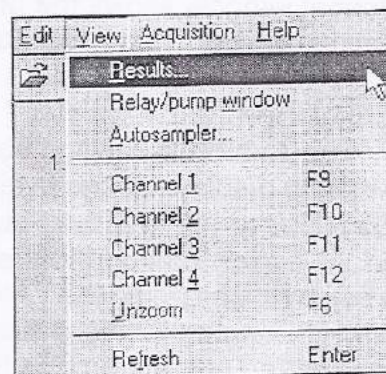
The View-Results Window (cont.)

Format

Selecting the Format button in the Results window opens up the Edit format window. The Edit format window allows the user to specify the information that is to be included in the Results table.

The **Available** options box in the Edit format window displays all the available options that can be included in the results but that aren't selected. An option is added to the **Selected** options box by highlighting the item in the Available box and clicking on the right facing arrow button. To deselect an option from the Selected box highlight the item and click on the left facing arrow button. The **Dec. places** dialogue box specifies how many decimal places a highlighted unit will display in the Results table.
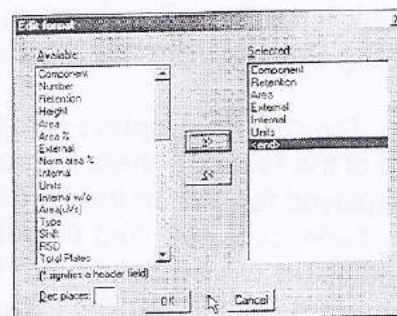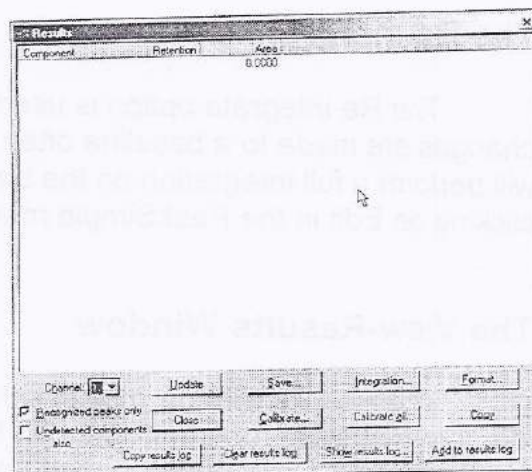
Close

The Close button exits the Results window and returns the user to the main screen.

Calibrate

The Calibrate button recalibrates a recognized peak in the Results table. Highlighting a peak name and selecting the Calibrate button opens up the Recalibration Level window. The window specifies which peak level should be calibrated. Following the Recalibration level window is the Calibration window which is discussed at further length in the Calibration section of this document.
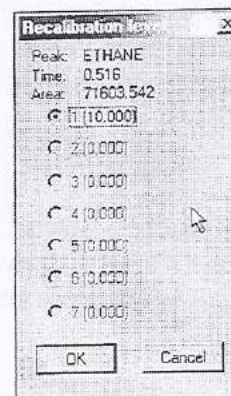
Calibrate All

The Calibrate all button recalibrates all the recognized peaks at once. The Calibrate all button calibrates all peaks with existing calibration curves on a particular calibration level. If named peaks are in the results table without calibration curves an error message, (NOT ENOUGH DATA POINTS), will be displayed. The calibration will
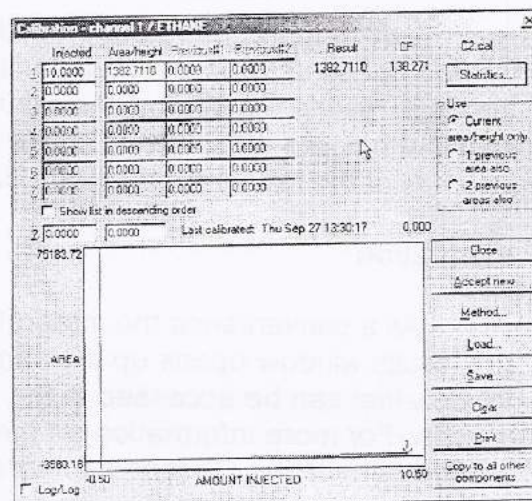
The View-Results Window (cont.)

Copy

The Copy button in the results window copies the results report to the Clipboard. Once the report is copied it can be pasted into other programs i.e. Excel.

Copy Results Log

The Copy results log button copies the .log file for the results to the Clipboard. This log file can be pasted into any Windows program. A certain number of lines in the results log will always be copied, by default the number is 20. If more than 20 lines are needed for an application the user must modify the peakwin.ini file located in the Windows folder. The default entry in the file is (SpareLines=20), delete the number 20 and insert the number of lines that are needed (up to a maximum of 100).

Clear Results Log
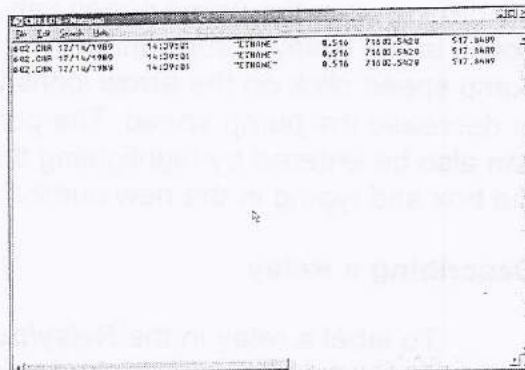
Clicking on the Clear results log button erases the results log file.

Show Results Log

The Show results log button when selected opens up Windows Notepad to view the results log.

Add to Results Log

To add the current report to the results log click the Add to results log button. The report can automatically be added to the results log at the end of each chromatogram run by checking the Add to results log checkbox in the Postrun window.

The View-Relay/Pump Window

The Relay/pump window manually controls the actions of the relays in PeakSimple. The Relay/pump window is opened up by opening the View menu and then selecting Relay/pump window from the list of options.

Selecting/Deselecting a Relay

To manually activate a relay click on the letter next to the relay label to make the button dark. To deactivate a relay select the specified lettered button to turn it black. Pressing the control button and the letter corresponding to the relay together also selects/deselects the relay.

Pump Speed

When an SRI HPLC pump is connected to the data system the pump speed can be controlled in the Relay/pump window. To change the pump speed click on the arrow icons to increase or decrease the pump speed. The pump speed can also be entered by highlighting the value in the box and typing in the new number.

Describing a Relay

To label a relay in the Relay/pump window right click on the main screen and select Events from the list of options. Once the Events window is opened up clicking on the Describe button opens up the Event description window. To enter a relay description click on the specified relay's dialogue box and type in the information. The description of the relays has no effect on the relay function and will not affect hardware.

PeakNTdoc-
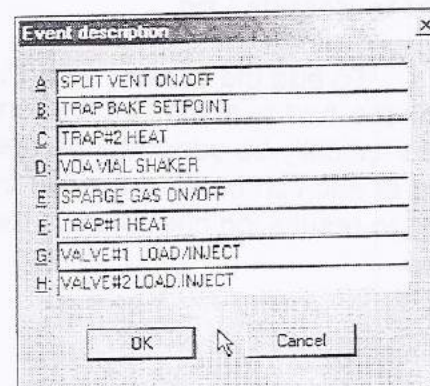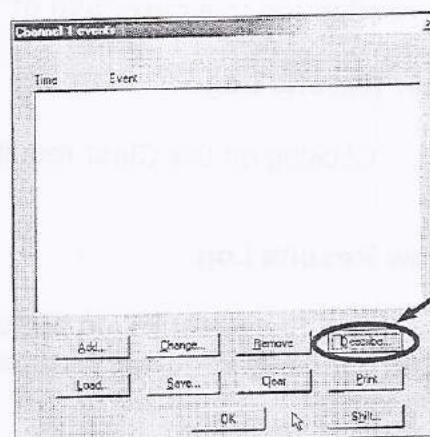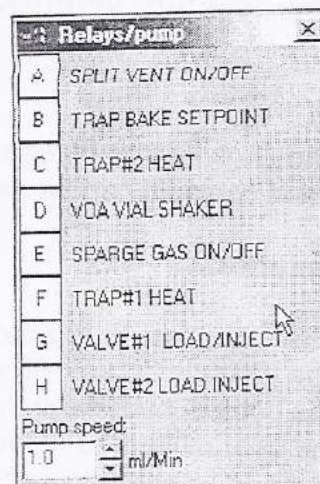
The View-Autosampler Window

The Autosampler window allows a list of control files to be run automatically. Control files are the master files which specify all parameters including temperature programming, component, and event files. These control files run tasks in PeakSimple. To open up the Autosampler window click on the View menu in the menu bar and then select Autosampler from the available options.
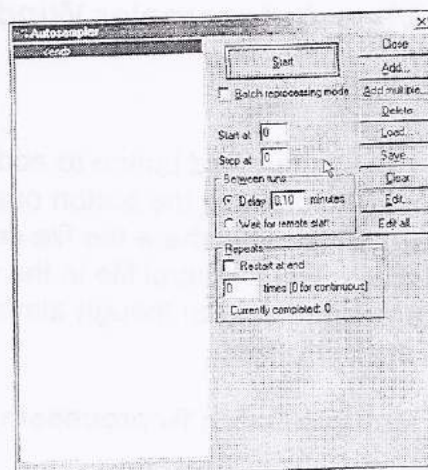
Start/Stop

The Start button when pressed begins the operation of the autosampler queue or reprocessing queue. A queue must be created or loaded before the control files can run. Once the autosampler is in operation the Start button changes into the Stop button. The Stop button ceases the autosampler operations that were previously running.

Batch Reprocessing Mode

To select Batch reprocessing mode click on the check box to the options left. While using the Batch reprocessing mode the user loads a list of previously stored chromatogram files in the list box to the left and then selects a control file which will reprocess the data files. When the operation begins PeakSimple will load each data file in the list into channel 1, perform the specified functions, and then increment to the next data file in the list.

The **Start at** dialogue box specifies which control file number to begin operation at first. If no number is entered the autosampler will begin at the first control file. The **Stop at** dialogue box specifies the last control file to be run before operations of the autosampler cease. If no number is entered in the dialogue box the autosampler will end after the last control file in the list is run.

The **Delay "x" minutes** radio button when selected specifies how many minutes PeakSimple will wait before running the next control file in the list box. The **Wait for remote start** radio button when selected instructs the autosampler to wait for a remote start signal before advancing to the next control file.

The **Restart at end** checkbox restarts the queue after getting to the end of the control files in the list box. In the **"x" times** the user enters the number of times the control files in the list box should be cycled if the Restart at end checkbox is selected. If the value 0 is selected the queue will be cycled continuously.

Close

The Close button closes the Autosampler window when it is selected.

Close

PeakNTdoc
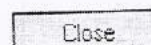
The View-Autosampler Window (cont.)

Add
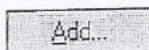
Select the Add button to add a control file to the queue. Selecting the button opens up the Select control file window where the file can be loaded into the list box. Each control file in the queue must have a different name even though almost identical actions are performed.
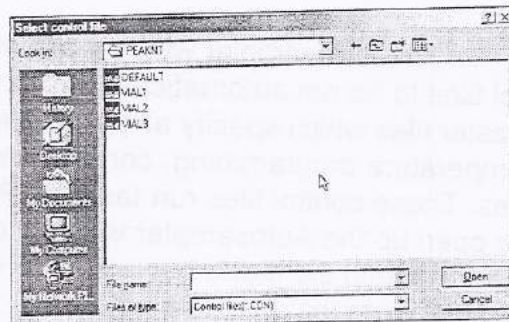
Add Multiple/Batch Reprocessing
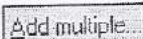
The Add multiple button allows the user to load multiple data files into the list box. Click on the button to open up the Select control file window and then click on a control file name to open up the Select data filenames window. Select as many data files as needed by pressing the shift button and clicking with the mouse cursor and then click on OK to load them into the queue. The Add multiple button is only useful for use with the Batch reprocessing mode.
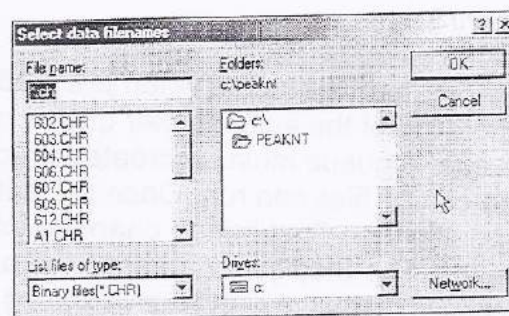
Delete
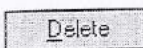
After highlighting a control file in the list box to the left select the Delete button to remove that control file from the queue.

Load

Select the Load button to open up a previously saved queue file. Clicking on the Load button opens up the Load autosampler queue window where the queue file can be selected and loaded.
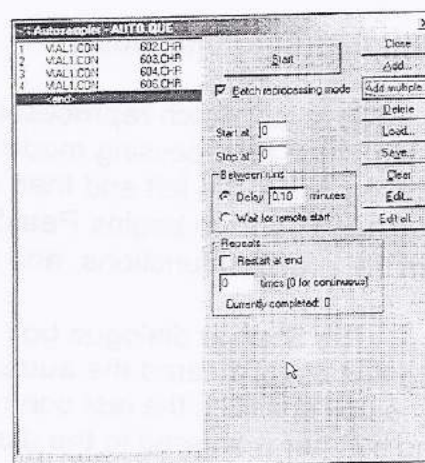
Save
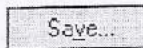
Selecting the Save button opens up the Save autosampler queue window. Save the queue in the file box by naming the file and selecting save. It is recommended that all files be saved to the Peak-Simple directory.
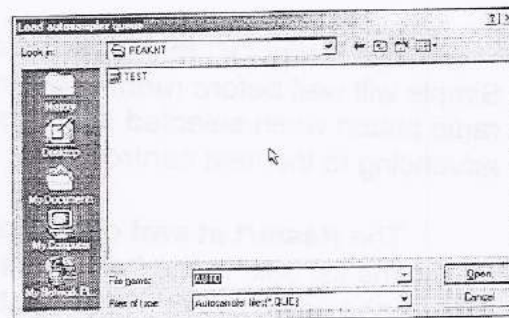
Clear
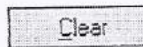
The Clear button erases the entire queue.

Edit

Edit...

After highlighting a control file select the Edit button to modify that control file. Selecting the Edit button loads the control file on the PeakSimple main screen. To make any changes click on the main screen, do all modifications, and then select Save all from the PeakSimple file menu.

Edit All

Edit all...

To edit all the control files in the queue at once click on the Edit all button to open up the Autosampler queue spreadsheet. Many of the commonly adjusted control file parameters are displayed in the spreadsheet enabling the user to input changes to the queue. Not all control file parameters can be modified using Edit all (only the parameters that are selected in Format) and so must be done individually with the Edit function.

Autosampler Queue Window

Close

Close

The Close button exits the window after prompting the user to save the spreadsheet.

Cancel

Cancel

The Cancel button exits the spreadsheet window without prompting the user to save.

Add
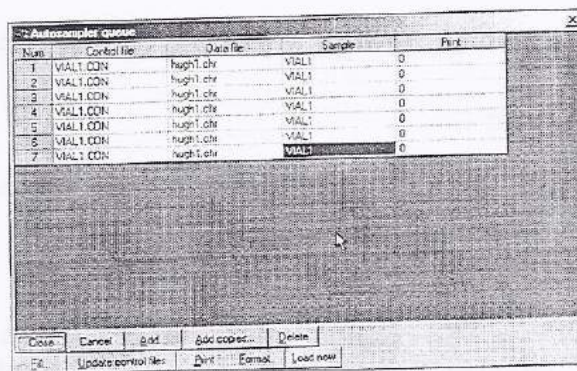
Add...

Selecting the Add button opens up the Select control file window where an existing control file can be added to the queue.
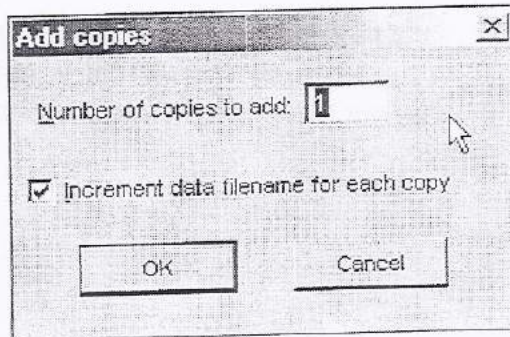
Add Copies

Add copies...

After highlighting a control file in the spreadsheet select the Add copies button to add copies of the file to the list. Once the Add copies window pops up input the number of copies to be made in the dialogue box and specify whether the file names should be incremented. The Add copies button is useful for creating a queue from scratch with a single control file.

Autosampler Queue Window (cont.)

Delete

Delete

The Delete button deletes a highlighted control file off the list. If no file is highlighted then the last file will be deleted from the queue.

Fill

Fill...

The Fill button fills a spreadsheet column, row, or cell with selected text. Once the desired cells are high-lighted clicking the Fill button opens up the Fill autosampler options box. Input the text to fill in the information field and specify whether the text should be incremented for each row.

Fill autosampler

Text to fill: _____

☐ Increment for each row

[OK]   [Cancel]
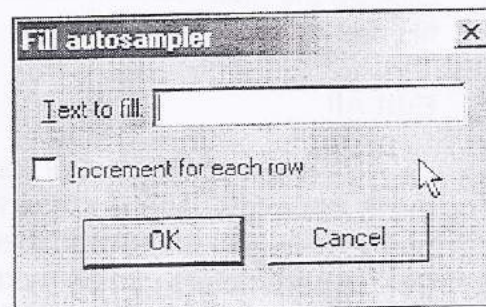
Update Control Files

Update control files

Selecting the Update control files button saves all changes to the control files in the list.

Print

Print

The Print button prints the queue spreadsheet.

Format

Format..

To change the format of the queue spread-sheet and open up the Edit format window select the Format button. In the Edit format window a format type can be added by selecting it in the Available window and then hitting the right facing arrow button. To remove a format type from being displayed in the spreadsheet highlight the format type in the Selected box and click on the left facing arrow.

Edit format

Available:
Lab name
Client
Client ID
Collection date
Holding time
Method
Lab ID
Description
Column
Carrier
Data file
Component file
Event file
Temperature file
Sample
Operator

Selected:
Data file
Sample
Print
<end>

Channels
☑ 1 ☐ 2 ☐ 3 ☐ 4

[OK]   [Cancel]
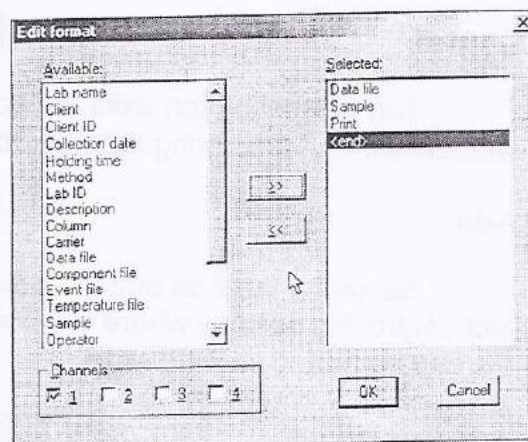
Load Now

Load now

After highlighting a control file select the Load now button to load that control file to the main PeakSimple screen. Click on the screen and make any changes to the control file and then select Save all to save the changes.

The View-Channel "X" Options

To view a specified chromatogram channel open the View menu in the PeakSimple menubar and select a channel to be viewed; either 1, 2, 3, or 4. Keyboard shortcuts can also be used to alternate viewing between chromatogram channels. Hitting F9 displays channel 1, F10 displays channel 2, F11 displays channel 3, and F12 displays channel 4. Furthermore the numerical icons in the PeakSimple toolbar can be used to toggle between chromatogram channels.

Unzoom

To unzoom from a close up view of a chromatogram select the Unzoom tool from the View menu or hit F6. PeakSimple will zoom out to the first level with the soriginal display units of the chromatogram when the Unzoom tool is used. The Unzoom button in the PeakSimple toolbar can also be used to unzoom a chromatogram or F6 on the keyboard.
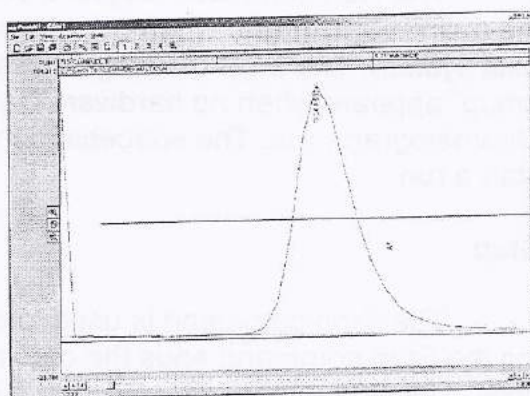
Refresh

The Refresh tool in the View menu redraws the chromatogram screen to fix any glitches or resolve an error. Pressing Enter on the keyboard also refreshes the screen.
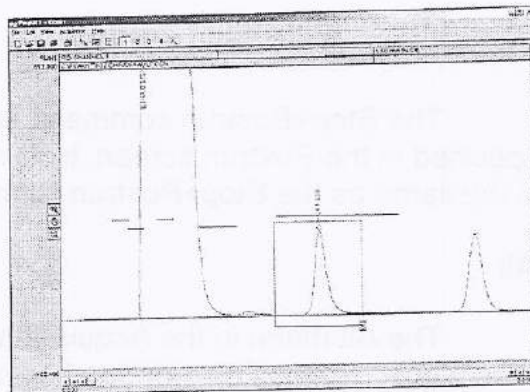
The Help Menu

About PeakNT

To view program information about Peak-Simple click on the About PeakNT option in the Help menu. The PeakNT window will pop up and display the information.

Show Tooltips

The Show tooltips option in the Help menu toggles the PeakSimple tooltips off or on. When Show tooltips is checked a helpful text tip will appear when the mouse cursor is held over a tool or button in PeakSimple. The tooltips provide relevant information to the operation and use of the PeakSimple data system.

PeakNT

Version 2.74 (MS)

Oct 3 2001

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OK
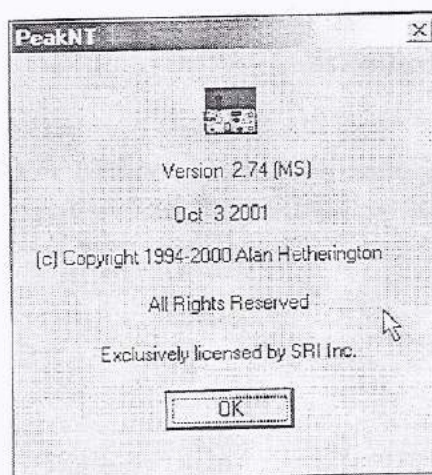
PeakNTdoc-

The Acquisition Menu

The Acquisition menu contains the commands to run a chromatogram run when hardware is connected to the PeakSimple data system. All Acquisition menu commands have corresponding keyboard hotkeys for convenience.

Run

The Run command begins a chromatogram run on the main trigger group when hardware is connected to the data system. The error message "No active channels in group" appears when no hardware is available to make a chromatograph run. The spacebar can also be used to start a run.

Stop

The Stop command is used to end a chromatogram run once it has been started. Using the Stop command ends the chromatogram run without running any of the Postrun operations. The End button can also stop a chromatogram run.

Stop+Postrun

The Stop+Postrun command ends a chromatogram run and executes the operations specified in the Postrun screen. Holding the Control button and pressing End on the keyboard is the same as the Stop+Postrun command.

Alt

The Alt menu in the Acquisition menu controls the acquisition commands for the alternate trigger group. The + button begins the alternate trigger chromatogram run, the - button stops the alternate trigger run, and the / button on the keyboard stops the run and begins the Postrun operations for the alternate trigger group.

Re-initialize

The Re-initialize command reestablishes the connection between the hardware and the PeakSimple data system. A connection between hardware and the data system has to exist for re-initialization to occur.

Loopback Test: For Data Validation

A loopback test may be performed if you are required to validate the precision of the G.C. or Data System's analog to digital conversion. This test requires the user to install a jumper wire on the A/D board inside the G.C. or Data System.

Description of Test:

A jumper wire is installed on the A/D board between 'temperature program one', (TP1), and 'channel one signal input', (Sig. 1+). A data file is then loaded into channel four. When the 'loopback' mode is selected in PeakSimple, the data on channel four is routed out TP1 to the channel one signal input. When a chromatogram run is started, channel one will begin to reproduce the data loaded into channel four. After the run has completed, area counts from a specific data peak may be collected and the run repeated several times. After at least three runs, the user may then calculate the average area counts and the percent relative standard deviation,(%RSD) and thus the precision of the A/D converter. Less than 0.5% RSD is typical for the SRI Model 202 and 203 A/D boards.

Setting Up The Hardware:

With the G.C. unplugged, remove the six screws securing the bottom cover. Flip the G.C. on its back and locate the A/D board on the right-hand side. Remove any wires from 'TP1' and 'SIG 1+' and add an insulated 22 AWG wire between TP1 and SIG 1. Refer to the diagram below for jumper placement.

Most systems will contain the Four Channel 202 Board. Also, verify that SIG 1- is grounded. Add another jumper if needed.

FOUR CHANNEL LAWSON 202 BOARD

CHANNEL INPUTS (5 VOLTS MAX.)
1 2 3 4

TEMP. PROG.

TO SERIAL DATA PORT FOR CONNECTION TO YOUR COMPUTER

+ - + - + - + -   1 2   A B C D E F G H RS1 RS2

ADD A JUMPER FROM TP1 TO SIG 1+ AND VERIFY SIG 1- IS GROUNDED

Some systems will contain the Single Channel 203 Board. Also, verify that SIG 1- is grounded. Add another jumper if needed.

SINGLE CHANNEL LAWSON 203 BOARD

TO SERIAL DATA PORT FOR CONNECTION TO YOUR COMPUTER

REM. ST.   GND   TEMP PROG 1 2   SIG 1+ -   GND

A B C D E F G H

ADD A JUMPER FROM TP1 TO SIG 1+ AND VERIFY SIG 1- IS GROUNDED

You could also run the TP1 wire to SIG 1+ through a relay for automatic hardware setup.
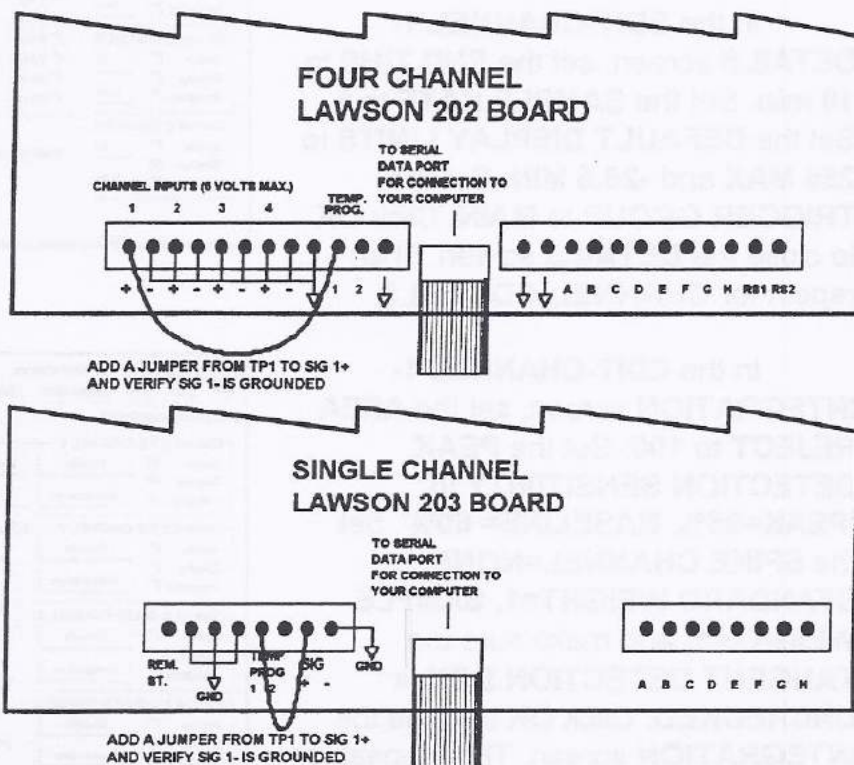
Loopback Test: (continued)

Setting Up The Software:

Re-attach the bottom cover and plug the G.C. back in. Turn the G.C. power on and start PeakSimple. Verify that the computer is communicating properly with the G.C..

In the **EDIT-OVERALL** screen, check the **LOOPBACK** box. Set the **START TIME** to 0 minutes and the **END TIME** to 10 minutes. Also verify that the **SHOW RETENTION WINDOWS** box is checked.

In the **EDIT-CHANNELS** screen, check the **ACTIVE, DISPLAY** and **INTEGRATE** boxes for channel 1. And check the **DISPLAY** and **INTEGRATE** boxes for channel 4.

In the **EDIT-CHANNEL 1-DETAILS** screen, set the **END TIME** to **10 min.** Set the **SAMPLE RATE** to **1.** Set the **DEFAULT DISPLAY LIMITS** to **256 MAX** and **-25.6 MIN.** Set the **TRIGGER GROUP** to **MAIN.** Click **OK** to close the **DETAILS** screen. Then repeat for **CHANNEL 4-DETAILS.**

In the **EDIT-CHANNEL 1-INTEGRATION** screen, set the **AREA REJECT** to **100.** Set the **PEAK DETECTION SENSITIVITY** to **'PEAK=95%, BASELINE= 60%'.** Set the **SPIKE CHANNEL=NONE, STANDARD WEIGHT=1, SAMPLE WEIGHT=1,** and make sure the **TANGENT DETECTION BOX** is **UNCHECKED.** Click **OK** to close the **INTEGRATION** screen. Then repeat for **CHANNEL 4-INTEGRATION.**
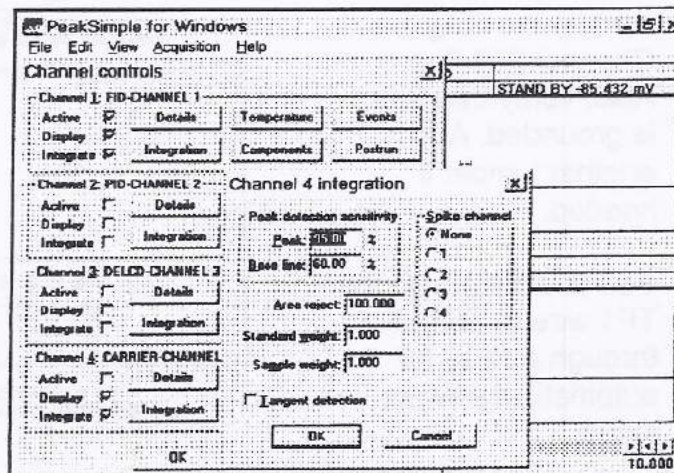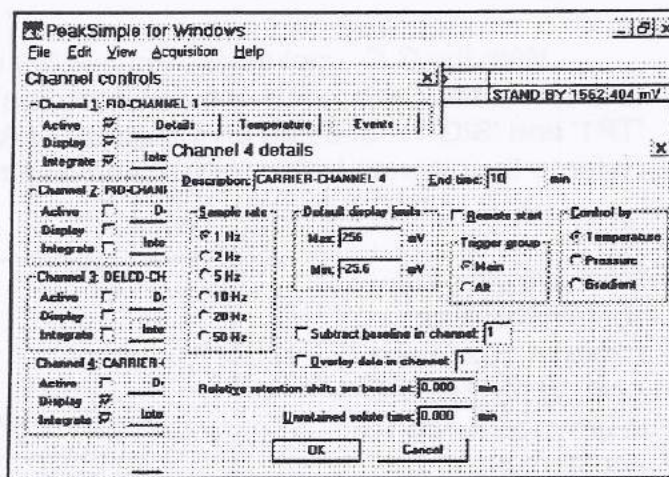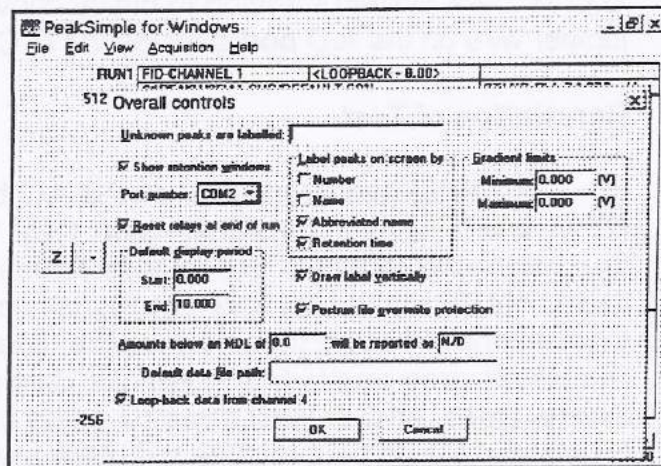
Loopback Test: (continued)

Software Setup: (continued)

In the **EDIT-CHANNEL 1-COMPONENTS-LOAD** screen, highlight the **602.cpt** sample components file and click **OPEN**. Click **OK** again to close the **COMPONENTS** screen. Then repeat for **CHANNEL 4-COMPONENTS**.

In the **FILE-OPEN** screen, select **CHANNEL 4** at the bottom of the window and then highlight the **602.chr** sample chromatogram file and select **OPEN**.

The 602.chr sample chromatogram that is now displayed on channel four represents the data that will be fed back through the A/D converter.

Starting the Run:

Auto-zero channel 1 by clicking the '**Z**' button. Depress the **SPACEBAR** and the chromatogram will start running. The data on **CHANNEL 1** should appear to be an exact replica of the data that was fed into **CHANNEL 4**.

Collecting the Data:

After the run has completed, make note of the area counts of one of the peaks by left-clicking on one of the peaks. Toluene, for example, may have an area count of 931.

Repeat the run three or more times; for each run, record the area counts of the same peak. Once the data has been collected from at least three runs; an average area count can be calculated as well as the percent relative standard deviation.
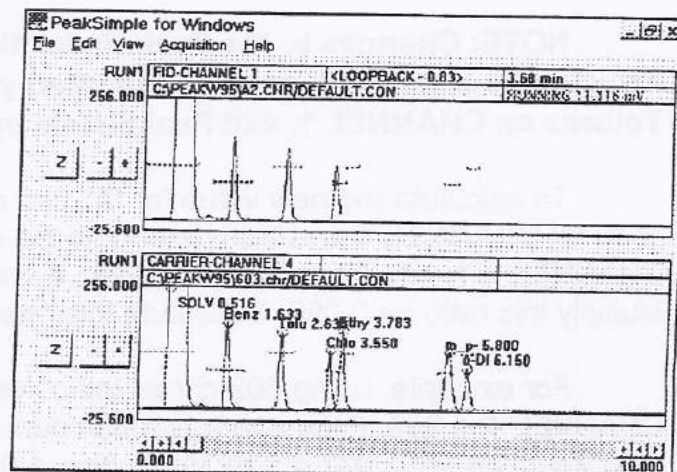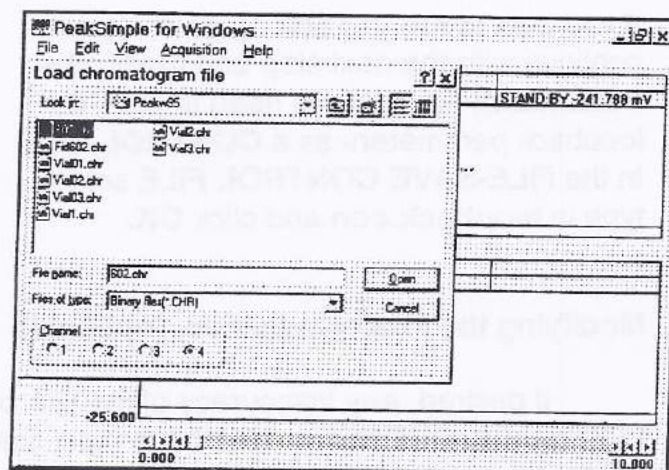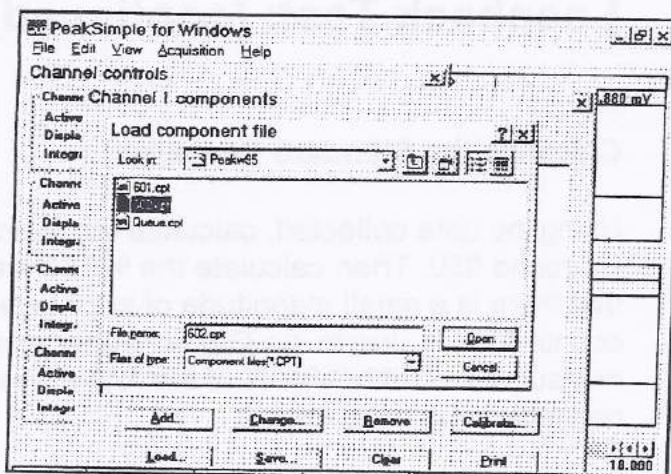
Loopback Test: (continued)

Calculate the Standard Deviation:

Using the data collected, calculate the average area counts for Toluene. Typically this value is around 950. Then calculate the %RSD which is usually less than 0.5%. You may notice that there is a small magnitude of error between the **CHANNEL 1** and **CHANNEL 4** area counts. This is due to the D/A converter and not the A/D converter. Since the loopback test measures the **PRECISION** of the A/D converter and not its **ACCURACY**, this minor discrepancy is insignificant.

Save Your Loopback Test as a CONTROL FILE:

If you wish to run this test again or if you continue with the next step and modify the **Peakwin.ini** file, you will need to save the loopback parameters as a **CONTROL FILE**. In the **FILE-SAVE CONTROL FILE** screen, type in **loopback.con** and click **OK**.
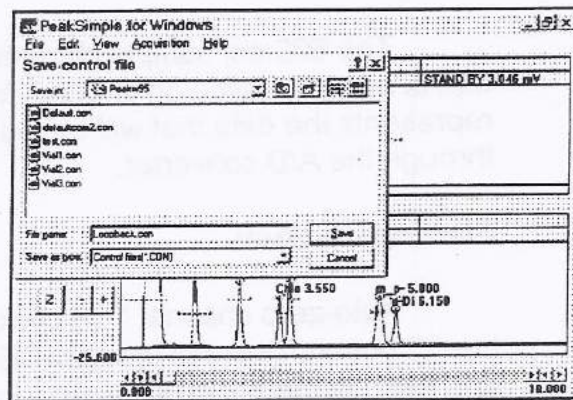
Modifying the Peakwin.ini File, (optional)

If desired, any inaccuracy of the D/A converter can be adjusted by attenuating the **LOOPBACK OUTPUT** to match the input signal. This adjustment can be made by entering the line **"LoopbackFactor=X"** in the [Lawson] section of the **PEAKWIN.INI** file located in the **WINDOWS** directory. The default value of '**X**' is **0.098**.

NOTE: Changes to the Peakwin.ini file will not be recognized unless the PeakSimple application is restarted. After you have obtained the average area count for Toluene on CHANNEL 1, exit PeakSimple by pressing 'Q', then 'Y'.
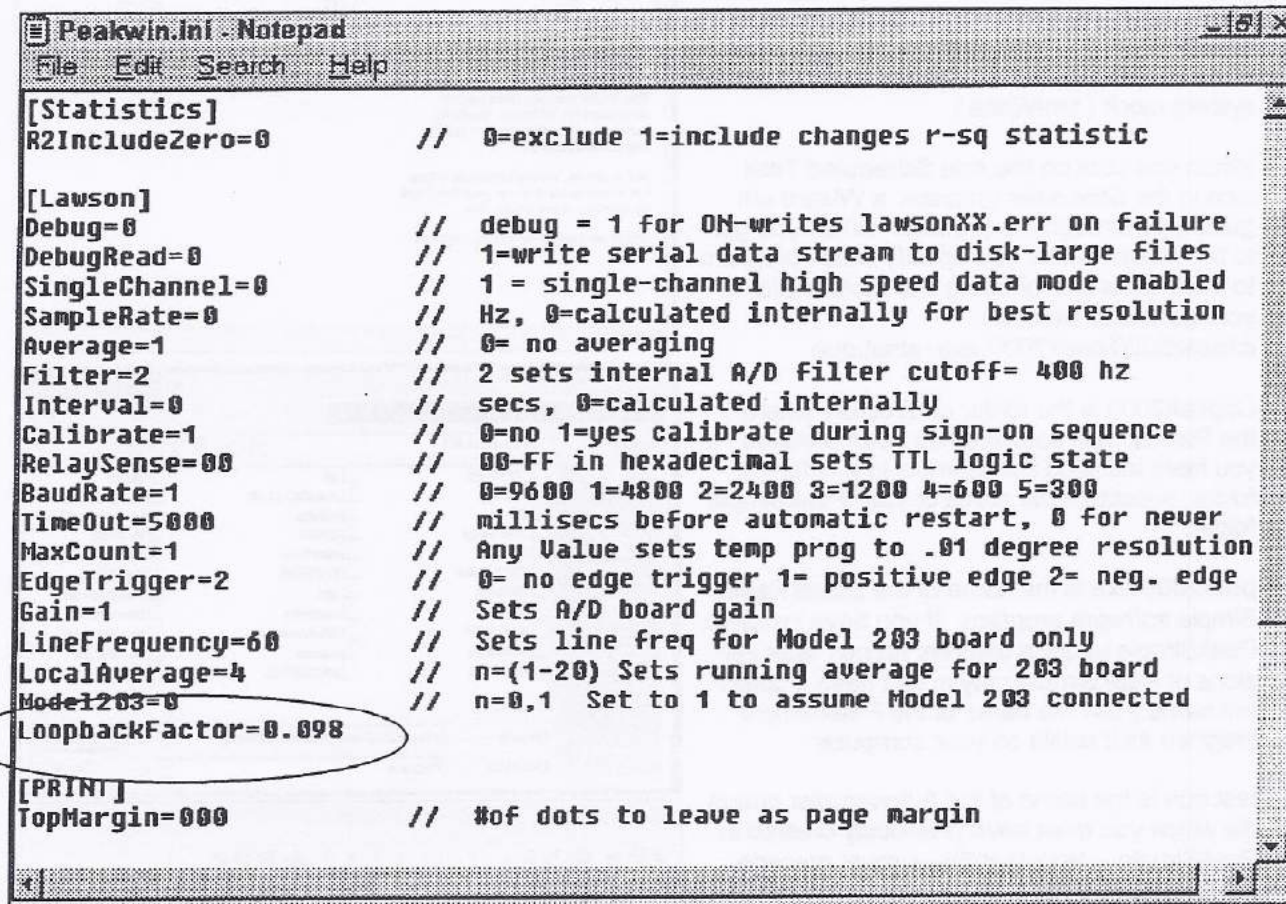
To calculate the new value for '**X**', first determine the average area counts of a specific peak for **CHANNEL 1** and also determine the area count of the corresponding peak on **CHANNEL 4**. Next, divide the **CHANNEL 4** area count by the **CHANNEL 1** area count. Multiply this ratio by 0.098. Substitute this new value for '**X**'.

For example, using 602.chr as the chromatogram file, the toluene area count for **CHANNEL 4** is **953**. If the average area count of toluene on **CHANNEL 1** is **931** then the ratio would be **953 / 931 = 1.0236**. Multiply **1.0236 x 0.098**. the answer is **0.1003128**. Round this new value for '**X**' to 0.1003.

Loopback Test: (continued)

Modifying the Peakwin.ini File, (continued)

Find the **PEAKWIN.INI** file in the **WINDOWS** sub-directory. Double-click to open it. Scroll down until you find the [Lawson] section. Place the cursor at the last line of the [Lawson] section and type **"LoopbackFactor=X"**, and then press **ENTER**. X is the value you calculated earlier. For example **"LoopbackFactor=0.1003"**.

```
Peakwin.ini - Notepad                                                    _ |@| X
File   Edit   Search   Help

[Statistics]
R2IncludeZero=0          //   0=exclude 1=include changes r-sq statistic

[Lawson]
Debug=0                  //   debug = 1 for ON-writes lawsonXX.err on failure
DebugRead=0              //   1=write serial data stream to disk-large files
SingleChannel=0          //   1 = single-channel high speed data mode enabled
SampleRate=0             //   Hz, 0=calculated internally for best resolution
Average=1                //   0= no averaging
Filter=2                 //   2 sets internal A/D filter cutoff= 400 hz
Interval=0               //   secs, 0=calculated internally
Calibrate=1              //   0=no 1=yes calibrate during sign-on sequence
RelaySense=00            //   00-FF in hexadecimal sets TTL logic state
BaudRate=1               //   0=9600 1=4800 2=2400 3=1200 4=600 5=300
TimeOut=5000             //   millisecs before automatic restart, 0 for never
MaxCount=1               //   Any Value sets temp prog to .01 degree resolution
EdgeTrigger=2            //   0= no edge trigger 1= positive edge 2= neg. edge
Gain=1                   //   Sets A/D board gain
LineFrequency=60         //   Sets line freq for Model 203 board only
LocalAverage=4           //   n=(1-20) Sets running average for 203 board
Model203=0               //   n=0,1  Set to 1 to assume Model 203 connected
LoopbackFactor=0.098

[PRINT]
TopMargin=000            //   #of dots to leave as page margin
```

Press **ALT-F** then **S** to save the file. Press **ALT-F** then **X** to exit. Restart **PeakSimple** and load the loopback.con control file you saved earlier. Run the loopback test again. The accuracy of the D/A converter should be improved. (Channel 1 toluene area counts should closely match the channel 4 toluene area counts).

End of Test:

Turn off G.C. power and re-connect the original A/D board wiring. The loopback test is completed.

The Windows Task Scheduler program is supplied with the Windows operating system. It is found under Programs/Accessories/System Tools. The Scheduler allows you to trigger a PeakSimple Autosampler Queue or a specific control file at a scheduled time and date, or on a regular repeating basis using the computer's system clock (time/date).

When you click on the Add Scheduled Task icon in the Scheduler program, a Wizard will guide you through the process. When you get to the screen where you specify which program to start, enter the following line modified for your particular situation:
c:\peak2000\peak2000.exe -atest.que

C:\peak2000 is the folder or directory where the PeakSimple software has been installed. If you have installed PeakSimple in a different folder, substitute the name of your PeakSimple folder.

peak2000.exe is the name of the actual PeakSimple software program. If you have installed PeakSimple under a different name (later versions of PeakSimple may in fact have a different name) use the name of the PeakSimple program as it exists on your computer.

test.que is the name of the Autosampler queue file which you must have previously created in PeakSimple. Note that the -a must precede the name of the .que file. When you create the .que file in PeakSimple you can save the que under any name you want. The -a is a Windows programming convention and must precede the name of the que file you want to run.

When the Scheduler starts PeakSimple, the specified queue will begin. At the end of the queue, PeakSimple will wait for the delay time specified in the queue, and then PeakSimple will Close automatically.
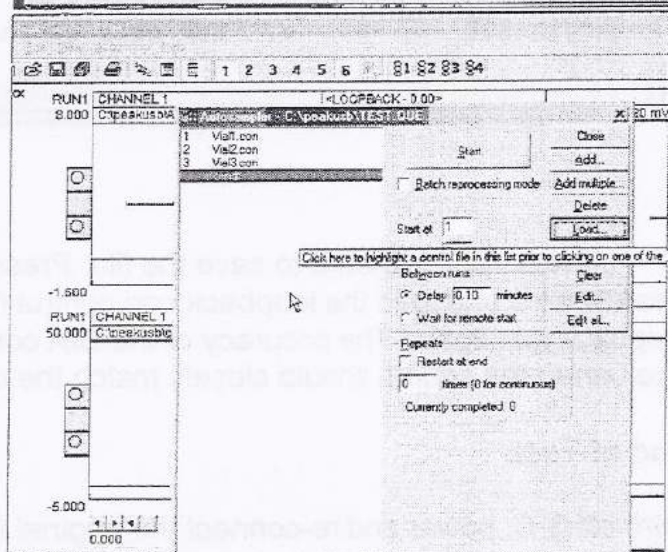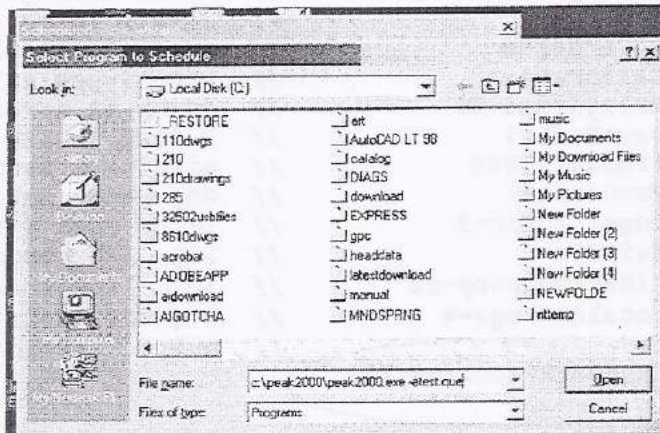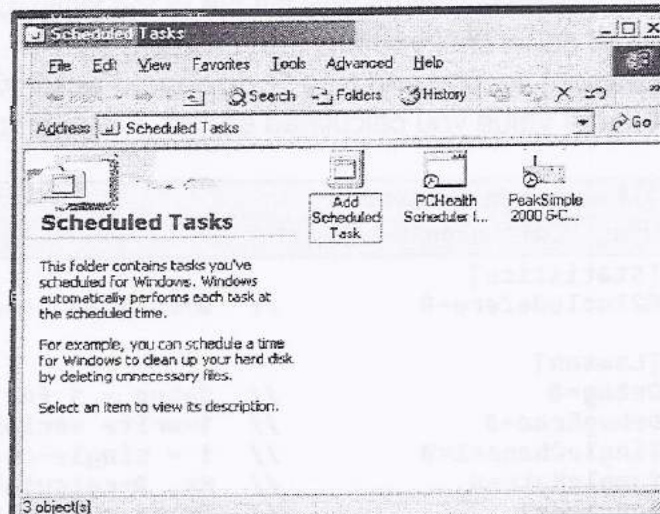
man264.pub

GPC From PeakSimple Data Acquisition

Introduction

The following is an outline of how PeakSimple data acquisition software/hardware can be used to acquire and analyze (in conjunction with an appropriate spreadsheet) gel permeation chromatography data. At this time, two different version of PeakSimple software were required for successful analysis. Version 2.08 was used to collect the data and obtain result tables for narrow polymer standard chromatograms, while version 2.09 was used to obtain the peak slice information for broad unknown polymers. That is, using 2.09, the voltage difference between the detector output and the subsequently drawn baseline was obtained for each data point and saved as an ASCII file, which was then imported into Excel for in-depth GPC analysis. Ultimately, it would be preferred to use only one version of PeakSimple. However, 2.09 (the latest version) was not stable while acquiring data. The program would crash after approximately 5 minutes. Furthermore, the time display in the upper right hand corner did not appear to work and retention windows were not visible on the screen although a component file was active. Thus, 2.09 was used only for obtaining slice information with non-active channels.

To illustrate how PeakSimple can be used for GPC analysis, I have included 3 narrow polystyrene standard chromatograms (4 standards per chromatogram) and two broad unknown polymer chromatograms. Chromatograms were obtained using a Waters 510 pump (U6K injector), an ethyl acetate mobile phase (1 mL/min), a series of Ultrastyrogel® columns (Waters 10^6, 10^4 and 500 Å) and Waters 2410 refractive index detector. All polymers were pre-dissolved in ethyl acetate and chromatograms were collected at 1 Hz. Polystyrene standard concentrations were 0.1 % w/w or less (50 µL injection volume) while broad unknown polymers were approximately 1 % w/w (75 µL injection volume). Also included are component files, containing the standard identities and expected retention windows, an event file for integration, and two ASCII data files containing slice information for the broad unknown polymers, and an Excel file with in-depth GPC analysis.

Obtaining a Calibration Curve

Polydisperse polymers in solution are fractionated according to size or hydrodynamic volume during GPC, which is also known as size exclusion chromatography. Molecular weight is related to the hydrodynamic volume. In GPC a dilute polymer solution is injected into a solvent stream which then flows through a series of columns packed with porous gel beads. Smaller molecules pass through and around the beads while larger molecules are excluded from all but the largest pores. Thus fractionation occurs with the largest molecules eluting first. The molecular weight of an eluting polymer molecule varies exponentially with eluting volume, the latter of which is proportional to time under constant flow rate conditions. To obtain molecular weight data and convert the GPC chromatogram into a molecular weight distribution, the relation between molecular weight and elution time is obtained from a series of polymer standards of known molecular weight. The calibration curve is thus obtained from a plot of the logarithm of molecular weight versus time. Given that GPC is a comparison of hydrodynamic volumes, unknown molecular weight determinations will be relative to the calibration standards. For a good introductory reference to polymer science, see R. J. Young and P. A. Lovell, Introduction to Polymers.

Using PeakSimple 2.08, the result table for each of the three polystyrene standard chromatograms was copied using DDE into Excel. The natural logarithm of molecular weight versus time was plotted and a best fit analytical approximation to the curve was obtained from a third order polynomial, $P(t_e)$. This is the calibration curve relating molecular weight to elution time.

Obtaining Molecular Weight Averages

The most common and convenient way to characterize a distribution of molecular weights making up a polymer sample is using molecular weight averages such as, number average molecular weight ($\overline{M}_n$), and weight average molecular weight ($\overline{M}_w$), as shown in the following figure for a typical polymer chromatogram. $\overline{M}_n$ is defined as a sum of products of the molecular weight of each fraction multiplied by its mole fraction. That is: $\overline{M}_n = \sum X_i M_i$ where X_i is the mole fraction of molecules of molecular weight mass M_i. The weight average molar mass is

defined as a sum of the products of the molecular weight of each fraction multiplied by its weight fraction, w_i. That is: $\overline{M}_w = \sum w_i M_i$. Additionally, it can be shown that the number average molecular weight, in terms of weight fraction, is equal to: $\overline{M}_n = 1/\sum(w_i/M_i)$. The ratio $\overline{M}_w/\overline{M}_n$ is known as the polydispersity or polydispersity index (PDI). The PDI is often used as a measure of the breadth of the molecular weight distribution. Polymers that are monodisperse (i.e. all chains have the same molecular weight) would have a PDI of 1.
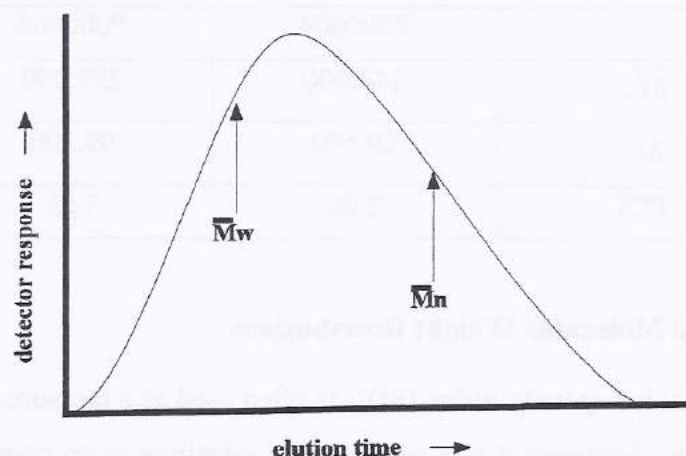
A typical polydisperse polymer molecular weight distribution showing the approximate locations of $\overline{M}_n$ and $\overline{M}_w$.

Using PeakSimple 2.09, polymers p000604 and p000606 were integrated (using the GPC event file) and the results saved in ASCII files. The ASCII files were imported into Excel and the corresponding sample times were added as a third column of data starting at time equal to zero. Only slice and time data corresponding to the major peak of interest were retained (columns A,B and J,K respectively). For each time slice, a corresponding molecular weight, M_i, was calculated using the analytical equation fitted to the calibration curve (columns C and L, respectively). Note that extrapolation of a few minutes outside of the last standard (MW = 1,000,000) is usually not a problem. Furthermore, the refractive index response of the detector is proportional to the weight concentration of eluting polymer, independent of molecular weight. Thus, the weight fraction, w_i, of polymer in any slice is equal to the detector voltage response or height (baseline subtracted) divided by the sum of detector voltage responses for each polymer

elution slice (i.e. $w_i = \text{height}_i / \sum \text{height}_i$, columns D and M respectively). $\overline{M}_w$ was obtained by multiplying w_i and M_i and summing the appropriate columns (see bottom of columns E and N). $1/\overline{M}_n$ was obtained by dividing each w_i by M_i and summing the appropriate columns (see bottom of columns F and O). Thus, the molecular weight averages for the two polymers were obtained and are summarized in the following table.

Polymer Molecular Weight Averages

	P000604	P000606
$\overline{M}_w$	143,000	299,000
$\overline{M}_n$	69,500	99,300
PDI	2.06	3.01

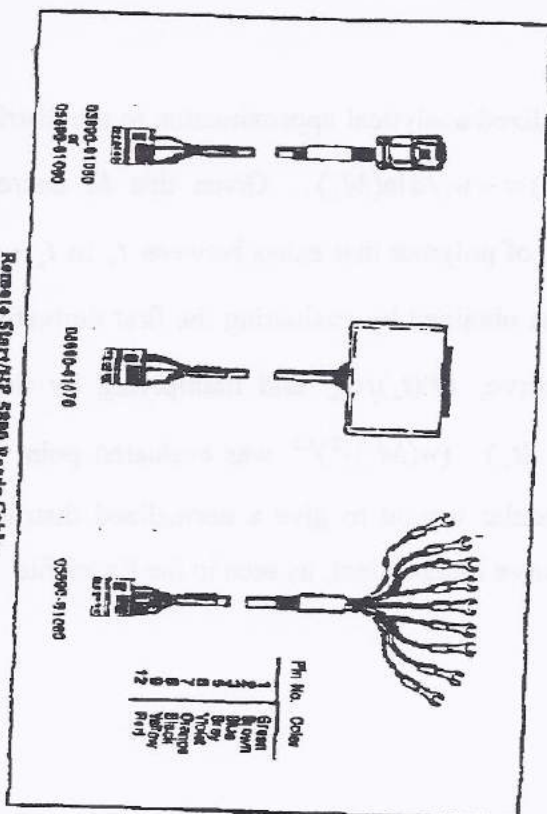## Obtaining Normalized Molecular Weight Distributions

As mentioned the polydispersity index (PDI) is often used as a measure of the breadth of the molar mass distribution. However it is a often a poor substitute when compared to a graphical representation of the complete molecular weight distribution curve, especially when comparing polymer distributions. To a first approximation, the raw chromatogram (a graph of detector response, $f(t_e)$, versus elution time, t_e) is a graphical representation of the distribution. However, the chromatogram height is injection concentration dependent, making comparisons difficult, and t_e is often non-linear with $\ln(M)$, as evidenced by a third order calibration curve.

A normalized molecular weight distribution function is given by $w(M) = -dw/d\ln(M)$. Conversion of $f(t_e)$ versus t_e to a normalized molecular weight distribution plot (i.e. $w(M)$ versus M or $\ln(M)$), is obtained by considering that the weight fraction, dw, of polymer which elutes between t_e and $t_e + dt_e$ is given by: $dw = f(t_e)dt_e \Big/ \int_0^\infty f(t_e)dt_e$ where the integral in the denominator is simply the area under the chromatogram. Thus, an analytical approximation of dw at the ith slice is w_i, the weight fraction of polymer

A normalized analytical approximation to the distribution function, $w(M_i)$, is thus obtained from: $w(M_i) = -w_i/d\ln(M_i)$. Given that M decreases as t_e increases, the same weight fraction, dw, of polymer that exists between t_e to $t_e + dt_e$ also exists between $\ln(M) - d\ln(M)$. $d\ln(M_i)$ was obtained by evaluating the first derivative of the analytical equation fitted to the calibration curve, $dP(t_e)/dt_e$ and multiplying by the time interval (i.e. the 1 Hz sampling frequency ~ dt_e). $(w(M_i))^2)^{1/2}$ was evaluated point by point (columns H and Q) and plotted against molecular weight to give a normalized distribution that is injection concentration and calibration curve independent, as seen in the Excel file.

The REMOTE receptacle provides a function used primarily to start an integrator or data system when an HP 5890 run begins, and also provides ready information.
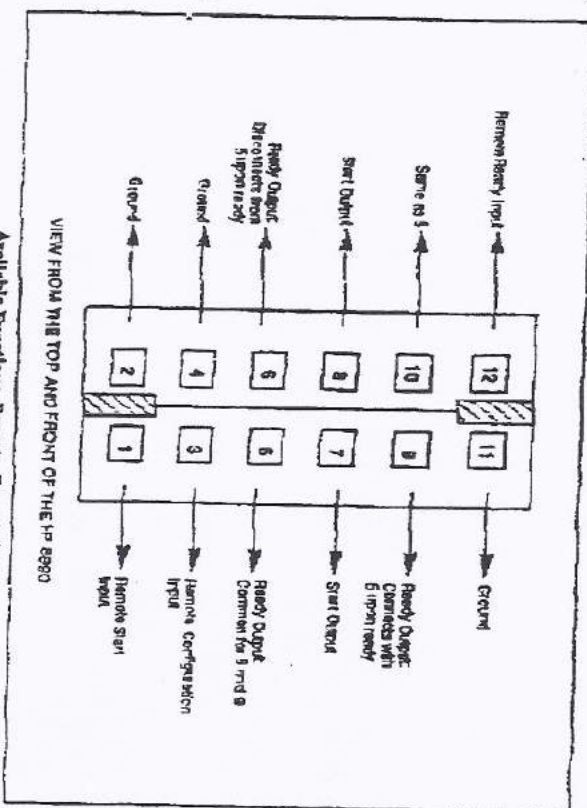
Remote Start/HP 5890 Ready Cables

Pin No.	Color
1	Green
2	Brown
3	Blue
4	Gray
5	Violet
6	Orange
7	Black
8	Yellow
9	Red
12	

Available Remote Start/HP 5890 Ready Cables

Part No.	Use	Length
D3384-60560	HP 3384/96A Integrator/Controller	2 m
05880-81080	HP 3392A Integrator/Controller	2 m
05890-81050	HP 3390A Integrator	2 m
05880-81070	HP 3350 Series LAB	2 m
05890-81080	General Purpose	2 m

Remote Receptacle

The 12-pin REMOTE receptacle provides a variety of functions, depending upon connection made via the cable. The figure below and the table following identify the function at each pin.

Top row (left to right): Remote Ready Input (12), Same as 9 (10), Start Output (8), Ready Output Differentiate from 5 upon ready (6), Ground (4), Ground (2)

Bottom row (left to right): Ground (11), Ready Output Connects with 6 upon ready (9), Start Output (7), Ready Output Common for 5 and 9 (5), Remote Configuration Input (3), Remote Start Input (1)

VIEW FROM THE TOP AND FRONT OF THE HP 5890

Available Functions, Remote Receptacle J2

Analog Signal Output Cables

The following figure and table show cables available to connect an HP 5890 ANALOG output channel (variable DC signals, +1 V or +1 mV maximum) to a recorder, integrator, and/or A/D converter for a computer system. If a second output channel is installed, a second cable is also required.

White + (High)
Black − (Low)
Orange Ground

05890-60760   05890-60780   05890-60780   05890-60800
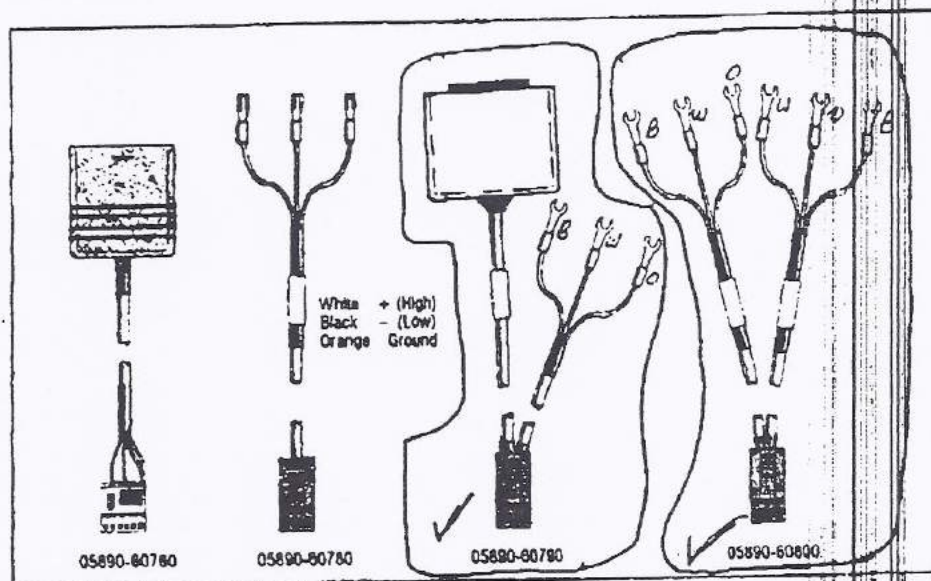
Available Analog Signal Output Cables

Note that the general purpose and HP 3350 Series LAS analog output cable assemblies consist of two independent cables, terminating together at a common, single, female plug at the HP 5890. One cable is labelled 1_mV, the other +1 V output. In general, the +1 V cable is connected to an integrating device or A/D converter, and the 1 mV cable is connected to a chart recording device.

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Installation 2-20

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03/25/1999